Adelina Maria Gaspar Gama Quaresma

Canine mammary tumours: new insights into prognosis and molecular classification

Orientador científico: Professor Doutor Fernando Carlos de Lander Schmitt
Faculdade de Medicina da Universidade do Porto
Instituto de Patologia e Imunologia Molecular da Universidade do Porto (IPATIMUP)

Co-orientador científico: Professora Doutora Anabela Gouveia Antunes Alves Universidade de Trás-os-Montes e Alto Douro



Departamento de Ciências Veterinárias Universidade de Trás-os-Montes e Alto Douro Vila Real, 2008

Tese de candidatura ao grau de doutor, de acordo com o disposto no nº1 do artigo 17º do Decreto-Lei nº216/92 de 13 de Outubro.

This study was supported by Fundação para a Ciência e Tecnologia, research project POCTI/CVT/57795/2004 and by the Centro de Ciência Animal e Veterinária (CECAV), Universidade de Trás-os-Montes e Alto Douro.
${f v}$

Abstract

In canine species, spontaneous mammary tumours constitute the second most frequent neoplasia, surpassed only by skin tumours. When considering female dogs, mammary tumours represent the most common neoplasia, with malignant tumours accounting for up to 50% of cases. These facts have raised an increasing interest on the research of reliable prognostic factors in canine mammary tumours, and similarly to humans, the veterinary pathologist might assume a fundamental role by providing both histological diagnosis as well as additional information regarding the prognosis of a particular animal.

At present, and despite several prognostic studies in this area, results are not consensual, which is mandatory for the validation of classical clinicopathological parameters and the search of novel prognostic factors. Therefore, the central goal of our thesis was the research of clinicopathological and molecular factors with potential impact on the prognosis of canine mammary tumours.

The present thesis is composed by seven chapters: an initial chapter (Chapter I) corresponding to the state of the art; Chapters II-VI, which correspond to scientific articles resulting from our investigation; and Chapter VII, which promotes a global and final discussion of the results.

Chapter I (General Introduction) is a review of the most recent literature concerning canine mammary tumours, especially with regard to prognostic studies. A particular emphasis is given to several molecular cell markers, in view of both canine and human scientific literature. At the end of this chapter, we have delineated the main goals of the present thesis.

In Chapter II (Canine mammary gland tumours: clinical and pathological parameters as predictors of overall and disease-free survival - a univariate and multivariate analysis), we have performed a clinical and histopathological characterization of a hundred and fifty six canine mammary tumour specimens (46 benign and 110 malignant). In order to investigate the prognostic value of clinical and pathological variables, a follow-up study was performed in 69 female dogs for a minimum period of 12 months after surgical procedure. Univariate analysis showed that tumour size, histological type, tumour growth, differentiation grade, stromal and lymphovascular invasion, lymph node status, mitotic and Ki-67 labelling indices were significantly associated with overall and

disease-free survival. Skin ulceration was only associated with poorer overall survival rates. Cox regression multivariate analysis revealed lymph node status as the only independent prognostic factor.

Chapter III (Expression of E-cadherin, P-cadherin and β-catenin in canine malignant mammary tumours in relation to clinicopathological parameters, proliferation and survival) describes the immunohistochemical evaluation of several adhesion molecules on a series of 65 canine malignant mammary tumours. Given the critical role assigned to cadherin-mediated cell adhesion during embryogenesis and in the maintenance of normal adult tissue architecture, as well as its putative involvement in tumour cell invasion and progression, we sought to investigate their expression in canine malignant mammary tumours and their association with clinicopathological variables, proliferation and survival.

Reduction in E-cadherin expression was significantly associated with increased tumour size, high histological and invasion grades, lymph node metastasis and high mitotic index, whereas reduced β -catenin expression was associated with high histological and invasion grades. P-cadherin expression was only associated with invasion. In 39 cases for which follow-up data was available, reduced E-cadherin and β -catenin expression was significantly associated with shorter overall survival and disease-free survival. Although this study has been performed with a relatively small number of cases, we have observed that an abnormal expression of adhesion molecules is a common phenomenon in canine mammary malignant tumours and, therefore, may play a central role in tumour progression. Further studies with a larger series will certainly highlight the prognostic value of these molecules in the context of canine mammary tumours.

In Chapter IV (Immunohistochemical expression of Epidermal Growth Factor Receptor (EGFR) in canine mammary tissues), an evaluation of EGFR immunohistochemical expression was performed in a series of 136 canine mammary tumours (46 benign and 90 malignant) and representative areas of adjacent normal and hyperplastic mammary tissue. Despite the availability of several biochemical studies of EGFR in canine mammary tumours, there are still no immunohistochemical studies concerning its expression, which directed us to its evaluation both in benign and malignant tumours. Immunohistochemistry has the advantage of disclosing the precise cellular location of a particular protein, which is not possible by using immunoenzymatic methodologies.

In normal and hyperplastic canine mammary glands, EGFR expression was consistently observed in myoepithelial cells, with luminal cells usually negative. Perilobular stroma was commonly positive. In benign tumours, EGFR was present in both epithelial cell components, but luminal cells were weakly positive, when compared to malignant tumours. In fact, EGFR overexpression was found in 9 benign (19.6%) and 38 malignant (42.2%) lesions, with EGFR positivity significantly related with malignancy. Besides animal age and tumour size, there were no significant associations between other clinicopathological parameters and EGFR overexpression. On survival analysis, tumours with EGFR overexpression showed a reduced disease-free and overall survival; however, these associations failed to reach statistically significant levels. Further studies are warranted, namely concerning the analysis of EGFR gene amplification, given that EGFR might represent a potential therapeutic target.

Chapter V (Expression and prognostic significance of cytokeratin (CK) 19 in canine malignant mammary tumours) describes the immunohistochemical evaluation of CK19 in a series of 102 malignant canine mammary tumours and investigates the possible association between CK19 pattern of expression and clinicopathological parameters, proliferation and survival. This study was based on recent evidence demonstrating a significant association between the reduction of luminal CK (such as CK19) and a more aggressive behaviour of human breast cancer, usually associated with a basal phenotype. Therefore, besides the evaluation of the prognostic potential of this luminal cell marker, we have also investigated its association with a basal/myoepithelial phenotype, by using additional specific cell differentiation markers.

Reduced/absent CK19 was significantly associated with histological type, invasiveness, high histological grade and an elevated Ki-67 index. CK19 positive expression was significantly associated with the presence of ER, whereas its reduced immunostaining was associated with basal/myoepithelial cell markers positive expression. Survival analysis demonstrated that down-regulation of this luminal CK is significantly associated with shorter overall and disease-free survival rates; however, CK19 was not an independent prognostic factor in multivariate analysis. In our series, CK19 down-regulation was significantly related to an aggressive phenotype; yet, the real implication of this phenomenon is not known, namely during tumour progression.

Chapter VI (Identification of molecular phenotypes in canine mammary carcinomas with clinical implications: application of the human classification) illustrates the application of a recently described classification for human breast carcinomas to a series

of 102 canine mammary carcinomas. This classification was initially based on gene expression profiling analysis and it was later on reinforced at the protein level, by using immunohistochemistry; both methodologies identified distinct phenotypes of human breast cancer associated with distinct clinical behaviours. Similarly to human studies, by using an immunohistochemistry surrogate panel based on five molecular markers (estrogen receptor, HER-2, cytokeratin 5, p63 and P-cadherin), we were able to classify canine mammary carcinomas into four different subtypes: luminal A (ER+/HER-2-), luminal B (ER+/HER-2+), basal (ER-/HER-2- and a basal marker positive) and HER-2 overexpressing tumours (ER-/HER-2+).

Luminal A-type tumours were characterized by lower grade and proliferation rate, whereas basal-type tumours were mostly high grade, high proliferative and positive for CK5, p63 and P-cadherin. In addition, as in humans, basal subtype was significantly associated with shorter disease-free and overall survival rates.

Although we consider these findings as preliminary results, which require further validation, this study pointed out to similar phenotypes to the ones described in the human literature. So, canine mammary carcinomas might represent a suitable natural model for the study of human breast carcinomas, in particular to the basal subset, given the putative high percentage of basal carcinomas identified in the dog.

The final Chapter (Chapter VII – General discussion and concluding remarks) encloses a global discussion of our investigation, stressing the most relevant and significant findings.

Resumo

Na espécie canina, os tumores mamários espontâneos representam a segunda neoplasia mais comum, sendo apenas ultrapassados pelos tumores de pele. Considerando os indivíduos do sexo feminino, os tumores de mama constituem a neoplasia espontânea mais frequente, representando os tumores malignos cerca de 50% dos casos observados. Estes factos suscitam um interesse crescente na pesquisa de factores de prognóstico credíveis na área dos tumores mamários caninos e à semelhança do que ocorre em Medicina Humana, o patologista veterinário pode assumir um papel fundamental ao fornecer não apenas um diagnóstico histológico, como também informação adicional acerca do prognóstico de um determinado indivíduo.

Actualmente, e apesar de vários estudos de prognóstico nesta área, os resultados não são consensuais pelo que se torna necessária a validação dos parâmetros clínico-patológicos considerados clássicos e a pesquisa de novos factores com valor prognóstico. Assim, tendo como objectivo central a pesquisa de factores com possível impacto no prognóstico dos tumores de mama de cadela, procedemos ao estudo de diversas características clínico-patológicas e moleculares, que se encontram discriminadas ao longo deste trabalho.

A presente dissertação é constituída por sete capítulos: um capítulo inicial de revisão bibliográfica (Capítulo I); os Capítulos II a VI, que correspondem aos artigos científicos resultantes da investigação desenvolvida; e o Capítulo VII, onde se promove uma discussão geral do trabalho efectuado.

O Capítulo I (Introdução Geral) consiste numa revisão bibliográfica actualizada acerca dos tumores de mama de cadela, em especial no que diz respeito a estudos de prognóstico. É ainda dado ênfase particular a alguns marcadores moleculares utilizados ao longo do nosso trabalho, tendo em consideração estudos efectuados em tumores mamários caninos e humanos. No fim deste capítulo, são enumerados os objectivos da presente dissertação.

Ao longo do Capítulo II (Tumores mamários caninos: parâmetros clínico-patológicos como factores preditivos da sobrevivência total e sobrevivência livre de doença – análise uni- e multivariada) procedeu-se à caracterização clínica e histopatológica de uma série de 156 tumores de mama de cadela (46 benignos e 110 malignos). Com o objectivo de investigar o valor prognóstico de variáveis clínico-patológicas, foi

efectuado um estudo de sobrevivência após exérese cirúrgica em 69 animais, durante um período mínimo de 12 meses. A análise univariada revelou que o tamanho do tumor, o tipo histológico, o modo de crescimento, o grau histológico, a invasão estromal e linfo-vascular, a presença de metástases ganglionares, e os índices de proliferação se encontravam significativamente associados com as sobrevivências total e livre de doença. A presença de ulceração cutânea encontrou-se associada apenas com a sobrevida total. A análise multivariada revelou a presença de metástases ganglionares como o único factor de prognóstico independente.

No Capítulo III (Expressão da caderina E, caderina P e β-catenina em tumores mamários caninos malignos em relação a parâmetros clínico-patológicos, proliferação e sobrevivência) efectuou-se a avaliação imunohistoquímica de moléculas de adesão numa série de 65 tumores mamários malignos de cadela. Tendo em conta vários estudos que demonstram a função importante da adesão mediada por caderinas durante os processos de desenvolvimento e na manutenção da arquitectura dos tecidos adultos, bem como o seu envolvimento durante a invasão e progressão tumoral, investigámos a expressão das moléculas acima descritas em tumores mamários malignos de cadela e a sua possível associação com parâmetros clínico-patológicos clássicos, índices de proliferação e sobrevivência.

Observámos que a redução da expressão da caderina E esteve significativamente associada com o tamanho do tumor, alto grau histológico, invasão, presença de metástases ganglionares e elevado índice mitótico; por outro lado, a redução da expressão da β-catenina encontrou-se significativamente associada com alto grau histológico e invasão. Relativamente à caderina P, a sua expressão encontrou-se significativamente associada apenas com a invasão. No que diz respeito ao estudo de sobrevivência, a redução da caderina E e β-catenina encontrou-se significativamente associada com menor tempo de sobrevivência total e livre de doença. Apesar deste estudo ter sido efectuado com um número reduzido de amostras, observou-se que a expressão alterada do complexo caderina-catenina é um evento comum nestas neoplasias. A realização de novos estudos com maior número de casos irá certamente esclarecer o valor prognóstico destas moléculas no contexto dos tumores mamários caninos.

No Capítulo IV (Expressão imunohistoquímica do Receptor para o Factor de Crescimento Epidérmico (*EGFR*) em tecidos mamários caninos), descreveu-se a

avaliação do EGFR através da técnica de imunohistoquímica numa série de 136 tumores mamários caninos (46 benignos e 90 malignos). Avaliou-se ainda a sua expressão na glândula mamária normal e hiperplásica adjacente. Apesar da existência de vários trabalhos em tumores mamários caninos com recurso a métodos imunoenzimáticos para a avaliação do EGFR, não existem ainda estudos de imunohistoquímica, pelo que considerámos importante avaliar a sua expressão em tumores benignos e malignos, nomeadamente a sua localização celular, informação que não é disponibilizada recorrendo às metodologias previamente descritas na literatura.

Na glândula mamária canina normal e hiperplásica, a expressão do EGFR foi observada principalmente ao nível das células mioepiteliais. No entanto, detectou-se positividade para este receptor em algumas células epiteliais luminais ductais, assim como no estroma perilobular. Relativamente aos tumores benignos, o EGFR foi observado no componente epitelial e mioepitelial, apresentando as células epiteliais um nível de expressão reduzido, quando comparado com os tumores malignos. De facto, a expressão de EGFR encontrou-se significativamente associada com a malignidade tumoral, tendo sido detectada uma imunoexpressão membranar completa de EGFR em mais de 10% das células neoplásicas em 42.2% de tumores malignos, versus 19.6% tumores benignos. Não se observou qualquer associação entre a expressão neoplásica do EGFR e os parâmetros clínico-patológicos, à excepção da idade e do tamanho do tumor. Apesar da sobre-expressão do EGFR mostrar uma tendência para um pior prognóstico, não foram encontradas associações estatisticamente significativas neste estudo. Acreditamos serem necessários estudos futuros acerca deste receptor, nomeadamente analisando a presença de amplificação do gene EGFR, já que este receptor pode constituir um potencial alvo terapêutico.

No Capítulo V (Expressão e valor prognóstico da citoqueratina (CK) 19 em tumores mamários malignos da cadela) procedeu-se à avaliação imunohistoquímica da CK19 numa série de 102 tumores mamários malignos de cadela, analisando-se a possível associação entre o seu padrão de expressão e parâmetros clínico-patológicos, proliferação e tempos de sobrevivência. À luz de estudos recentes em carcinomas humanos que demonstram uma associação entre a redução da expressão de CK luminais e uma maior agressividade biológica, julgámos pertinente investigar o padrão de expressão da CK19 (CK luminal) nos tumores mamários malignos da cadela, nomeadamente o seu potencial valor prognóstico e também a sua possível associação a

um fenótipo basal/mioepitelial, para tal utilizando marcadores específicos de diferenciação celular.

Neste trabalho, observámos que a redução ou ausência da CK19 se encontrou significativamente associada com o tipo histológico, invasão, alto grau histológico e índice Ki-67 elevado. A expressão da CK19 encontrou-se significativamente associada com a presença de receptores de estrogénio (ER), enquanto a sua redução se revelou associada com a presença de marcadores basais/mioepiteliais. Relativamente ao estudo de sobrevivência, a redução ou ausência da expressão deste marcador luminal provou estar associada a menores tempos de sobrevivência; no entanto, a CK19 não foi considerada como factor de prognóstico independente em análise multivariada. Apesar de, neste estudo, a ausência ou redução da expressão da CK19 se encontrar associada a um fenótipo tumoral mais agressivo, o significado biológico deste achado relativamente à progressão neoplásica permanece por esclarecer.

O Capítulo VI (Identificação de fenótipos moleculares em carcinomas mamários caninos com implicação clínica: aplicação de uma classificação humana) reflecte a tentativa de aplicação de uma classificação recentemente descrita para os carcinomas mamários humanos a uma série de 102 carcinomas mamários caninos. Esta classificação teve como base estudos de expressão genética e foi posteriormente comprovada através da técnica de imunohistoquímica, distinguindo diferentes fenótipos moleculares de cancro de mama humano. Recorrendo a marcadores válidos em Medicina Humana para a sua identificação (ER, HER-2, CK5, p63 and caderina P), classificámos os carcinomas mamários caninos em 4 subtipos principais: luminal A (ER+, HER-2-), luminal B (ER+, HER-2+), basal (ER-, HER-2- e um marcador basal positivo) e HER-2 (ER-, HER-2+). À semelhança da mulher, os tumores classificados como luminal A apresentaram baixo grau histológico e menores índices de proliferação, enquanto os carcinomas "basais" se caracterizaram geralmente por alto grau histológico e elevados índices de proliferação. Quanto à sobrevivência, também nos carcinomas de mama de cadela observámos uma associação entre o fenótipo basal e tempos de sobrevivência menores. Estes resultados parecem apontar para a existência de fenótipos moleculares semelhantes aos descritos em Medicina Humana, sugerindo o carcinoma de mama da cadela como um potencial modelo natural de estudo para o carcinoma de mama da mulher.

No Capítulo VII (Discussão geral e conclusões) procede-se à discussão global do trabalho desenvolvido, evidenciando-se os seus aspectos mais relevantes.

Ao meu Pai (*in memoriam*) À minha Mãe Ao Miguel, Jorge e Inês

Acknowledgements • Agradecimentos

Ao longo deste percurso, foram muitas as pessoas que prestaram o seu contributo e apoio. A elas a expressão do nosso agradecimento sincero.

À Universidade de Trás-os-Montes e Alto Douro, na pessoa do seu Magnífico Reitor, Professor Doutor Armando Mascarenhas Ferreira, manifestamos o nosso reconhecimento por ter autorizado a realização deste trabalho e pela disponibilidade dos meios para a sua execução prática.

Ao Senhor Professor Doutor Fernando Schmitt, a nossa especial gratidão por ter aceitado o encargo da orientação científica da nossa dissertação. A sua visão dinâmica, o seu rigor científico e o seu entusiasmo são qualidades que não esqueceremos. Muito obrigada por estar sempre presente e pelo apoio incondicional. Permaneceremos sempre devedoras do seu saber, confiança e amizade.

À Senhora Professora Doutora Anabela Alves, co-orientadora científica deste trabalho, expressamos o nosso profundo reconhecimento pela sua presença e amizade constantes, pelos conhecimentos transmitidos e pela disponibilização de meios para a execução da componente prática.

À Senhora Professora Doutora Fátima Gärtner, pela amizade e simpatia, pela disposição em colaborar e pelo precioso apoio na realização deste trabalho.

À Senhora Professora Doutora Conceição Martins, actual Coordenadora do Departamento de Ciências Veterinárias da UTAD, e ao Senhor Professor Doutor Jorge Rodrigues, à época Coordenador do Departamento de Ciências Veterinárias da UTAD, pelas facilidades concedidas para a realização deste trabalho.

À Senhora Professora Doutora Maria dos Anjos Pires, responsável pelo Laboratório de Histologia e Anatomia Patológica da UTAD, pela amizade, apoio e incentivo durante a realização deste estudo.

À Senhora Professora Doutora Paula Rodrigues, pela sua simpatia e disponibilidade em colaborar ao longo deste trabalho.

Às Senhoras Professoras Doutoras Isabel Pires e Justina Oliveira, nossas colegas de disciplina, pela simpatia e interesse demonstrados ao longo do nosso percurso.

Às funcionárias do Laboratório de Histologia e Anatomia Patológica da UTAD, Senhora D. Lígia Lourenço, Senhora D. Ana Plácido e Senhora D. Glória Milagres, pela simpatia, amizade e colaboração prestada. Agradecemos de modo especial à Senhora D. Lígia, pelas horas infindáveis ao micrótomo.

À Senhora D. Alda Magalhães, funcionária do Hospital Veterinário da UTAD, pela simpatia e disponibilidade sempre demonstrada.

A todos os elementos do Grupo de Patologia Mamária do IPATIMUP, pela simpatia e interesse demonstrados e pelas reuniões semanais sempre inovadoras no âmbito do cancro de mama. Um agradecimento especial à Senhora Professora Doutora Joana Paredes, pela sua amizade, partilha de conhecimentos e pela disponibilidade constante em colaborar.

Às técnicas do Laboratório de Anatomia Patológica do IPATIMUP, agradecemos a simpatia com que sempre nos acolheram. À Senhora Dra. Sara Ricardo, em particular, agradecemos a disponibilidade e ajuda na realização da técnica de imunohistoquímica.

À Reprografía da UTAD, na pessoa do Senhor Francisco Ribeiro, pelo profissionalismo durante a impressão e encadernação da presente dissertação.

Aos colegas das várias Clínicas e Hospitais Veterinários e aos proprietários dos animais que colaboraram neste estudo, o nosso muito obrigado.

Às Senhoras Professoras Doutoras Ana Margarida Calado e Paula Oliveira, pela simpatia e amizade demonstradas ao longo destes anos.

À Nanda, pela amizade com que sempre acompanhou o nosso trajecto, pelo seu apoio e encorajamento constantes e pelo "eterno" debate científico em torno dos tumores de mama.

À Milú, pela amizade com que acompanhou o nosso trabalho, pelo encorajamento e apoio que sempre manifestou.

À nossa família e amigos, pela paciência, amor e carinho que nunca nos negaram, mesmo quando a distância nos separava.

Ao Miguel, Jorge e Inês, que são a minha vida, pelo vosso amor incondicional.

Contents

Chapter I General Introduction	
1. Canine mammary gland tumours	3
General considerations	3
Etiopathogenesis	3
Pathology and natural behaviour	5
Histogenesis and differentiation	6
Prognosis	8
2. Differentiation and prognostic markers in mammary cancer: from dogs and humans	11
Differentiation cell markers in mammary cancer	11
Cytokeratins	12
Placental cadherin	13
P63	14
Prognostic markers in mammary cancer	15
Tumour cell proliferation	16
Hormone receptors	16
Human Epidermal Growth Factor Receptor-2 (HER-2)	18
Epidermal Growth Factor Receptor (EGFR)	20
Epithelial Cadherin	22
References	25
3. Aims and outline of the thesis	47
Chapter II Canine mammary gland tumours: clinical and pathological parameters as	
predictors of overall and disease-free survival - a univariate and multivariate analysis	49
Chapter III Expression of E-cadherin, P-cadherin and β-catenin in canine malignant	
mammary tumours in relation to clinicopathological parameters, proliferation and survival	89
Chapter IV Immunohistochemical expression of Epidermal Growth Factor Receptor	
(EGFR) in canine mammary tissues	105
Chapter V Expression and prognostic significance of CK19 in canine malignant	
mammary tumours	125
Chapter VI Identification of molecular phenotypes in canine mammary carcinomas with	
clinical implications: application of the human classification	147
Chapter VII General discussion and concluding remarks	169

List of Figures*

Fig. 1. Normal canine mammary gland: schematic representation (A) and	
immunohistochemical reactivity to basal/myoepithelial cell markers (B).	11
Fig. 2. Simplified overview of intracellular estrogen action mechanisms.	17
Fig. 3. Signalling pathways of HER family members.	19
Fig. 4. Schematic representation of the classical cadherin-catenin complex.	22

List of Tables*

Table 1. Histological classification of canine mammary tumours

*Tables and figures included in the papers are not listed here.

5

List of Abbreviations and Symbols

AgNORs Silver-stained nucleolar organizer regions

BRCA Breast Cancer

Ca Calcium cat catenin

CD Cluster of differentiation

CECAV Centro de Ciência Animal e Veterinária

CI Confidence interval

CK Cytokeratin
Cm centimetre

DAB 3,3-diaminobenzidine tetrahydrochloride

DFS Disease-free survival
DNA Deoxyribonucleic Acid
E-cadherin Epithelial cadherin

EGF Epidermal Growth Factor

EGFR Epidermal Growth Factor Receptor

ER Oestrogen Receptor

ERE Oestrogen responsive element

ERK Extracellular signal regulated kinase

FDA Food and Drug Administration

Fig. Figure

GPR G protein coupled receptor HE Haematoxylin and Eosin

HER Human Epidermal Growth Factor Receptor

HR Hazard ratio

H₂O₂ Hidrogen Peroxide

IGF-1 R Insulin Growth Factor-1 Receptor

IPATIMUP Instituto de Patologia e Imunologia Molecular da Universidade do Porto

kDa KiloDalton

MAPK Mitogen Activated Protein Kinase

MMP Matrix Metalloproteinase

μm micrometre
 Max Maximum
 Min Minimum
 min minute
 mg milligram
 mL millilitre
 mM milliMolar

mm² square millimetre

mRNA Messenger Ribonucleic Acid

OS Overall survival

PBS Phosphate buffered saline

P-cadherin Placental cadherin

PCNA Proliferating cell nuclear antigen
PI3K Phosphatidylinositol 3 kinase

PR Progesterone Receptor SMA Smooth muscle actin

SPSS Statistical Package for the Social Sciences

TA Transactivation
TF Transcription factor

TGF Transforming Growth Factor
USA United States of America
WHO World Health Organization

1. CANINE MAMMARY GLAND TUMOURS

General considerations

Similarly to rodents, felines and humans, canine mammary gland is frequently affected by spontaneous tumours, which represent the second most frequent neoplasia (Ferguson, 1985; Madewell and Theilen, 1987; Moulton, 1990). Spontaneous mammary gland tumours occur almost exclusively in female dogs, representing the most commonly occurring neoplasm and accounting for 25 to 50% of all neoplasias (Moulton, 1990; Misdorp *et al.*, 1999, 2002; Sorenmo, 2003). The exact incidence of these tumours is difficult to determine but it has been estimated in 105:100.000, which is three times higher compared to women (Brodey *et al.*, 1983).

Canine mammary tumours mainly affect middle-aged bitches (Loar, 1989; Moulton, 1990; Hellmén, 1996; Rutteman *et al.*, 2001), occur more often in caudal mammary glands and are clinically manifested as single or multiple nodules. Purebred dogs, namely spaniel breeds, pointers and dachshunds seem to be predisposed (Rutteman *et al.*, 2001; Misdorp, 2002).

Based on histological and biological criteria, it can be estimated that approximately one third to half of the surgically removed canine mammary tumours are malignant (Misdorp, 2002). Therefore, this disease represents a serious problem in worldwide veterinary practice and is a matter of concern for both oncologists and pathologists, which is ultimately reflected on the escalating number of studies in this research area. Furthermore, canine mammary tumours have attracted considerable attention over the years as possible animal models for human mammary neoplasia, based on their morphological and biological similarities (Gilbertson *et al.*, 1983).

Etiopathogenesis

Tumourigenesis is a multistep process comprising initiation, promotion and progression. The initiation of breast cancer is due to transforming (genetic and epigenetic) events in a single cell. Promotion and subsequent tumour progression are driven by the accumulation of additional genetic changes combined with clonal expansion and selection (Beckmann *et al.*, 1997; Porter *et al.*, 2001). So, invasive

mammary cancer is the endpoint of a multiple-step evolution that can be tracked as a series of progressive histological and molecular lesions. Under the microscope we can occasionally recognize the progression of hyperplasia, in situ carcinoma and invasive cancer, however the molecular evolution is less clearly understood (Ross, 1998).

The etiopathogenesis of canine mammary tumours is still unclear, despite several reported genetic alterations concerning oncogenes (Ahern *et al.*, 1996; Rungsipipat *et al.*, 1999; Martin de las Mulas *et al.*, 2003), tumour suppressor genes (Van Leeuwen *et al.*, 1996; Chu *et al.*, 1998; Veldhoen *et al.*, 1999) and the breast cancer susceptibility gene *BRCA*1 (Yuzbasiyan-Gurkan *et al.*, 1999). In addition, gross abnormalities in the nuclear DNA content (DNA aneuploidy) have been found in 50 to 60% of canine malignant mammary tumours. This aneuploidy reflects genetic instability, which is commonly at the basis of malignant transformation. In fact, some benign tumours were also found aneuploid, possibly reflecting their potential to progress to malignancy (Rutteman *et al.*, 1988a; Hellmén *et al.*, 1993; Rutteman *et al.*, 2001).

Human breast cancer represents a complex disease modulated by host factors, such as the hormonal status, which is involved in breast tumour development and progression (Ross, 1998). The participation of steroid receptors in the development of canine mammary tumours is not fully understood. It is known that early ovariohysterectomy offers a considerable protective effect, with the risk of developing mammary tumours increasing from 0.5% to 8%, and to 26%, depending on whether the ovariohysterectomy is performed before the first, second, or any oestrus thereafter, respectively (Schneider et al., 1969). Moreover, the administration of steroid hormones and their synthetic derivatives (progestins or progestin-estrogen combinations at high dosage) was found to promote the formation of mammary tumours in the dog (Misdorp et al., 1988; Stovring et al., 1997; Rutteman et al., 2001; Misdorp, 2002). It was also shown that mechanisms involved in progesterone-induced mammary gland tumours include an upregulation of growth hormone production within the mammary gland, where it has a direct growth stimulatory effect (Selman et al., 1994; Mol et al., 1999). Taken together, these findings suggest that steroid hormones might play an important role in the pathogenesis of canine mammary tumours (Sorenmo et al., 2000).

As in humans, advancing age, obesity and diet also seem to increase the risk of mammary tumours in the dog (Perez Alenza *et al.*, 2000), but a protective effect of early pregnancy has not been demonstrated (Rutteman *et al.*, 2001).

Pathology and natural behaviour

The classification of canine mammary neoplasms has been based mainly on standard histopathology (descriptive morphology), and to a lesser extent on the histogenetic origin and prognosis (Fowler *et al.*, 1974; Hampe and Misdorp, 1974; Destexhe *et al.*, 1993b; Benjamin *et al.*, 1999; Misdorp *et al.*, 1999; Misdorp, 2002). Canine mammary tumours are characterized by a complex morphology forming epithelial, mixed and mesenchymal tumours. All types can exist as benign and malignant forms: complex adenoma and mixed benign tumours constitute the dominant benign histotypes, whereas carcinomas represent the majority of malignant tumours. True malignant mixed tumours (carcinosarcomas) and sarcomas do exist but are uncommon (Misdorp *et al.*, 1999; Hellmén *et al.*, 2000). The current WHO classification of canine mammary tumours (Table 1) is both descriptive and prognostic, and subdivides carcinomas into noninfiltrating carcinomas, complex carcinomas (two cell types) and simple carcinomas (one cell type), in an attempt to rank the tumours by increasing malignant potential (Misdorp *et al.*, 1999).

Table 1. Histological classification of canine mammary tumours (Misdorp et al., 1999).

HISTOLOGICAL CLAS	SIFICATION OF CANINE MAMMARY TUMOURS
	Adenoma
	Simple adenoma
	Complex adenoma
BENIGN	Basaloid adenoma
TUMOURS	Fibroadenoma
	Low-cellularity fibroadenoma
	High-cellularity fibroadenoma
	Benign mixed tumour
	Duct papilloma
	Noninfiltrating (in situ) carcinoma
	Complex carcinoma
	Simple carcinoma
	Tubulopapillary carcinoma
	Solid carcinoma
	Anaplastic carcinoma
MALIGNANT	Special types of carcinomas
TUMOURS	Spindle cell carcinoma
1011100115	Squamous cell carcinoma
	Mucinous carcinoma
	Lipid-rich carcinoma
	Sarcoma
	Fibrosarcoma/ Osteosarcoma/ Other sarcomas
	Carcinosarcoma
	Carcinoma or sarcoma in benign tumour

Between 41 and 53% of all mammary tumours that occur in the female dog are considered malignant (Brodey *et al.*, 1983; Gilbertson *et al.*, 1983; Misdorp *et al.*, 1999; Moe, 2001; Rutteman *et al.*, 2001; Hellmén, 2005). Small, non-invasive, well-differentiated tumours are often treated effectively with surgery alone, but dogs with large, invasive, or poorly differentiated tumours are at risk of developing metastasis and dying of the disease. Approximately 30% of carcinomas cause metastases, usually via the lymphatics to the regional lymph nodes and the lungs, whereas more than 75% of sarcomas give rise to metastases, usually by the haematogenous route (Sorenmo, 2003; Hellmén, 2005). Nevertheless, considerable variations are observed in the biological behaviour among canine mammary tumours and histomorphological evidence of malignancy does not invariably imply a malignant clinical course (Rutteman *et al.*, 2001).

Histogenesis and differentiation

Canine mammary tumours are known for their biological and morphological heterogeneity (Nerurkar *et al.*, 1989; Moulton, 1990) and their precise histogenesis (especially of mixed tumours) has challenged veterinary pathologists ever since the early days of diagnostic pathology.

Mammary gland has a tubulo-alveolar structure composed of two cell layers, an inner luminal cell layer composed of glandular epithelial cells, and a distinct outer basal cell layer, juxtaposed to the basement membrane (Fig. 1A), composed of spindle-shaped or cuboidal myoepithelial cells, depending on their location and the hormonal status (Gusterson *et al.*, 2005). These cells have a common origin, arising from progenitor cells located in a suprabasal compartment between the luminal and the basal layer (Boecker *et al.*, 2002; Boecker and Buerger, 2003; Birnbaum *et al.*, 2004). It was shown that cells seem to exist at intermediate state of maturation, in both the epithelial and myoepithelial cell lineage (Boecker and Buerger, 2003).

There is overwhelming evidence that virtually all tumours are clonal and represent the progeny of a single cell. What is less clear is which cells within the tumour clone possess tumour-initiating cell function and are capable of maintaining cell growth (Dick, 2003). Substantial data suggest that both stem and progenitor cells may be the

targets of transformation during tumourigenesis, leading to breast cancer heterogeneity (Dontu *et al.*, 2004; Kalirai and Clarke, 2006).

Despite rare in the human breast, mixed mammary tumours are very frequent lesions in dogs and show many histological similarities with human pleomorphic adenomas of salivary glands (Genelhu et al., 2007). These tumours are characterized by the proliferation of an epithelial and a mesenchymal component, and a number of studies have addressed the histogenesis of metaplastic elements, like cartilage and bone (Misdorp et al., 1999). Besides glandular cells, morphologically different types of myoepithelial cells are observed, and several authors favoured a myoepithelial cell role in this tumour type histogenesis, based on immunohistochemical, electron microscopy and cell line studies (Pulley, 1973; Fowler et al., 1974; Tateyama and Cotchin, 1977, 1978; Destexhe et al., 1993b; Griffey et al., 1993; Arai et al., 1995; Gärtner et al., 1999; Misdorp et al., 1999; Tateyama et al., 2001; Espinosa de Los Monteros et al., 2002; Gama et al., 2003; Ramalho et al., 2006). Similarly, some human studies have also disclosed a myoepithelial cell histogenesis for benign pleomorphic adenoma of salivary glands (Erlandson et al., 1984). However, others have refuted this hypothesis and have supported an epithelial (Monlux et al., 1977), stromal (Palmer and Monlux, 1979; Nerurkar et al., 1989; Vos et al., 1993) or, more recently, a stem cell ontogeny (Hellmén and Lindgren, 1989; Hellmén et al., 2000) for canine mixed neoplasms. As for human metaplastic carcinomas (which include mixed malignant tumours), several lines of evidence favour a monoclonal origin for both epithelial and mesenchymal elements (Thompson et al., 1996; Zhuang et al., 1997; Wada et al., 1998), and a number of studies support a basal/myoepithelial histogenesis or differentiation (Sapino et al., 1992; Reis-Filho et al., 2003; Leibl et al., 2005).

Human breast cancers, as determined morphologically, were thought to arise exclusively from the inner, luminal epithelial cell compartment of the terminal-duct lobular unit of the breast. Irrespective of the true histogenesis of breast carcinoma, it has become increasingly clear that a small proportion of cancers (up to 18%) exhibit a partial or complete basal/myoepithelial phenotype, meaning they express molecules normally seen in the basal/myoepithelial compartment of the normal breast (Zhuang *et al.*, 1997; Tsuda *et al.*, 2000; Lakhani and O'Hare, 2001).

Although mixed tumours are rather frequent canine benign mammary lesions, the malignant counterpart is uncommon. However, myoepithelial differentiation is a

frequent finding in canine mammary malignant tumours, accompanied by the proliferation of glandular epithelial cells in the so-called complex carcinomas. Other frequent carcinomas are of simple type, usually thought to arise from luminal epithelial cells. Nevertheless, as in humans, several immunohistochemical studies pointed out to the presence of a basal/myoepithelial cell phenotype in a subset of simple carcinoma cases, which was not readily recognizable by routine histological evaluation only (Destexhe *et al.*, 1993b; Griffey *et al.*, 1993; Gama *et al.*, 2003). This "basal" differentiation has raised the attention from pathologists, since these were high grade tumours that presented an aggressive behaviour and poor patient prognosis (Griffey *et al.*, 1993; Tsuda *et al.*, 2000; Jones *et al.*, 2001; Laakso *et al.*, 2005).

In human breast cancer studies, this intriguing phenotype has reemerged in the past few years, due to the introduction of high-throughput technologies. Recent gene expression cDNA microarray studies have made it possible to distinguish two major tumour classes of breast cancer: one with the characteristics of basal/myoepithelial and the other with the characteristics of luminal cells (Perou *et al.*, 2000). Of major importance is the prognostic significance of basal-like cancers, which are frequently associated with poor clinical outcome. Basal-like tumours are hormonal receptor negative and express genes characteristic of basal and myoepithelial cells (Sorlie *et al.*, 2001, 2003; Sotiriou *et al.*, 2003).

Prognosis

As in humans, the identification of parameters with prognostic relevance constitutes a major area of investigation in canine mammary cancer (Bratulic *et al.*, 1996; Lohr *et al.*, 1997; Funakoshi *et al.*, 2000; Geraldes *et al.*, 2000; Nieto *et al.*, 2000) and in the last years an increasing number of potential prognostic factors (clinicopathological and molecular factors) have been investigated.

Although not consensual, several clinicopathological features have been recognized as prognostic factors in the vast majority of canine mammary cancer studies, based on univariate and/or multivariate analysis: tumour size (Bostock, 1975; Misdorp and Hart, 1976; Yamagami *et al.*, 1996b; Chang *et al.*, 2005; Martin de las Mulas *et al.*, 2005), ulceration (Hellmén *et al.*, 1993; Peña *et al.*, 1998; Queiroga and Lopes, 2002), tumour histological type (Misdorp and Hart, 1976; Hellmén *et al.*, 1993; Chang *et al.*, 2005)

and grade (Karayannopoulou *et al.*, 2005; Martin de las Mulas *et al.*, 2005), degree of invasion (Bostock, 1975; Misdorp and Hart, 1976; Gilbertson *et al.*, 1983; Hellmén *et al.*, 1993; Martin de las Mulas *et al.*, 2005), presence of lymph node and distant metastasis (Hellmén *et al.*, 1993; Yamagami *et al.*, 1996b; Queiroga and Lopes, 2002; Philibert *et al.*, 2003; Chang *et al.*, 2005).

Tumour size has been found to be an independent prognostic factor in a number of studies, with tumours smaller than 3 cm in diameter associated with a significantly better prognosis (Misdorp and Hart, 1976; Yamagami *et al.*, 1996b).

Tumour type was found to be an important factor in several studies, which described a range of increasing malignancy from complex carcinoma to simple carcinoma to sarcoma (Misdorp and Hart, 1976; Hellmén *et al.*, 1993; Philibert *et al.*, 2003). Carcinomas with myoepithelial cell proliferation (complex carcinomas) are usually associated with longer survival times, and although the mechanisms still remain unclear, this fact may be related to the putative role of myoepithelial cells as natural invasion tumour suppressors (Yamagami *et al.*, 1996b). Within the group of simple carcinomas, an increasing order of malignancy was observed from tubulopapillary to solid to anaplastic carcinoma (Bostock, 1975; Misdorp *et al.*, 1999). Carcinosarcomas were also associated with poor prognosis, with most dogs developing metastasis within the first year after surgery (Benjamin *et al.*, 1999).

Most studies found histological grade as being a reliable prognostic factor in canine mammary tumours (Misdorp and Hart, 1976; Gilbertson *et al.*, 1983; Peña *et al.*, 1998; Benjamin *et al.*, 1999). Initial criteria proposed for histological grading of canine mammary carcinomas by Misdorp and Hart (1976) and Gilbertson *et al.* (1983) were based on a combination of rather subjective cellular features. Recently, Karayannopoulou *et al.* (2005) applied the Elston and Ellis human grading method for histological grading and found it predictive for dog mammary tumours. This method (based on tubule formation, nuclear pleomorphism and mitotic count evaluation) is apparently more consistent and shows reproductive results (Elston and Ellis, 1998).

Factors that do not seem to be associated with prognosis are breed, tumour location, number of tumours and type of surgery (as long as histologically adequate resection is achieved) (Schneider *et al.*, 1969; Misdorp and Hart, 1976; Hellmén *et al.*, 1993; Yamagami *et al.*, 1996b; Rutteman *et al.*, 2001). Controversial findings exist regarding animal age and ovariohysterectomy at the time of surgery. Some studies state that

older animals are associated with poorer survival (Schneider *et al.*, 1969; Hellmén *et al.*, 1993; Peña *et al.*, 1998) while others report no statistical influence of age on survival (Hellmén *et al.*, 1993; Queiroga and Lopes, 2002; Philibert *et al.*, 2003; Chang *et al.*, 2005). As for ovariectomy, two distinct groups demonstrated that dogs spayed at the time of surgery survived longer than intact dogs (Sorenmo *et al.*, 2000; Chang *et al.*, 2005), in contrast to other studies reporting no effect of simultaneous ovariectomy on survival (Yamagami *et al.*, 1996a; Morris *et al.*, 1998; Philibert *et al.*, 2003).

Classical clinicopathological factors are not always sufficient to predict the biological behaviour of canine mammary tumours and the availability and application of new methodologies have allowed the identification of new prognostic factors, some potentially relevant as therapeutic targets. Next, we will focus on some molecular factors with potential prognostic value in canine mammary cancer.

2. DIFFERENTIATION AND PROGNOSTIC MARKERS IN MAMMARY CANCER: FROM DOGS AND HUMANS

Differentiation cell markers in mammary cancer

Mammary epithelial cells can be recognized by their distinct immunoprofile: luminal epithelial cells are characterized by the expression of luminal cytokeratins (CK) 7, 8, 18 and 19, whereas basal/myoepithelial cells express basal CK 5, 14 and 17, p63, P-cadherin, CD10 and EGFR, among other markers (Malzahn *et al.*, 1998; Boecker and Buerger, 2003). Due to its contractile phenotype, myoepithelial cells also express smooth muscle-specific proteins such as smooth muscle actin (SMA) and calponin (Adriance *et al.*, 2005; Espinosa de los Monteros *et al.*, 2002). The hunt for specific markers of the different types of epithelial cells is ongoing and several markers have already been successfully used in human and canine mammary tissues (Gama *et al.*, 2003; Reis-Filho *et al.*, 2003) (Fig. 1B). Some fundamental considerations will now be addressed, regarding a few selected molecular markers.

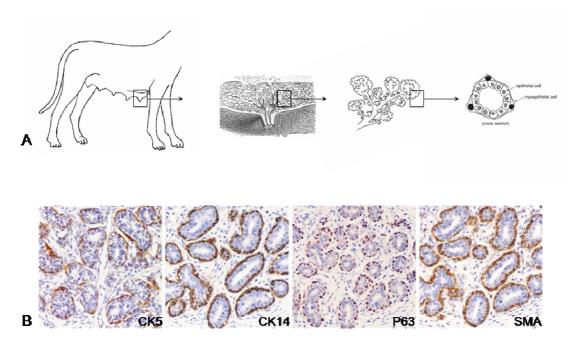


Fig. 1. Normal canine mammary gland: schematic representation (A) and immunohistochemical reactivity to basal/myoepithelial cell markers (B). A. Schematic representation of canine mammary gland, showing its anatomical location and organization. Mammary epithelium is organized as a bilayer, with a luminal layer of secretory epithelial cells, and a basal layer of myoepithelial cells. B. Normal mammary gland stained with antibodies to CK5, CK14, p63 and SMA, highlighting the basally located myoepithelial cells.

Cytokeratins (CK). CK are the typical intermediate filament proteins of epithelia and are essential for normal tissue structure and function (Schweizer *et al.*, 2006). Besides major components of the epithelial cytoskeleton, CK are highly dynamic and have also been involved in intracellular signalling pathways (Moll *et al.*, 2008).

The human keratin family shows an outstanding degree of molecular diversity and includes 54 distinct elements (Moll *et al.*, 2008). CK are encoded by *KRT* genes mostly clustered on paralogous regions of 12q and 17q chromosome arms and are classified, either upon type and isoelectric point, i.e. type I acidic (CK9-10, CK12-28 and CK31-40) and type II neutral-basic (CK1-8 and CK71-86) or upon molecular mass, i.e. low and high molecular weight CK (such as CK18/19 and CK5/6, respectively) (Chu and Weiss, 2002; Moll *et al.*, 2008).

The stability of intermediate filaments makes it possible to characterize and study tumour histogenesis but although epithelial tissues tend to retain their characteristic CK pattern throughout carcinogenesis, modulations may occur within a certain range of possibilities during carcinoma development and progression (Malzahn *et al.*, 1998; Abd El-Rehim *et al.*, 2004; Birnbaum *et al.*, 2004; Laakso *et al.*, 2005). It has long been suggested that certain constituent proteins of the cytoskeletal intermediate filaments may be of relevance with respect to the biological behaviour and prognosis of human breast carcinomas (Dairkee *et al.*, 1987; Raymond and Leong, 1989; Takei *et al.*, 1995; Schaller *et al.*, 1996). In fact, the immunoexpression of basal-type CK, such as CK5, CK14 and CK17, has been associated with a poor prognosis for many years (Dairkee *et al.*, 1987; Malzahn *et al.*, 1998). After the rediscovery of basal-like carcinomas by gene expression microarray analysis, numerous immunohistochemical studies confirmed these earlier findings (van de Rijn *et al.*, 2002; Abd El-Rehim *et al.*, 2004; Gusterson *et al.*, 2005).

A few number of studies are available concerning cytokeratin expression in canine mammary tumours (Hellmén and Lindgren, 1989; Destexhe *et al.*, 1993b; Griffey *et al.*, 1993); however, Griffey and coworkers (1993), based on CK14 expression, already described a basal phenotype for canine mammary carcinomas, which was also characterized by an aggressive clinical behaviour (Griffey *et al.*, 1993).

Placental cadherin (P-cadherin). P-cadherin is a member of the cadherin family, along with epithelial cadherin (E-cadherin) and neural cadherin (N-cadherin). Cadherins are calcium-dependent cell-to-cell adhesion molecules which play critical roles during embryonic development and in the maintenance of normal tissue architecture (Nose and Takeichi, 1986; Nose et al., 1987; Takeichi, 1991, 1993, 1995; Gumbiner, 1996). In a development setting, P-cadherin is transiently expressed in various tissues (Hirai et al., 1989); it was localized to the cap cells of terminal end buds in the developing murine mammary gland, which might represent mammary stem cells (Williams and Daniel, 1983; Daniel et al., 1995).

P-cadherin expression in adult tissues is limited to epithelium, located at cell-cell boundaries (Shimoyama *et al.*, 1989; Shimoyama and Hirohashi, 1991). Unlike E-cadherin, which is broadly distributed in all epithelial tissues, P-cadherin exhibits a singular pattern of expression, co-localizing partially with E-cadherin and being restricted to the basal proliferative cell layer of the majority of stratified epithelia (reviewed by Paredes *et al.*, 2007).

In normal human and canine mammary gland, P-cadherin is restricted to myoepithelial cells, representing a sensitive marker for this cell type (Rasbridge et al., 1993; Palacios et al., 1995; Kovacs and Walker, 2003; Gama et al., 2004). However, during lactation, P-cadherin is not found at cell-cell borders, as expected for an adhesion molecule, but rather appears to be secreted by epithelial cells (Soler et al., 2002; Gama et al., 2002). In human breast cancer, P-cadherin was found to be expressed by a subset of carcinomas, frequently with a basal epithelial phenotype (Arnes et al., 2005). Pcadherin is commonly identified in medullary and metaplastic carcinomas, further suggesting a basal/myoepithelial cell histogenetic origin or line of differentiation for these tumours (Han et al., 1999; Reis-Filho et al., 2003). P-cadherin expressing tumours are usually associated with aggressive behaviour and poor outcome (Palacios et al., 1995; Peralta Soler et al., 1999; Gamallo et al., 2001; Kovacs et al., 2003; Paredes et al., 2002, 2005; Arnes et al., 2005), which has raised the interest on Pcadherin as a potential prognostic marker for breast cancer. P-cadherin expression was found inversely correlated with hormonal receptor status (Paredes et al., 2002a, 2002b) and it seems to be associated with an estrogen-independent tumour growth (Paredes et al., 2002a). Some authors actually described this molecule as an independent marker of

poor prognosis (Peralta Soler *et al.*, 1999), with its expression highly predictive of a poor outcome in small, node-negative breast cancers (Arnes *et al.*, 2005).

In canine mammary tumours, we have also described P-cadherin expression in a subset of malignant carcinomas. A significant association was found between its expression and tumour type, being highly positive in carcinosarcoma and spindle cell carcinoma, favouring a probable basal/myoepithelial differentiation (Gama *et al.*, 2004).

P63. P63 is a recently characterized member of the p53 family (Yang et al., 1998; Kaelin, 1999; Little and Jochemsen, 2002). P63 gene is located on chromosome 3q27 (Yang et al., 1998) and exhibits a high homology to p53, leading to the early speculation that p63 would also function as a tumour suppressor (Yang and McKeon, 2000; Westfall and Pietenpol, 2004). Despite its homology with p53, p63 gene encodes at least six different proteins, grouped in two distinct classes: one containing a region that is similar to p53 transactivation domain (TAp63 isoforms) and another lacking this domain (ΔNp63 isoforms) (Yang et al., 1998). P63 is rarely mutated (Osada et al., 1998; Hagiwara et al., 1999) and several studies described an overexpression of p63 splice variants in a subset of human epithelial tumours, sometimes associated with gene amplification, suggesting that p63 can act as an oncogene (Crook et al., 2000; Hibi et al., 2000; Park et al., 2000; Yamaguchi et al., 2000; Massion et al., 2003). Supporting this hypothesis, it was recently shown that deregulated TAp63 isoform predisposes to tumour development and progression (Koster et al., 2006) and that p63 contributes to cell invasion and migration in squamous cell carcinoma of the head and neck (Gu et al., 2008).

Although p63 function is not fully understood, the striking epithelial defects seen in p63-deficient mice suggest that this gene plays a key role in maintaining basal/progenitor epithelial cell populations (DiRenzo *et al.*, 2002). Whereas p53-/mice are developmentally normal but prone to neoplastic disease (Donehower *et al.*, 1992), p63 knockout mice have severe developmental abnormalities. Specifically, the p63-/- mice die shortly after birth and are deficient in the development of several epithelial tissues such as skin, prostate, mammary gland, and urothelia (Mills *et al.*, 1999; Yang *et al.*, 1999). Recently, Koster *et al.* (2004) demonstrated that TAp63 isoforms are the first to be expressed during embryogenesis and are required for

initiation of epithelial stratification. This program further involves a shift in the balance between p63 isoforms towards Δ Np63 to allow terminal differentiation. After epidermis maturation, persistently elevated p63 levels in the basal layer are required for the maintenance of the basal cell proliferative potential (Koster *et al.*, 2004).

Immunohistochemical analyses show p63 protein localization and expression in the basal/progenitor cells of several adult epithelial tissues such as the epidermis, hair follicles, sweat glands, cervix, tongue, esophagus, mammary glands, prostate, and urogenital tract, with Δ Np63 α being the predominant, if not only, p63 variant expressed (Yang *et al.*, 1998; Parsa *et al.*, 1999; Signoretti *et al.*, 2000; Barbareschi *et al.*, 2001; Pellegrini *et al.*, 2001; Di Como *et al.*, 2002; Westfall *et al.*, 2003).

P63 is consistently expressed in basal/myoepithelial cells of normal human and canine mammary gland and in tumours with basal/myoepithelial cell features (Barbareschi *et al.*, 2001; Nylander *et al.*, 2002; Wang *et al.*, 2002; Gama *et al.*, 2003; Reis-Filho and Schmitt, 2003; Ribeiro-Silva *et al.*, 2003b; Ramalho *et al.*, 2006). Thus, p63 has been proposed as a reliable basal/myoepithelial cell marker, being expressed by basal-like breast carcinomas, including metaplastic type (Reis-Filho *et al.*, 2003; Laakso *et al.*, 2005; Matos *et al.*, 2005). Some human breast cancer studies found an association between its expression and high grade, large tumour size, nodal metastasis and ER negativity (Ribeiro-Silva *et al.*, 2003a).

Prognostic markers in mammary cancer

Recently, prognostic value has been claimed for several molecular variables in canine mammary cancer studies (Zaidan Dagli, 2008), including cell proliferation markers (Peña *et al.*, 1998; Sarli *et al.*, 2002; De Matos *et al.*, 2006), receptor proteins (Graham *et al.*, 1999; Geraldes *et al.*, 2000; Nieto *et al.*, 2000), oncogenes/tumour suppressor genes (Ahern *et al.*, 1996; Lee *et al.*, 2004) and adhesion molecules (Brunetti *et al.*, 2005; Matos *et al.*, 2006), among others (Queiroga *et al.*, 2005; Pinho *et al.*, 2007). Despite their recognition as relevant prognostic indicators in human breast cancer, some of these factors have generated contradictory results and still lack validation in the veterinary area.

Tumour cell proliferation. Proliferation is a key feature in tumour progression and has been extensively investigated to evaluate prognosis in canine mammary cancer (Destexhe et al., 1993a; Bratulic et al., 1996; Peña et al., 1998; Sarli et al., 2002; Zuccari et al., 2004; De Matos et al., 2006). Several methods were performed including AgNOR quantification (Bostock et al., 1992; Bratulic et al., 1996; Lohr et al., 1997), DNA flow cytometry measurement of S-phase fraction (Hellmén et al., 1993) and immunohistochemical analysis of PCNA and Ki-67 (Preziosi et al., 1995; Peña et al., 1998; Geraldes et al., 2000; Sarli et al., 2002; Zacchetti et al., 2003; De Matos et al., 2006).

Currently, proliferation is widely estimated by immunohistochemical assessment of Ki-67, both in human (Bouzubar *et al.*, 1989; Brown and Gatter, 1990; Veronese and Gambacorta, 1992; Barginear *et al.*, 2008) and in canine cancers (Peña *et al.*, 1998; Sarli *et al.*, 2002; De Matos *et al.*, 2006), with most studies describing an association between high proliferation and poor prognosis. Ki-67 is a non-histone nuclear protein detected in all cell cycle phases except the resting phase (G0) and is therefore a direct indicator of the tumour growth fraction (Durchow *et al.*, 1994).

Hormone receptors. Oestrogen and oestrogen receptors (ER) play essential roles in both normal breast development and breast cancer progression (Pearce and Jordan, 2004). Oestrogen exerts its biological effects usually by binding to ER (ERα and ERβ), which mainly exists in the nucleus as a member of the nuclear receptor superfamily of transcription factors. The oestrogen–ER complex through genomic and nongenomic pathways, leads to nuclear and extranuclear processes that promote cellular proliferation and differentiation (Fig. 2) (Murphy and Watson, 2002; Yamaguchi, 2007). Growth factor signalling pathways can also activate ER via phosphorylation, in a ligand-independent manner (Le Goff *et al.*, 1994; Lee *et al.*, 2001).

Progesterone Receptor (PR) is an oestrogen-regulated gene and its expression is therefore thought to indicate a functioning ER pathway. Theoretically, the assessment of PR should assist in predicting response to hormonal therapy more accurately. In keeping with this, there is some evidence that tumours positive for PR are more likely to respond to tamoxifen (Ravdin *et al.*, 1992; Bardou *et al.*, 2003) but the predictive value of PR positivity in the absence of ER is still controversial (Payne *et al.*, 2008).

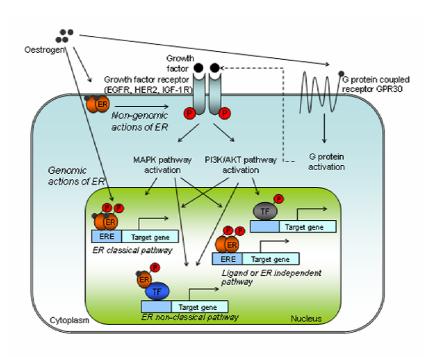


Fig. 2. Simplified overview of intracellular estrogen action mechanisms. In the classical genomic pathway, ligand-bound ERs bind directly to the estrogen response element (ERE) present in the target gene promoters. Alternatively, in the non-classical pathway ER acts as a coactivator via interaction with other transcription factors (TF), which regulates the gene transcriptions at their specific DNA sites. Membrane-initiated (non-genomic) steroid signalling has also been reported, either, controversially, as a small pool of ER within the plasma membrane, or via non-ER proteins, such as GPR30. The former can contribute to the oestrogenic response via cross-talk with growth factor-mediated pathways, which activate targeted transcription factors and/or coactivators via phosphorylation, in an ER dependent or independent manner. The latter activates G-protein signalling pathways, which result in cleavage and release of membrane-bound growth factors such as EGF, which activates its receptor and initiates intracellular kinase cascades (adapted from Yamaguchi, 2007 and Speirs and Walker, 2007).

In humans, it has been standard practice for 25 years to analyze all invasive breast cancers for hormone receptor content as a means of estimating prognosis and predicting responsiveness to endocrine treatment (Yeh and Mies, 2008). The majority of available data in canine literature concerning steroid receptors are based on biochemical assays (Mialot *et al.*, 1982; Martin *et al.*, 1984; Parodi *et al.*, 1984;

Rutteman et al., 1988b; Sartin et al., 1992; Donnay et al., 1993). In human breast cancer studies, these techniques were replaced by immunohistochemical methods after the development of reliable monoclonal antibodies against oestrogen and progesterone receptors (Allred et al., 1990). Recent studies performed on canine tissues have proven steroid receptors value in characterizing subgroups with different prognosis among female dogs with mammary cancer (Graham et al., 1999; Nieto et al., 2000). Despite some studies describing an association between the absence of ER/PR or both and shorter survival time (Martin et al., 1984; Sartin et al., 1992; Nieto et al., 2000; Martin de las Mulas, 2005), others failed to find such a correlation (Millanta et al., 2005). Thus, there is still insufficient data in the literature on the prognostic significance of hormonal status and its application into diagnostic routine remains a matter of debate. ER inhibition through endocrine targeting, either directly using weak oestrogen agonists (Selective Oestrogen Receptor Modulators) or indirectly by blocking the conversion of androgens to oestrogen (e.g. aromatase inhibitors), forms the mainstay of adjuvant and metastatic human breast cancer therapy (Payne et al., 2008). In dogs, there is no convincing data indicating that hormonal treatment improves prognosis. In fact, although ER-positive canine mammary tumour cell lines have been shown to respond to the selective ER modulator tamoxifen (Sartin et al., 1993), adjuvant endocrine therapy is not currently used. Treatment of bitches with tamoxifen has been reported to produce estrogen-like side effects and at present is not advised for dogs (Rutteman et al., 2001).

Human Epidermal Growth Factor Receptor-2 (HER-2). The HER family of receptor tyrosine kinases includes four closely related members: Epidermal Growth Factor Receptor (EGFR, also called HER-1 or c-erbB-1), HER-2 (also called c-erbB-2 or neu), HER-3 (also called c-erbB-3), and HER-4 (also called c-erbB-4) (Holbro and Hynes, 2004). These receptors share a common structure comprising an extracellular ligand-binding domain, a transmembrane domain and an intracellular domain with tyrosine kinase activity. Binding of growth factor ligands causes homo- or hetero-dimerization with another family member, which leads to receptor-linked tyrosine kinase activation. This activation triggers a network of intracellular signalling pathways, mainly the mitogen activated protein kinase (MAPK) and the

phosphatidylinositol 3 kinase (PI3K)-AKT pathways, which produce diverse cellular events including cell proliferation, adhesion, migration, differentiation, angiogenesis and inhibition of apoptosis (Fig. 3) (Hanna *et al.*, 1999; Yarden and Sliwkowski, 2001; Kumar and Wang, 2002).

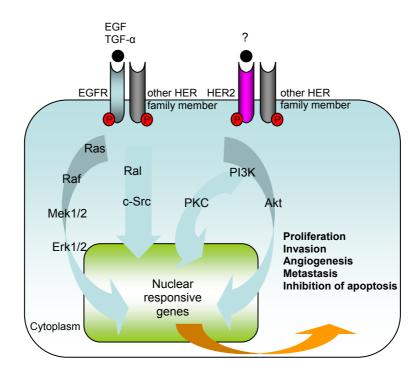


Fig. 3. Signalling pathways of HER family members. Ligand binding induces receptor dimerization and subsequent autophosphorylation of distinct tyrosine residues, creating docking sites for adaptor molecules and leading to the activation of downstream effector molecules. A variety of signalling pathways results in pleiotropic effects, including cell proliferation, control of the cell cycle, regulation of apoptosis and survival, and alterations in cell migration and invasiveness (adapted from Prenzel *et al.*, 2001 and Vlahovic and Crawford, 2003).

The human *HER2* gene maps to chromosome 17q21 and encodes a 185 kDa glycoprotein. It is reported to be amplified and overexpressed in several types of human tumours, including breast cancer (Hynes and Lane, 2005). HER-2 overexpression is found in 15–30% of human breast carcinomas and correlates with more aggressive clinicopathological features, drug resistance or sensitivity to specific chemotherapy and hormonal therapy regimens in breast cancer (Slamon *et al.*, 1987; Revillion *et al.*, 1998; Yamauchi *et al.*, 2001; Burstein, 2005). Amplification is the

predominant mechanism of gene overexpression and is present in about 85–90% of the cases (Hoang *et al.*, 2000; Jimenez *et al.*, 2000).

HER-2 was one of the first oncogenes studied in clinical samples of invasive breast cancer. Its early significance as a prognostic factor has been surpassed by its key importance as a predictive factor of response to particular systemic therapies, notably trastuzumab (HerceptinTM), a humanized monoclonal antibody which has been shown in several studies to improve response rates, time to progression and overall survival (Slamon *et al.*, 2001; Romond *et al.*, 2005; Smith *et al.*, 2007).

While the expression of HER-2 has been extensively investigated in human breast tumours, only a limited number of studies are available concerning HER-2 status in canine tumours (Ahern *et al.*, 1996; Schafer *et al.*, 1998; Rungsipipat *et al.*, 1999; Matsuyama *et al.*, 2001; Martin de las Mulas *et al.*, 2003). Furthermore, and despite HER-2 recognition as a prognostic factor in human breast cancer (Slamon *et al.*, 1987; Revillion *et al.*, 1998), the significance of HER-2 overexpression in dogs with mammary carcinoma is still unclear. Some studies have shown that either HER-2 amplification (Ahern *et al.*, 1996) or protein overexpression (Rungsipipat *et al.*, 1999) are present in canine mammary carcinomas; nevertheless, a subsequent study addressing simultaneously HER-2 protein and gene status found no gene amplification in overexpressing tumours (Martin de las Mulas *et al.*, 2003).

Canine *HER-2* has been mapped to 1q13.1 and cytogenetic studies of canine tumours revealed that this region is very often affected by clonal chromosome aberrations, which might be associated with HER-2 protein overexpression (Murua Escobar *et al.*, 2001). In canine mammary carcinomas, HER-2 overexpression was found usually associated with established indicators of poor prognosis (Martin de las Mulas *et al.*, 2003; Dutra *et al.*, 2004). However, a recent study performed by Hsu *et al.* (2007) revealed that HER-2 overexpression in canine malignant mammary tumours is associated with higher survival rates (Hsu *et al.*, 2007).

Epidermal Growth Factor Receptor (EGFR). The *EGFR* gene maps to human chromosome 7p11.2-p2 and encodes for a 170 kDa transmembrane tyrosine kinase which is activated by several ligands, including EGF and TGF- α (reviewed in Suo and Nesland, 2002). EGFR was the first tyrosine kinase transmembrane receptor to be

directly linked with human cancer (Hynes and Lane, 2005). Its expression in normal and neoplastic breast has been extensively studied, since EGFR is required for normal mammary development and lactation (Kumar and Wang, 2002). Recent studies have showed EGFR to be frequently expressed in basal cell layers and in myoepithelial cells (Santini *et al.*, 2002; DiRenzo *et al.*, 2002) and several authors pointed out EGFR as a possible basal cell marker (Korsching *et al.*, 2002; Nielsen *et al.*, 2004).

EGFR overexpression is observed in approximately 16-48% of all breast cancers, although methodology and positivity criteria differ widely among studies (Klijn *et al.*, 1992; Fox *et al.*, 1994; Toi *et al.*, 1994; Tsutsui *et al.*, 2002; Rampaul *et al.*, 2005). EGFR expression is common in basal-like breast cancers, being found in up to 60% of basal-like breast carcinomas (Nielsen *et al.*, 2004; Reis-Filho *et al.*, 2005, 2006; Livasy *et al.*, 2006; Turner and Reis-Filho, 2006).

EGFR overexpression has been shown to be associated with aggressive biological properties and poor clinical outcome (Nicholson *et al.*, 1990; Tsutsui *et al.*, 2002; Tovey *et al.*, 2004), but the validity of EGFR as a useful prognostic factor for human breast cancer is still uncertain (Nicholson *et al.*, 2001; Rampaul *et al.*, 2004; Park *et al.*, 2007). The interest in EGFR is further enhanced by the availability and recent FDA approval of specific EGFR tyrosine kinase inhibitors (Bhargava *et al.*, 2005).

Molecular mechanisms for EGFR overexpression in the majority of breast cancer cases are yet to be identified (Reis-Filho *et al.*, 2006). *EGFR* gene amplifications are rather uncommon in breast cancer, with the exception of basal-like carcinomas (Reis-Filho *et al.*, 2005, 2006; Park *et al.*, 2007). In addition, *EGFR* activating mutations represent remarkably rare findings (Bhargava *et al.*, 2005; Weber *et al.*, 2005; Reis-Filho *et al.*, 2006) and other regulatory mechanisms are certainly involved, possibly at the transcriptional level (Kersting *et al.*, 2004, 2006; Park *et al.*, 2007; Sassen *et al.*, 2008). In contrast to human literature (Klijn *et al.*, 1992; Bhargava *et al.*, 2005; Reis Filho *et al.*, 2005, 2006; Park *et al.*, 2007), a limited number of reports are available concerning EGFR status in canine mammary tissues, with most studies based on biochemical assays (Nerurkar *et al.*, 1987; Rutteman *et al.*, 1990; Donnay *et al.*, 1993; Rutteman *et al.*, 1994; Donnay *et al.*, 1996). These studies failed to find a relation between EGFR concentrations and clinicopathological parameters. Yet, some studies found an inverse correlation between EGFR and ER concentrations in malignant tumours (Nerurkar *et al.*, 1987), while others described a positive (Donnay *et al.*, 1993; Donnay *et al.*, 1996)

or no existing correlation (Rutteman *et al.*, 1994). EGFR mRNA expression was recently described in canine mammary tissues, with controversial results in normal mammary gland (Matsuyama *et al.*, 2001). According to this study, EGFR expression was present in tumours but it was not observed in normal tissues, contradicting previous canine (Rutteman *et al.*, 1994) and human findings (Moller *et al.*, 1989; Santini *et al.*, 2002).

E-Cadherin. E-cadherin (also called uvomorulin, L-Cam, cell-Cam120/80 or Arc-1) is a 120 kDa transmembrane glycoprotein whose extracellular domain promotes cell-to-cell adhesion, while the intracellular domain interacts with catenins (α -, β - and γ -catenins) which link cadherins to the actin cytoskeleton (Fig. 4) (Ozawa *et al.*, 1989; Vleminckx *et al.*, 1991; Knudsen *et al.*, 1998). β-catenin also participates in a signal transduction cascade as part of the Wnt signalling pathway (Berx *et al.*, 2001; Brown, 2001). Another catenin-like molecule, p120, has been identified in association with E-cadherin at the cell-cell junctions, although this complex does not appear to form a link with the actin cytoskeleton (Reynolds *et al.*, 1994).

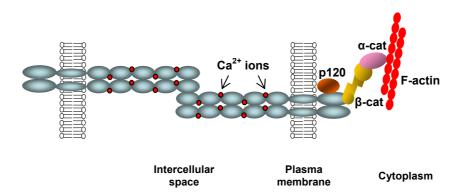


Fig. 4. Schematic representation of the classical cadherin-catenin complex. Classical cadherins (blue), which mediate calcium-dependent (red) intercellular adhesion, are composed by an extracellular domain, a transmembrane domain and a cytoplasmic domain. This last domain comprises a juxtamembrane domain, which binds p120-catenin (orange), and a catenin-binding domain, which binds β -catenin (yellow). β -catenin binds α -catenin (violet), which establishes a direct link between the cadherin-catenin complex and the actin cytoskeleton (red) (adapted from Paredes *et al.*, 2007).

Normal E-cadherin expression and function are essential for the induction and maintenance of a polarized and differentiated epithelium during embryonic development (Takeichi, 1991). In adult epithelial tissues, an intact complex is required for the maintenance of normal intercellular adhesion. In the light of this, several authors have proposed that E-cadherin might function as an invasion suppressor molecule such that a disturbed function of E-cadherin-catenin complex theorethically enhances the tumour cell invasive potential (Wijnhoven *et al.*, 2000).

In normal human and canine mammary gland, E-cadherin is expressed by both luminal and myoepithelial cells at cell-cell borders (Rasbridge *et al.*, 1993; Palacios *et al.*, 1995; Restucci *et al.*, 1997). In human breast cancer, the loss or reduction of the E-cadherin-catenin complex has been extensively associated with tumour progression (Oka *et al.*, 1993; Siitonen *et al.*, 1996; Bukholm *et al.*, 1998; Heimann *et al.*, 2000; Madhavan *et al.*, 2001). In general, loss of E-cadherin expression correlates with undifferentiated breast carcinomas, but the available studies differ with regard to its association with survival and its value as a prognostic marker is still controversial (Knudsen and Wheelock, 2005; Gould Rothberg and Bracken, 2006). Some studies report that the combination of E-cadherin and one of the catenins is of better prognostic value than the evaluation of individual components (Zschiesche *et al.*, 1997; Gofuku *et al.*, 1999).

The role of E-cadherin and catenins in canine mammary tumours is still poorly understood (Restucci *et al.*, 1997; Reis *et al.*, 2003; Sarli *et al.*, 2004; Brunetti *et al.*, 2005; Matos *et al.*, 2006; de Matos *et al.*, 2007; Nowak *et al.*, 2007; Rodo and Malicka, 2008). Brunetti *et al.* (2005) reported that reduced E-cadherin/β-catenin expression was associated with invasion, but no correlation was found regarding survival. More recent studies also described a significant correlation between E-cadherin loss and several classic prognostic features (Matos *et al.* 2006), as well as with invasion and lymph node metastases, suggesting this molecule as a potential prognostic marker for canine mammary cancer (de Matos *et al.*, 2007).

REFERENCES

- Abd El-Rehim DM, Pinder SE, Paish CE, Bell J, Blamey RW, Robertson JF, Nicholson RI, Ellis IO. (2004). Expression of luminal and basal cytokeratins in human breast carcinoma. J Pathol 203:661–671.
- Adriance MC, Inman JL, Petersen OW, Bissell MJ. (2005). Myoepithelial cells: good fences make good neighbors. Breast Cancer Res 7:190-197.
- Ahern TE, Bird RC, Bird AE, Wolfe LG. (1996). Expression of the oncogene c-erbB-2 in canine mammary cancers and tumor derived cell lines. Am J Vet Res 57:693–696.
- Allred DC, Bustamante MA, Daniel CO, Gaskill HV, Cruz AB Jr. (1990). Immunocytochemical analysis of estrogen receptors in human breast carcinomas: evaluation of 130 cases and review of the literature regarding concordance with biochemical assay and clinical relevance. Arch Surg 125:107–113.
- Arai K, Uehara K, Naoi M. (1995). Simultaneous expression of type IX collagen and an inhibin-related antigen in proliferative myoepithelial cells with pleomorphic adenoma of canine mammary glands. Jpn J Cancer Res 86:577-584.
- Arnes JB, Brunet JS, Stefansson I, Begin LR, Wong N, Chappuis PO, Akslen LA, Foulkes WD. (2005). Placental cadherin and the basal epithelial phenotype of BRCA1-related breast cancer. Clin. Cancer Res. 11:4003-4011.
- Barbareschi M, Pecciarini L, Cangi MG, Macri E, Rizzo A, Viale G, Doglioni C. (2001). p63, a p53 homologue, is a selective nuclear marker of myoepithelial cells of the human breast. Am J Surg Pathol 25:1054-1060.
- Bardou VJ, Arpino G, Elledge RM, Osborne CK, Clark GM. (2003). Progesterone receptor status significantly improves outcome prediction over estrogen receptor status alone for adjuvant endocrine therapy in two large breast cancer databases. J Clin Oncol 21:1973–1979.
- Barginear MF, Bradley T, Shapira I, Budman DR. (2008). Implications of applied research for prognosis and therapy of breast cancer. Crit Rev Oncol Hematol 65:223–234.
- Beckmann MW, Niederacher D, Schnürch H-G, Gusterson BA, Bender HG (1997). Multistep carcinogenesis of breast cancer and tumour heterogeneity. J Mol Med 75:429–439.
- Benjamin SA, Lee AC, Saunders WJ. (1999). Classification and behaviour of canine mammary epithelial neoplasms based on life-span observations in beagles. Vet Pathol 36:423-436.
- Berx G, van Roy R. (2001). The E-cadherin/catenin complex: An important gatekeeper in breast cancer tumorigenesis and malignant progression. Breast Cancer Res 3:289–293.

- Bhargava R, Gerald WL, Li AR, Pan Q, Lal P, Ladanyi M, Chen B. (2005). EGFR gene amplification in breast cancer: correlation with epidermal growth factor receptor mRNA and protein expression and HER-2 status and absence of EGFR-activating mutations. Modern Pathol 18:1027-1033.
- Birnbaum D, Bertucci F, Ginestier C, Tagett R, Jacquemier J, Charafe-Jauffret E. (2004). Basal and luminal breast cancers: basic or luminous? Int J Oncol 25:249–258.
- Boecker W, Moll R, Poremba C, Holland R, Van Diest PJ, Dervan P, Buerger H, Wai D, Diallo RI, Brandt B, Herbst H, Schmidt A, Lerch MM, Buchwallow IB. (2002). Common adult stem cells in the human breast give rise to glandular and myoepithelial cell lineages: a new cell biological concept. Lab Invest 82:737–746.
- Boecker W, Buerger H. (2003). Evidence of progenitor cells of glandular and myoepithelial cell lineages in the human adult female breast epithelium: a new progenitor (adult stem) cell concept. Cell Prolif 36(Suppl 1):73–84.
- Bostock DE (1975). The prognosis following the surgical excision of canine mammary neoplasms. Eur J Cancer 11:389-396.
- Bostock DE, Moriarty J, Crocker J. (1992). Correlation between histologic diagnosis means nucleolar organizer region count and prognosis in canine mammary tumors. Vet Pathol 29:381–385.
- Bouzubar N, Walker KJ, Griffiths K, Ellis IO, Elston CW, Robertson JF, Blamey RW, Nicholson RI. (1989). Ki67 immunostaining in primary breast cancer: pathological and clinical associations. Br J Cancer 59:943-947.
- Bratulic M, Grabarevic Z, Artukovic B, Capak D. (1996). Number of nucleoli and nucleolar organizer regions per nucleous and nucleolus- prognostic value in canine mammary tumors. Vet Pathol 33:527-532.
- Brodey RS, Goldschmidt MH, Roszel JR. (1983). Canine mammary gland neoplasms. J Am Animal Hosp Assoc 19:61-90.
- Brown AM. (2001). Wnt signaling in breast cancer: have we come full circle? Breast Cancer Res 3:351-355.
- Brown DC, Gatter KC. (1990). Monoclonal antibody Ki-67: its use in histopathology. Histopathology 17:489-503.
- Brunetti B, Sarli G, Preziosi R, Monari I, Benazzi C. (2005). E-cadherin and β-catenin reduction influence invasion but not proliferation in canine malignant mammary tumors. Vet Pathol 42:781-787.
- Bukholm IK, Nesland JM, Karesen R, Jacobsen U, Borresen-Dale A-L. (1998). E-cadherin and α -, β -, and γ -catenin protein expression in relation to metastasis in human breast carcinoma. J Pathol 185:262-266.

- Burstein HJ. (2005). The distinctive nature of HER2-positive breast cancers. N Engl J Med 353:1652–1654.
- Chang SC, Chang CC, Chang TJ, Wong ML. (2005). Prognostic factors associated with survival two years after surgery in dogs with malignant mammary tumours: 79 cases (1998-2002). J Am Vet Med Ass 227:1625-1629.
- Chu LL, Rutteman GR, Kong JM, Ghahremani M, Schmeing M, Misdorp W, van Garderen E, Pelletier J. (1998). Genomic organization of the canine p53 gene and its mutational status in canine mammary neoplasia. Breast Cancer Res Treat 50:11-25.
- Chu PG, Weiss LM. (2002). Keratin expression in human tissues and neoplasms. Histopathology 40:403-439.
- Crook T, Nicholls JM, Brooks L, O'Nions J, Allday MJ. (2000). High level expression of deltaN-p63: a mechanism for the inactivation of p53 in undifferentiated nasopharyngeal carcinoma (NPC)? Oncogene 19:3439-3444.
- Dairkee SH, Ljung BM, Smith H, Hackett A. (1987). Immunolocalization of a human basal epithelium specific keratin in benign and malignant breast disease. Breast Cancer Res Treat 10:11–20.
- Daniel CW, Strickland P, Friedmann Y. (1995). Expression and functional role of E- and P-cadherins in mouse mammary ductal morphogenesis and growth. Dev Biol 169:511-519.
- De Matos AJ, Lopes CC, Faustino AM, Carvalheira JG, Dos Santos MS, Rutteman GR, Gärtner MF. (2006). MIB-1 labelling indices according to clinico-pathological variables in canine mammary tumours: a multivariate study. Anticancer Res 26:1821–1826.
- De Matos AJF, Lopes C, Faustino AMR, Carvalheira J, Rutteman GR, Gärtner F. (2007). E-cadherin, β-catenin, invasion and lymph node metastases in canine malignant mammary tumours. APMIS 115:327-334.
- Destexhe E, Bicker E, Coignoul F. (1993a). Image analysis evaluation of ploidy, S-phase fraction and nuclear area in canine mammary tumours. J Comp Pathol 113:205-216.
- Destexhe E, Lespagnard L, Degeyter M, Heymann R, Coignoul F. (1993b). Immunohistochemical identification of myoepithelial, epithelial, and connective tissue cells in canine mammary tumors. Vet Pathol 30:146-154.
- Di Como CJ, Urist MJ, Babayan I, Drobnjak M, Hedvat CV, Teruya-Feldstein J, Pohar K, Hoos A, Cordon-Cardo C. (2002). p63 expression profiles in human normal and tumor tissues. Clin Cancer Res 8:494-501.
- Dick JE. (2003). Breast cancer stem cells revealed. Proc Natl Acad Sci USA 100:3547-3549.

- DiRenzo J, Signoretti S, Nakamura N, Rivera-Gonzalez R, Sellers W, Loda M, Brown M. (2002). Growth factor requirements and basal phenotype of an immortalized mammary epithelial cell line. Cancer Res 62:89-98.
- Donehower LA, Harvey M, Slagle BL, McArthur MJ, Montgomery CA, Jr Butel JS, Bradley A. (1992). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. Nature 356:215-221.
- Donnay I, Rauis J, Wouters-Ballman P, Devleeschouwer N, Leclercq G, Verstegen JP. (1993). Receptors for oestrogen, progesterone and epidermal growth factor in normal and tumorous canine mammary tissues. J Repr Fertil Suppl 47:501-512.
- Donnay I, Devleeschouwer N, Wouters-Ballman P, Leclercq G, Verstegen J. (1996). Relationship between receptors for epidermal growth factor and steroid hormones in normal, dysplastic and neoplastic canine mammary tissues. Res Vet Sci 60:251-254.
- Dontu G, El-Ashry D, Wicha MS. (2004). Breast cancer, stem/progenitor cells and the estrogen receptor. Trends Endocrinol Metab 15:193-197.
- Durchow M, Gerdes J, Schluter C. (1994). The proliferation-associated Ki-67 protein: definition in molecular terms. Cell Proliferation 27:235-242.
- Dutra AP, Granja NVM, Schmitt FC, Cassali GD. (2004). c-erbB-2 expression and nuclear pleomorphism in canine mammary tumors. Braz J Med Biol Res 37:1673-1681.
- Elston CW, Ellis IO (1998) Assessment of histological grade. In Systemic Pathology The Breast, Ed. CW Elston and IO Ellis. 3rd Edition, vol 13, Churchill and Livingstone, London, pp. 365–384.
- Erlandson RA, Cardon-Cardo C, Higgins PJ. (1984). Histogenesis of benign pleomorphic adenoma (mixed tumor) of the major salivary glands. An ultrastructural and immunohistochemical study. Am J Surg Pathol 8:803-820.
- Espinosa de Los Monteros A, Millán MY, Ordás J, Carrasco L, Reymundo C, Martín de Las Mulas J. (2002). Immunolocalization of the smooth muscle-specific protein calponin in complex and mixed tumors of the mammary gland of the dog: assessment of the morphogenetic role of the myoepithelium. Vet Pathol 39: 247-256.
- Ferguson HR. (1985). Canine mammary gland tumors. Vet Clin North Am Small Anim Pract 15:501-511.
- Fox SB, Smith K, Hollyer J, Greenall M, Hastrich D, Harris AL. (1994). The epidermal growth factor receptor as a prognostic marker: results of 370 patients and review of 3009 patients. Breast Cancer Res Treat 29:41–49.
- Fowler EH, Wilson GP, Koester A. (1974). Biologic behaviour of canine mammary neoplasms based on a histogenetic classification. Vet Pathol 11: 212-229.

- Funakoshi Y, Nakayama H, Uetsuka K, Nishimura R, Sasaki N, Doi K. (2000). Cellular proliferative and telomerase activity in canine mammary gland tumors. Vet Pathol 37:177-183.
- Gama A, Paredes J, Milanezi MF, Reis-Filho JS, Gärtner F, Schmitt FC. (2002). P-cadherin expression in canine lactating mammary gland. J Cell Biochem 86:420-421.
- Gama A, Alves A, Gärtner F, Schmitt F. (2003). P63: a novel myoepithelial cell marker in canine mammary tissues. Vet Pathol 40:412-420.
- Gama A, Paredes J, Albergaria A, Gärtner F, Schmitt F. (2004). P-cadherin expression in canine mammary tissues. J Comp Pathol 130:13-20.
- Gamallo C, Moreno-Bueno G, Sarrio D, Calero F, Hardisson D, Palacios J. (2001). The prognostic significance of P-cadherin in infiltrating ductal breast carcinoma. Mod Pathol 14:650-654.
- Gärtner F, Geraldes M, Gassali G, Rema A, Schmitt F (1999) DNA measurement and immunohistochemical characterization of epithelial and mesenchymal cells in canine mixed mammary tumours: putative evidence for a common histogenesis. Vet J 158:39-47.
- Genelhu MCLS, Cardoso SV, Gobbi H, Cassali GD. (2007). A comparative study between mixed-type tumours from human salivary and canine mammary glands. BMC Cancer 7:218
- Geraldes M, Gärtner F, Schmitt F. (2000). Immunohistochemical study of hormonal receptors and cell proliferation in normal canine mammary glands and spontaneous mammary tumours. Vet Rec 146:403-406.
- Gilbertson SR, Kurzman ID, Zachrau RE, Hurvitz AI, Black MM. (1983). Canine mammary epithelial neoplasms: biologic implications of morphologic characteristics assessed in 232 dogs. Vet Pathol 20:127-142.
- Gofuku J, Shiozaki H, Tsujinaka T, Inoue M, Tamura S, Doki Y, Matsui S, Tsukita S, Kikkawa N, Monden M. (1999). Expression of E-cadherin and alpha-catenin in patients with colorectal carcinoma. Correlation with cancer invasion and metastasis. Am J Clin Pathol 111:29-37.
- Gould Rothberg BE, Bracken MB. (2006). E-cadherin immunohistochemical expression as a prognostic factor in infiltrating ductal carcinoma of the breast: a systematic review and meta-analysis. Breast Cancer Res Treat 100:139-148.
- Graham JC, O'Keefe DA, Gelberg HB. (1999). Immunohistochemical assay for detecting estrogen receptors in canine mammary tumors. Am J Vet Res 60:627-630.
- Griffey SM, Madewell BR, Dairkee SH, Hunt JE, Naydan DK, Higgins RJ. (1993). Immunohistochemical reactivity of basal and luminal epithelium-specific cytokeratin

- antibodies within normal and neoplastic canine mammary glands. Vet Pathol 30:155-161.
- Gu X, Coates PJ, Boldrup L, Nylander K. (2008). P63 contributes to cell invasion and migration in squamous cell carcinoma of the head and neck. Cancer Letters 263:26-34.
- Gumbiner BM. (1996). Cell adhesion: the molecular basis of tissue architecture and morphogenesis. Cell 84:345-357.
- Gusterson BA, Ross DT, Heath VJ, Stein T. (2005). Basal cytokeratins and their relationship to the cellular origin and functional classification of breast cancer. Breast Cancer Res 7:143-148.
- Hagiwara K, McMenamin MG, Miura K, Harris CC. (1999). Mutational analysis of the p63/p73L/p51/p40/CUSP/KET gene in human cancer cell lines using intronic primers. Cancer Res 59:4165-4169.
- Hampe JF, Misdorp W. (1974). Tumors and dysplasias of the mammary gland. Bull WHO 50:11-133.
- Han AC, Soler AP, Knudsen KA, Salazar H. (1999). Distinct cadherin profiles in special variant carcinomas and other tumors of the breast. Hum Pathol 30:1035-1039.
- Hanna W, Kahn HJ, Trudeau M. (1999). Evaluation of HER-2/neu (erbB-2) status in breast cancer: from bench to bedside. Mod Pathol 12:827–834.
- Heimann R, Lan F, McBride R, Hellman S. (2000). Separating favorable from unfavorable prognostic markers in breast cancer: the role of E-cadherin. Cancer Res 60:298-304.
- Hellmén E. (1996). The pathogenesis of canine mammary tumors. Cancer J 9:282-286.
- Hellmén E. (2005). Complex mammary tumours in the female dog: a review. J Dairy Res 72:90-97.
- Hellmén E, Bergstrom R, Holmberg L, Spangberg I-B, Hansson K, Lindgren A. (1993). Prognostic factors in canine mammary tumors: a multivariate study of 202 consecutive cases. Vet Pathol 30:20-27.
- Hellmén E, Lindgren A. (1989). The expression of intermediate filaments in canine mammary glands and their tumors. Vet Pathol 26:420-428.
- Hellmén E, Moller M, Blankenstein MA, Andersson L, Westermark B. (2000). Expression of different phenotypes in cell lines from canine mammary spindle cell tumours and osteosarcomas indicating a pluripotent mammary stem cell origin. Breast Cancer Res Treat 61:197-210.
- Hibi K, Trink B, Patturajan M, Westra WH, Caballero OL, Hill DE, Ratovitski EA, Jen J, Sidransky D. (2000). AIS is an oncogene amplified in squamous cell carcinoma. Proc Natl Acad Sci USA 97:5462-5467.

- Hirai Y, Nose A, Kobayashi S, Takeichi M. (1989). Expression and role of E- and P-cadherin adhesion molecules in embryonic histogenesis. I. Lung epithelial morphogenesis. Development 105:263-270.
- Hoang MP, Sahin AA, Ordonez NG, Sneige N. (2000). HER-2/neu gene amplification compared with HER-2/neu protein overexpression and interobserver reproducibility in invasive breast carcinoma. Am J Clin Pathol 113: 852–859
- Holbro T, Hynes NE. (2004). ErbB receptors: directing key signalling networks throughout life. Annu Rev Pharmacol Toxicol 44:195-217.
- Hsu W-L, Huang H-M, Liao J-W, Wong M-L, Chang S-C. (2007). Increased survival in dogs with malignant mammary tumours overexpressing HER-2 protein and detection of a silent single nucleotide polymorphism in the canine HER-2 gene. Vet J Doi:10.1016/j.tvjl.2007.10.013
- Hynes NE, Lane HA. (2005). ERBB receptors and cancer: the complexity of targeted inhibitors. Nature Rev Cancer 5:341-354.
- Jimenez RE, Wallis T, Tabasczka P, Visscher DW. (2000). Determination of HER-2/neu status in breast carcinoma: comparative analysis of immunohistochemistry and fluorescent *in situ* hybridization. Modern Pathol 13:37–45.
- Jones C, Nonni AV, Fulford L, Merrett S, Chaqqar R, Eusebi V, Lakhani SR. (2001). CGH analysis of ductal carcinoma of the breast with basaloid/myoepithelial cell differentiation. Br J Cancer 85:422-427.
- Kaelin WGJ. (1999). The emerging p53 gene family. J Natl Cancer Inst 91:594-598.
- Kalirai H, Clarke RB. (2006). Human breast epithelial stem cells and their regulation. J Pathol 208:7-16.
- Karayannopoulou M, Kaldrymidou E, Constantinidis TC, Dessiris A. (2005). Histological grading and prognosis in dogs with mammary carcinomas: application of a human grading method. J Comp Pathol 133:246–252
- Kersting C, Kuijper A, Schmidt H, Packeisen J, Liedtke C, Tidow N, Gustmann C, Hinrichs B, Wulfing P, Tio J, Boecker W, van Diest P, Brandt B, Buerger H. (2006). Amplifications of the epidermal growth factor receptor gene (egfr) are common in phyllodes tumors of the breast and are associated with tumor progression. Lab Invest 86:54–61.
- Kersting C, Tidow N, Schmidt H, Liedtke C, Neumann J, Boecker W, van Diest PJ, Brandt B, Buerger H. (2004). Gene dosage PCR and fluorescence in situ hybridization reveal low frequency of EGFR amplifications despite protein overexpression in invasive breast carcinoma. Lab Invest 84:582-587.

- Klijn JG, Berns PM, Schmitz PI, Foekens JA. (1992). The clinical significance of epidermal growth factor receptor (EGF-R) in human breast cancer: a review on 5232 patients. Endocr Rev 13:3–17.
- Knudsen KA, Frankowski C, Johnson KR, Wheelock MJ. (1998). A role for cadherins in cellular signaling and differentiation. J Cell Biochem Suppl 30-31:168-176.
- Knudsen KA, Wheelock MJ. (2005). Cadherins and the mammary gland. J Cell Biochem 95:488-496.
- Korsching, E. Packeisen J, Agelopoulos K, Eisenacher M, Voss R, Isola J, van Diest PJ, Brandt B, Boecker W, Buerger H. (2002). Cytogenetic alterations and cytokeratin expression patterns in breast cancer: integrating a new model of breast differentiation into cytogenetic pathways of breast carcinogenesis. Lab Invest 82:1525-1533.
- Koster MI, Kim S, Mills AA, DeMayo FJ, Roop DR. (2004). p63 is the molecular switch for initiation of an epithelial stratification program. Genes & Dev. 18:126–131.
- Koster MI, Lu S-L, White LD, Wang X-J, Roop DR. (2006). Reactivation of developmentally expressed p63 isoforms predisposes to tumor development and progression. Cancer Res 66:3981-3986.
- Kovacs A, Dhillon J, Walker RA. (2003). Expression of P-cadherin, but not E-cadherin or N-cadherin, relates to pathological and functional differentiation of breast carcinomas. Mol Pathol 56:318-322.
- Kovacs A, Walker RA. (2003). P-cadherin as a marker in the differential diagnosis of breast lesions. J Clin Pathol 56:139-141.
- Kumar R, Wang R-A. (2002). Protein Kinases in Mammary Gland Development and Cancer. Microsc Res Tech 59:49–57.
- Laakso M, Loman N, Borg A, Isola J. (2005). Cytokeratin 5/14-positive breast cancer: true basal phenotype confined to BRCA1 tumors. Mod Pathol 18:1321-1328.
- Lakhani SR, O'Hare MJ. (2001). The mammary myoepithelial cell- Cinderella or ugly sister? Breast Cancer Res 3:1-4.
- Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS. (1994). Phosphorylation of the human estrogen receptor. Identification of hormone-regulated sites and examination of their influence on transcriptional activity. J Biol Chem 269:4458-4466.
- Lee AV, Cui X, Oesterreich S. (2001). Cross-talk among estrogen receptor, epidermal growth factor and insulin-like growth factor signalling in breast cancer. Clin Cancer Res 7:4429s–4435s.
- Lee CH, Kim WH, Lim JH, Kang MS, Kim DY, Kweon OK. (2004). Mutation and overexpression of p53 as a prognostic factor in canine mammary tumors. J Vet Sci 5:63–69.

- Leibl S, Gogg-Kammerer M, Sommersacher A, Denk H, Moinfar F. (2005). Metaplastic breast carcinomas: are they of myoepithelial differentiation?: immunohistochemical profile of the sarcomatoid subtype using novel myoepithelial markers. Am J Surg Pathol 29:347-353.
- Little NA, Jochemsen AG. (2002). p63. Int J Biochem Cell Biol 34: 6-9.
- Livasy CA, Karaca G, Nanda R, Tretiakova MS, Olopade OI, Moore DT, Perou CM. (2006). Phenotypic evaluation of the basal-like subtype of invasive breast carcinoma. Mod Pathol 19:264-271.
- Loar AS. (1989) Tumors of the genital system and mammary glands. In Textbook of Veterinary Internal Medicine: diseases of the dog and cat. Ed. SJ Ettinger. 3rd Edition. WB Saunders Company, Philadelphia, vol. 2, pp. 1814-1825.
- Lohr CV, Teifke JP, Failing K, Weiss E. (1997). Characterization of the proliferation state in canine mammary tumors by the standardized AgNOR method with postfixation and immunohistologic detection of Ki-67 and PCNA. Vet Pathol 34:212-221.
- Madewell BR, Theilen GH. (1987). Tumors of the urogenital tract. In Veterinary Cancer Medicine, Ed. GH Theilen and BR Madeweel. 2nd Edition. Lea & Febiger, Philadelphia, pp. 583.
- Madhavan M, Srinivas P, Abraham E, Ahmed I, Mathew A, Vijayalekshmi NR, Balaram P. (2001). Cadherins as predictive markers of nodal metastasis in breast cancer. Mod Pathol 14:423-427.
- Malzahn K, Mitze M,Thoenes M, Moll R. (1998). Biological and prognostic significance of stratified epithelial cytokeratins in infiltrating ductal breast carcinomas. Virchows Arch 433:119-129.
- Martin PM, Cotard M, Mialot JP, André F, Raynaud JP. (1984). Animal models for hormone dependent human breast cancer. Relationship between steroid receptor profiles in canine and feline mammary tumours and survival rate. Cancer Chemother Pharmacol 12:13-17.
- Martin de las Mulas J, Ordás J, Millán Y, Fernández-Soria V, Ramón y Cajal S. (2003). Oncogene HER-2 in canine mammary gland carcinomas: an immunohistochemical and chromogenic in situ hybridization study. Breast Cancer Res Treat 80:363–367.
- Martin de las Mulas J, Millán Y, Dios R. (2005). A prospective analysis of immunohistochemically determined Estrogen Receptor α and Progesterone Receptor Expression and host and tumor factors as predictors of disease-free period in mammary tumors of the dog. Vet Pathol 42:200–212.
- Massion PP, Taflan PM, Jamshedur Rahman SM, Yildiz P, Shyr Y, Edgerton ME, Westfall MD, Roberts JR, Pietenpol JA, Carbone DP, Gonzalez AL. (2003). Significance of p63

- amplification and overexpression in lung cancer development and prognosis. Cancer Res 63:7113-7121.
- Matos AJF, Lopes C, Carvalheira J, Santos M, Rutteman GR, Gärtner F. (2006). E-cadherin expression in canine malignant mammary tumours: relationship to other clinicopathological variables. J Comp Pathol 134:182-189.
- Matos I, Dufloth R, Alvarenga M, Zeferino LC, Schmitt F. (2005). p63, cytokeratin 5, and P-cadherin: three molecular markers to distinguish basal phenotype in breast carcinomas. Virchows Arch 447:688-694.
- Matsuyama S, Nakamura M, Yonezawa K, Shimada T, Ohashi F, Takamori Y, Kubo K. (2001). Expression patterns of the erbB subfamily mRNA in canine benign and malignant mammary tumours. J Vet Med Sci 63:949-954.
- Mialot JP, André F, Martin PM, Cotard DM, Raynaud JP. (1982). Etude de récepteurs des hormones stéroides dans les tumeurs mammaires de la chienne I. Mise en évidence, caracterisation et relation avec le type histologique. Rec Vet Med 158:215–221.
- Millanta F, Calandrella M, Bari G, Niccolini M, Vannozzi I, Poli A. (2005). Comparison of steroid receptor expression in normal, dysplastic, and neoplastic canine and feline mammary tissues. Res Vet Sci 79:225-232.
- Mills AA, Zheng B, Wang XJ, Vogel H, Roop DR, Bradley A. (1999). p63 is a p53 homologue required for limb and epidermal morphogenesis. Nature 398:708-713.
- Misdorp W. (1988). Canine mammary tumours: protective effect of late ovariectomy and stimulating effect of progestins. Vet Q 10:26-33.
- Misdorp W. (2002). Tumours of the mammary gland. In Tumors in Domestic Animals, DJ Meuten Ed., 4th Edition, Iowa State Press, Blackwell Publishing Company, pp. 575-606.
- Misdorp W, Else RW, Hellmén E, Lipscomb TP. (1999). Histological classification of mammary tumors of the dog and the cat. 2nd series. Vol VII. Washington DC, Armed Forces Institute of Pathology, American Registry of Pathology and the World Health Organization Collaborating Center for Worldwide reference on Comparative Oncology, pp. 1-59.
- Misdorp W, Hart AA. (1976). Prognostic factors in canine mammary cancer. J Natl Cancer Inst 56:779-786.
- Moe L. (2001). Population-based incidence of mammary tumours in some dog breeds. J Repr Fertil Suppl 57:439–443.
- Mol JA, Lantinga-van Leeuwen IS, van Garderen E, Selman PJ, Oosterlaken-Dijksterhuis MA, Schalken JA, Rijnberk A. (1999). Mammary growth hormone and tumorigenesis lessons from the dog. Vet Q 21:111-115.

- Moll R, Divo M, Langbein L. (2008). The human keratins: biology and pathology. Histochem Cell Biol 129:705-733.
- Moller P, Mechtersheimer G, Kaufmann M, Moldenhauer G, Momburg F, Mattfeldt T, Otto HF. (1989). Expression of epidermal growth factor receptor in benign and malignant primary tumours of the breast. Virchows Arch A: Pathol Anat Histopathol 414:157-164.
- Monlux AW, Roszel JF, MacVean DW, Palmer TW. (1977). Classification of epithelial canine mammary tumors in a defined population. Vet Pathol 14:194-217.
- Morris JS, Dobson JM, Bostock DE, O'Farrell E. (1998). Effect of ovariohysterectomy in bitches with mammary neoplasms. Vet Rec 142:656-658.
- Moulton JE (1990). Tumors of the mammary gland. In Tumors in domestic animals. Ed. JE Moulton, 3rd Edition, California Press, Berkeley, California, pp. 518-553.
- Murphy LC, Watson P. (2002). Steroid receptors in human breast carcinogenesis and breast cancer progression. Biomed Pharmacother 56:65–77.
- Murua Escobar H, Becker K, Bullerdiek J, Nolte I. (2001). The canine ERBB2 gene maps to a chromosome region frequently affected by aberrations in tumors of the dog (*Canis familiaris*). Cytogenet Cell Genet 94:194–195.
- Nerurkar VR, Chitale AR, Jalnakurpar BV, Naik SN, Lalitha VS. (1989). Comparative pathology of canine mammary tumours. J Comp Pathol 101: 389-397.
- Nerurkar VR, Seshadri R, Mulherkar R, Ishwad CS, Lalitha VS, Naik SN. (1987). Receptors for epidermal growth factor and estradiol in canine mammary tumors. Int J Cancer 40:230-232.
- Nicholson RI, Gee JM, Harper ME. (2001). EGFR and cancer prognosis. Eur J Cancer 37(Suppl 4):9-15.
- Nicholson S, Wright C, Sainsbury JR, Halcrow P, Kelly P, Angus B, Farndon JR, Harris AL. (1990). Epidermal growth factor receptor (EGFr) as a marker for poor prognosis in node-negative breast cancer patients: neu and tamoxifen failure. J Steroid Biochem Mol Biol 37:811-814.
- Nielsen TO, Hsu FD, Jensen K, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslen LA, Regaz J, Gown AM, Gilks CB, van de Rijn M, Perou CM. (2004). Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. Clin Cancer Res 10:5367–5374.
- Nieto A, Peña L, Perez-Alenza MD, Sanchez MA, Flores JM, Castaño M. (2000). Immunohistologic detection of estrogen receptor alpha in canine mammary tumors: clinical and pathologic associations and prognostic significance. Vet Pathol 37:239–247.

- Nose A, Takeichi M. (1986). A novel cadherin cell adhesion molecule: its expression patterns associated with implantation and organogenesis of mouse embryos. J Cell Biol 103:2649-2658.
- Nose A, Nagafuchi A, Takeichi M. (1987). Isolation of placental cadherin cDNA: identification of a novel gene family of cell-cell adhesion molecules. EMBO J 6:3655-3661.
- Nowak M, Madej JA, Dziêgiel P. (2007). Expression of E-cadherin, β-catenin and Ki-67 antigen and their reciprocal relationships in mammary adenocarcinomas in bitches. Folia Histochem Cytobiol 45: 233-238.
- Nylander K, Vojtesek B, Nenutil R, Lindgren B, Roos G, Zhanxiang W, Sjostrom B, Dahlqvist A, Coates PJ. (2002). Differential expression of p63 isoforms in normal tissues and neoplastic cells. J Pathol 198:417-427.
- Oka H, Shiozaki H, Kobayashi K, Inoue M, Tahara H, Kobayashi T, Takatsuka Y, Matsuyoshi N, Hirano S, Takeichi M. (1993). Expression of E-cadherin cell adhesion molecules in human breast cancer tissues and its relationship to metastasis. Cancer Res 53:1696-1701.
- Osada M, Ohba M, Kawahara C, Ishioka C, Kanamaru R, Katoh I, Ikawa Y, Nimura Y, Nakagawara A, Obinata M, Ikawa S. (1998). Cloning and functional analysis of human p51, which structurally and functionally resembles p53. Nat Med 4:839-843.
- Ozawa M, Baribault H, Kemler R. (1989). The cytoplasmic domain of the cell adhesion molecule uvomorulin associates with three independent proteins structurally related in different species. EMBO J 8:1711-1717.
- Palacios J, Benito N, Pizarro A, Suarez A, Espada J, Cano A, Gamallo C. (1995). Anomalous expression of P-cadherin in breast carcinoma. Correlation with E-cadherin expression and pathological features. Am J Pathol 146:605-612.
- Palmer TE, Monlux AW. (1979). Acid mucopolysacharides in mammary tumors of dog. Vet Pathol 16:493-509.
- Paredes J, Milanezi F, Reis-Filho JS, Leitao D, Athanazio D, Schmitt F. (2002a). Aberrant P-cadherin expression: is it associated with estrogen-independent growth in breast cancer? Pathol Res Pract 198:795-801.
- Paredes J, Milanezi F, Reis-Filho JS, Leitao D, Athanazio D, Schmitt FC. (2002b). Correlation between P-cadherin and estrogen receptor expression in breast cancer. J Br Patol Med Lab 38:307-313.
- Paredes J, Milanezi F, Viegas L, Amendoeira I, Schmitt F. (2002c). P-cadherin expression is associated with high-grade ductal carcinoma in situ of the breast. Virchows Archiv 440:16-21.

- Paredes J, Stove C, Stove V, Milanezi F, van Marck V, Derycke L, Mareel M, Bracke M, Schmitt F. (2004). P-cadherin is up-regulated by the antiestrogen ICI 182,780 and promotes invasion of human breast cancer cells. Cancer Res 64:8309-8317.
- Paredes J, Albergaria A, Oliveira J, Jerónimo C, Milanezi F, Schmitt F. (2005). P-cadherin overexpression is an indicator of clinical outcome in invasive breast carcinomas and is associated with CDH3 promoter hypomethylation. Clin Cancer Res 11:5869-5877.
- Paredes J, Correia AL, Ribeiro AS, Albergaria A, Milanezi F, Schmitt FC. (2007). P-cadherin expression in breast cancer: a review. Breast Cancer Res 9:214.
- Park BJ, Lee SJ, Kim JI, Lee SJ, Lee CH, Chang SG, Park JH, Chi SG. (2000). Frequent alteration of p63 expression in human primary bladder carcinomas. Cancer Res 60:3370-3374.
- Park K, Han S, Shin E, Kim HJ, Kim JY. (2007). EGFR gene and protein expression in breast cancers. Eur J Surg Oncol 33:956-960.
- Parodi AL, Mialot JP, Martin PM. (1984). Canine and feline mammary cancers as animal models for hormone-dependent human breast tumors: relationships between steroid receptor profiles and survival rates. Prognosis Cancer Res Ther 31:357–365.
- Parsa R, Yang A, McKeon F, Green H. (1999). Association of p63 with proliferative potential in normal and neoplastic human keratinocytes. J Invest Dermatol 113:1099-1105.
- Payne SJL, Bowen RL, Jones JL, Wells CA. (2008). Predictive markers in breast cancer the present. Histopathology 52:82–90.
- Pearce ST, Jordan VC. (2004). The biological role of estrogen receptors α and β in cancer. Crit Rev Oncol Hematol 50:3–22.
- Pellegrini G, Dellambra E, Golisano O, Martinelli E, Fantozzi I, Bondanza S, Ponzin D, McKeon F, De Luca M. (2001). p63 identifies keratinocyte stem cells. Proc Natl Acad Sci 98:3156-3161.
- Peña L, Nieto A, Pérez-Alenza D, Cuesta P, Castano M. (1998). Immunohistochemical detection of Ki-67 and PCNA in canine mammary tumors: relationship to clinical and pathologic variables. J Vet Diagn Invest 10:237-246.
- Peralta Soler A, Knudsen KA, Salazar H, Han AC, Keshgegian AA. (1999). P-cadherin expression in breast carcinoma indicates poor survival. Cancer 86:1263-1272.
- Perez Alenza MD, Peña L, del Castillo N, Nieto A. (2000). Factors influencing the incidence and prognosis of canine mammary tumours. J Small Anim Pract 41:287-291.
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lonning PE, Borresen-Dale AL, Brown PO, Botstein D. (2000). Molecular portraits of human breast tumours. Nature 406:747-752.

- Philibert JC, Snyder PW, Glickman N, Glickman LT, Knapp DW, Waters DJ. (2003). Influence of host factors on survival in dogs with malignant mammary gland tumours. J Vet Intern Med 17:102-106.
- Pinho SS, Matos AJ, Lopes C, Marcos NT, Carvalheira J, Reis CA, Gärtner F. (2007). Sialyl Lewis X expression in canine malignant mammary tumours: correlation with clinicopathological features and E-cadherin expression. BMC Cancer 7:124.
- Porter DA, Krop IE, Nasser S, Sgroi D, Kaelin CM, Marks JR, Riggins G, Polyak K. (2001). A SAGE (serial analysis of gene expression) view of breast tumor progression. Cancer Res 61:5697–5702.
- Prenzel N, Fischer OM, Streit S, Hart S, Ullrich A. (2001). The epidermal growth factor receptor family as a central element for cellular signal transduction and diversification. Endocr Relat Cancer 8:11-31.
- Preziosi R, Sarli G, Benazzi C, Marcato PS. (1995). Detection of proliferating cell nuclear antigen (PCNA) in canine and feline mammary tumours. J Comp Pathol 113: 301-312.
- Pulley LT. (1973). Ultrastructural and histochemical demonstration of myoepithelium in mixed tumors of the canine mammary gland. Am J Vet Res 34:1513-1522.
- Queiroga F, Lopes C. (2002). Canine mammary tumours, research on new prognostic factors. Rev Port Ciências Veterinárias 97:119-127.
- Queiroga FL, Perez-Alenza MD, Silvan G, Peña L, Lopes C, Illera JC. (2005). Cox-2 levels in canine mammary tumors, including inflammatory mammary carcinoma: clinicopathological features and prognostic significance. Anticancer Res 25:4269-4275.
- Ramalho LN, Ribeiro-Silva A, Cassali GD, Zucoloto S. (2006). The expression of p63 and cytokeratin 5 in mixed tumors of the canine mammary gland provides new insights into the histogenesis of these neoplasms. Vet Pathol 43:424-429.
- Rampaul RS, Pinder SE, Nicholson RI, Gullick WJ, Robertson JF, Ellis IO. (2005). Clinical value of epidermal growth factor receptor expression in primary breast cancer. Adv Anat Pathol 12:271–273.
- Rampaul RS, Pinder SE, Wencyk PM, Nicholson RI, Blamey RW, Robertson JF, Ellis IO. (2004). Epidermal growth factor receptor status in operable invasive breast cancer: is it of any prognostic value? Clin Cancer Res 10:2578.
- Rasbridge SA, Gillett CE, Sampson SA, Walsh FS, Millis RR. (1993). Epithelial (E-) and placental (P-) cadherin cell adhesion molecule expression in breast carcinoma. J Pathol 169:245-250.
- Ravdin PM, Green S, Dorr TM, McGuire WL, Fabian C, Pugh RP, Carter RD, Rivkin SE, Borst JR, Belt RJ *et al.* (1992). Prognostic significance of progesterone receptor levels in estrogen receptor-positive patients with metastatic breast cancer treated with

- tamoxifen: results of a prospective Southwest Oncology Group study. J Clin Oncol 10:1284–1291.
- Raymond WA, Leong AS. (1989). Vimentin-a new prognostic parameter in breast carcinoma? J Pathol 158:107-114.
- Reis AL, Carvalheira J, Schmitt FC, Gärtner F. (2003). Immunohistochemical study of Ecadherin expression in canine mammary tumours. Vet Rec 152:621-624.
- Reis-Filho JS, Milanezi F, Paredes J, Silva P, Pereira EM, Maeda SA, de Carvalho LV, Schmitt FC. (2003). Novel and classic myoepithelial/stem cell markers in metaplastic carcinomas of the breast. Appl Immunohistochem Mol Morphol 11:1-8.
- Reis-Filho JS, Milanezi F, Carvalho S, Simpson PT, Steele D, Savage K, Lambros MB, Pereira EM, Nesland JM, Lakhani SR, Schmitt FC. (2005). Metaplastic breast carcinomas exhibit EGFR, but not HER2, gene amplification and overexpression: immunohistochemical and chromogenic in situ hybridization analysis. Breast Cancer Res 7:RA1028-1035.
- Reis-Filho JS, Milanezi F, Steele D, Savage K, Simpson PT, Nesland JM, Pereira EM, Lakhani SR, Schmitt FC. (2006). Metaplastic breast carcinomas are basal-like tumours. Histopathology 49:10-21.
- Reis-Filho JS, Pinheiro C, Lambros MB, Milanezi F, Carvalho S, Savage K, Simpson PT, Jones C, Swift S, Mackay A, Reis RM, Hornick JL, Pereira EM, Baltazar F, Fletcher CD, Ashworth A, Lakhani SR, Schmitt FC. (2006). EGFR amplification and lack of activating mutations in metaplastic breast carcinomas. J Pathol 209:445–453.
- Reis-Filho JS, Schmitt FC. (2003). p63 expression in sarcomatoid/metaplastic carcinomas of the breast. Histopathology 42:94-95.
- Restucci B, Papparella S, De Vico G, Maiolino P. (1997). E cadherin expression in normal and neoplastic canine mammary gland. J Comp Pathol 116:191-202.
- Revillion F, Bonnetarre J, Peyrat JP. (1998). ErbB-2 oncogene in human breast cancer and its clinical significance. Eur J Cancer 34:791–808.
- Reynolds AB, Daniel J, McCrea PD, Wheelock MJ, Wu J, Zhang Z. (1994). Identification of a new catenin: the tyrosine kinase substrate p120cas associates with E-cadherin complexes. Mol Cell Biol 14:8333-8342.
- Ribeiro-Silva A, Zambelli Ramalho LN, Britto Garcia S, Zucoloto S. (2003a). The relationship between p63 and p53 expression in normal and neoplastic breast tissue. Arch Pathol Lab Med 127:336–340.
- Ribeiro-Silva A, Zamzelli Ramalho LN, Garcia SB, Zucoloto S. (2003b). Is p63 reliable in detecting microinvasion in ductal carcinoma in situ of the breast? Pathol Oncol Res 9:20-23.

- Rodo A, Malicka E. (2008). E-cadherin immunohistochemical expression in mammary gland neoplasms in bitches. Po J Vet Sci 11:47-54.
- Romond EH, Perez EA, Bryant J, Suman VJ, Geyer CEJr, Davidson NE, Tan-Chiu E, Martino S, Paik S, Kaufman PA, Swain SM, Pisansky TM, Fehrenbacher L, Kutteh LA, Vogel VG, Visscher DW, Yothers G, Jenkens RB, Brown AM, Dakhil SR, Mamounas EP, Lingle WL, Klein PM, Ingle JN, Wolmark N. (2005). Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer. N Engl J Med 353:1673–1684.
- Ross DW. (1998). Introduction to Oncogenes and Molecular Cancer Medicine. Springer-Verlag, New York.
- Rungsipipat A, Tateyama S, Yamaguchi R, Uchida K, Miyoshi N, Hayashi T. (1999). Immunohistochemical analysis of c-yes and c-erbB-2 oncogene products and p53 tumor suppressor protein in canine mammary tumors. J Vet Med Sci 61:27–32.
- Rutteman GR, Cornelisse CJ, Dijkshoorn NJ, Poortman J, Misdorp W. (1988a). Flow cytometric analysis of DNA ploidy in canine mammary tumours. Cancer Res 48:3411-3417.
- Rutteman GR, Misdorp W, Blankenstein MA, van den Brom WE. (1988b). Oestrogen (ER) and progestin receptors (PR) in mammary tissue of the female dog: Different receptor profile in non-malignant and malignant states. Brit J Cancer 58:594-599.
- Rutteman GR, Foekens JA, Blankenstein MA, Vos JH, Misdorp W. (1990). EGF-receptors in non-affected and tumorous dog mammary tissues. Eur J Cancer 26:182-186.
- Rutteman GR, Withrow SJ, MacEwen EG. (2001). Tumors of the mammary gland. In: Small Animal Clinical Oncology. SJ Withrow and BR MacEwen (Ed.), WB Saunders Company, Philadelphia, pp. 455–477.
- Santini D, Ceccarelli C, Tardio ML, Taffurelli M, Marrano D. (2002). Immunocytochemical expression of epidermal growth factor receptor in myoepithelial cells of the breast. Appl Immunohistochem Mol Morphol 10:29-33.
- Sapino A, Papotti M, Sanfilippo B, Gugliotta P, Bussolati G. (1992). Tumor types derived from epithelial and myoepithelial cell lines of R3230AC rat mammary carcinoma. Cancer Res 52:1553-1560.
- Sarli G, Preziosi R, Benazzi C, Castellani G, Marcato PS. (2002). Prognostic value of histologic stage and proliferative activity in canine malignant mammary tumors. J Vet Diagn Invest 14:25–34.
- Sarli G, Preziosi R, De Tolla L, Brunetti B, Benazzi C. (2004). E-cadherin immunoreactivity in canine mammary tumors. J Vet Diagn Invest 16:542-547.

- Sartin EA, Barnes S, Kwapien RP, Wolfe LG. (1992). Estrogen and progesterone receptor status of mammary carcinomas and correlation with clinical outcome in dogs. Am J Vet Res 53:2196–2200.
- Sartin EA, Barnes S, Toivio-Kinnucan M, Wright JC, Wolfe LG. (1993). Heterogenic properties of clonal cell lines derived from canine mammary carcinomas and sensitivity to tamoxifen and doxorubicin. Anticancer Res 13:229–236.
- Sassen A, Rochon J, Wild PJ, Hartmann A, Hofstaedter F, Schwarz S, Brockhoff G. (2008). Cytogenetic analysis of HER1/EGFR, HER2, HER3, and HER4 in 278 breast cancer patients. Breast Cancer Res 10:R2.
- Schafer KA, Kelly G, Schrader R, Griffith WC, Muggenburg BA, Tierney LA, Lechner JF, Janovitz EB, Hahn FF. (1998). A canine model of familial mammary gland neoplasia. Vet Pathol 35:168-177.
- Schaller G, Fuchs I, Pritze W, Ebert A, Herbst H, Pantel K, Weitzel H, Lengyel E. (1996). Elevated keratin 18 protein expression indicates a favorable prognosis in patients with breast cancer. Clin Cancer Res 2:1879-1885.
- Schneider R, Dorn CR, Taylor DON. (1969). Factors influencing canine mammary cancer development and postsurgical survival. J Natl Cancer Inst 43:1249-1261.
- Schweizer J, Bowden PE, Coulombe PA, Langbein L, Lane EB, Magin TM, Maltais L, Omary MB, Parry DA, Rogers MA, Wright MW. (2006). New consensus nomenclature for mammalian keratins. J Cell Biol 174:169-174.
- Selman PJ, Mol JA, Rutteman GR, van Garderen E, Rijnberk A. (1994). Progestin-induced growth hormone excess in the dog originates in the mammary gland. Endocrinology 134:287-292.
- Shimoyama Y, Hirohashi S. (1991). Expression of E- and P-cadherin in gastric carcinomas. Cancer Res 51:2185-2192.
- Shimoyama Y, Yoshida T, Terada M, Shimosato Y, Abe O, Hirohashi S. (1989). Molecular cloning of a human Ca2+-dependent cell-cell adhesion molecule homologous to mouse placental cadherin: its low expression in human placental tissues. J Cell Biol 109:1787-1794.
- Signoretti S, Waltregny D, Dilks J, Isaac B, Lin D, Garraway L, Yang A, Montironi R, McKeon F, Loda M. (2000). p63 is a prostate basal cell marker and is required for prostate development. Am J Pathol 157:1764-1775.
- Siitonen SM, Kononen JT, Helin HJ, Rantala IS, Holli KA, Isola JJ. (1996). Reduced E-cadherin expression is associated with invasiveness and unfavorable prognosis in breast cancer. Am J Clin Pathol 105:394-402.

- Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. (1987). Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. Science 235:177–182.
- Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, Fleming T, Eiermann W, Wolter J, Pegram M, Baselga J, Norton L. (2001). Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. N Engl J Med 344:783–792.
- Soler AP, Russo J, Russo IH, Knudsen KA. (2002). Soluble fragment of P-cadherin adhesion protein found in human milk. J Cell Biochem 85:180-184.
- Sorenmo K, Shofer FS, Goldschmidt MH. (2000). Effect of spaying and timing of spaying on survival of dogs with mammary carcinoma. J Vet Intern Med 14:266-270.
- Sorenmo K. (2003). Canine mammary gland tumors. Vet Clin North Am 33: 573-596.
- Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, Rijn MV, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Lonning PE, Borresen-Dale AL. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98:10869–10874
- Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lonning PE, Brown PO, Borresen-Dale AL, Botstein D. (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 100:8418–8423.
- Sotiriou C, Neo SY, McShane LM, Korn EL, Long PM, Jazaeri A, Martiat P, Fox SB, Harris AL, Liu ET. (2003). Breast cancer classification and prognosis based on gene expression profiles from a population-based study. Proc Natl Acad Sci. USA 100:10393–10398.
- Speirs V, Walker RA. (2007). New perspectives into the biological and clinical relevance of oestrogen receptors in the human breast. J Pathol 211:499-506.
- Stovring M, Moe L, Glattre E. (1997). A population based case-control study of canine mammary tumors and clinical use of medroxyprogesterone acetate. APMIS 105:590-596.
- Suo Z, Nesland JM. (2002). Type 1 Protein Tyrosine Kinases in Breast Carcinoma: A Review. Ultrastruct Pathol 26:125-135.
- Takei H, Iino Y, Horiguchi J, Kanoh T, Takao Y, Oyama T, Morishita Y. (1995). Immunohistochemical analysis of cytokeratin #8 as a prognostic factor in invasive breast carcinoma. Anticancer Res 15:1101-1105.
- Takeichi M. (1991). Cadherin cell adhesion receptors as a morphogenetic regulator. Science 251:1451-1455.

- Takeichi M. (1993). Cadherins in cancer: implications for invasion and metastasis. Cur Opin Cell Biol 5:806-811.
- Takeichi M. (1995). Morphogenetic roles of classic cadherins. Cur Opin Cell Biol 7:619-627.
- Tateyama S, Cotchin E. (1977). Alkaline phosphatase reaction of canine mixed mammary tumours: a light and electron microscopic study. Res Vet Sci 23:356-364.
- Tateyama S, Cotchin E. (1978). Electron microscopic observations on canine mixed mammary tumours, with special reference to cytoplasmic filamentous components. Am J Vet Res 39:1494-1501.
- Tateyama S, Uchida K, Hidaka T, Hirao M, Yamaguchi R. (2001). Expression of bone morphogenetic protein-6 (BMP-6) in myoepithelial cells in canine mammary gland tumors. Vet Pathol 38:703-709.
- Thompson L, Chang B, Barsky SH. (1996). Monoclonal origins of malignant mixed tumors (carcinosarcomas). Evidence for a divergent histogenesis. Am J Surg Pathol 20:277-285.
- Toi M, Tominaga T, Osaki A, Toge T. (1994). Role of epidermal growth factor receptor expression in primary breast cancer: results of a biochemical study and an immunocytochemical study. Breast Cancer Res Treat 29:51-58.
- Tovey SM, Witton CJ, Bartlett JMS, Stanton PD, Reeves JR, Cooke TG. (2004). Outcome and human epidermal growth factor receptor (HER) 1-4 status in invasive breast carcinomas with proliferation indices evaluated by bromodeoxyuridine labelling. Breast Cancer Res 6:R246–R251.
- Tsuda H, Takarabe T, Hasegawa F, Fukutomi T, Hirohashi S. (2000). Large, central acellular zones indicating myoepithelial tumor differentiation in high-grade invasive ductal carcinomas as markers of predisposition to lung and brain metastases. Am J Surg Pathol 24:197–202.
- Tsutsui S, Ohno S, Murakami S, Hachitanda Y, Oda S. (2002). Prognostic value of epidermal growth factor receptor (EGFR) and its relationship to the estrogen receptor status in 1029 patients with breast cancer. Breast Cancer Res Treat 71:67-75.
- Turner NC, Reis-Filho JS. (2006). Basal-like breast cancer and the BRCA1 phenotype. Oncogene 25:5846–5853.
- van de Rijn M, Perou CM, Tibshirani R, Haas P, Kallioniemi O, Kononen J, Torhorst J, Sauter G, Zuber M, Kochli OR, Mross F, Dieterich H, Seitz R, Ross D, Botstein D, Brown P. (2002). Expression of cytokeratins 17 and 5 identifies a group of breast carcinomas with poor clinical outcome. Am J Pathol 161:1991–1996.

- Van Leeuwen IS, Hellmen E, Cornelisse CJ, Van der Burg B, Rutteman GR. (1996). P53 mutations in mammary tumor cell lines and corresponding tumor tissues in the dog. Anticancer Res 16:3737-3743.
- Veldhoen N, Watterson J, Brash M, Milner J. (1999). Identification of tumour-associated and germ line p53 mutations in canine mammary cancer. Br J Cancer 81:409-415.
- Veronese SM, Gambacorta M. (1992). Detection of Ki-67 proliferation rate in breast cancer-correlation with clinical and pathologic features. Am J Clin Pathol 95:30-34.
- Vlahovic G, Crawford J. (2003). Activation of tyrosine kinases in cancer. The Oncologist 8:531-538.
- Vleminckx K, Vakaet Jr L, Mareel M, Fiers W, Van Roy F. (1991). Genetic manipulation of E-cadherin expression by epithelial tumour cells reveals an invasion suppressor role. Cell 66:107-119.
- Vos JH, Van Den Ingh TSGAM, Misdorp W, Molenbeek RF, Van Mil FN, Rutteman GR, Ivanyi D, Ramaekers FCS. (1993). Immunohistochemistry with keratin, vimentin, desmin, and α-smooth muscle actin monoclonal antibodies in canine mammary gland: benign mammary tumors and duct ectasias. Vet Q 14:89-95.
- Wada H, Enomoto T, Tsujimoto M, Nomura T, Murata Y, Shroyer KR. (1998). Carcinosarcoma of the breast: molecular-biological study for analysis of histogenesis. Hum Pathol 29:1324-1328.
- Wang X, Mori I, Tang W, Nakamura M, Nakamura Y, Sato M, Sakurai T, Kakudo K. (2002). p63 expression in normal, hyperplastic and malignant breast tissues. Breast Cancer 9:216-219.
- Weber F, Fukino K, Sawada T, Williams N, Sweet K, Brena RM, Plass C, Caldes T, Mutter GL, Villalona-Calero MA, Eng C. (2005). Variability in organ-specific EGFR mutational spectra in tumour epithelium and stroma may be the biological basis for differential responses to tyrosine kinase inhibitors. Br J Cancer 92:1922-1926.
- Westfall MD, Mays DJ, Sniezek JC, Pietenpol JA. (2003). The Δ Np63 α phosphoprotein binds the p21 and 14-3-3 σ promoters in vivo and has transcriptional repressor activity that is reduced by Hay-Wells syndrome-derived mutations. Mol Cell Biol 23:2264-2276.
- Westfall MD, Pietenpol JA. (2004). p63: Molecular complexity in development and cancer. Carcinogenesis 25:857-864.
- Wijnhoven BP, Dinjens WN, Pignatelli M. (2000). E-cadherin-catenin cell-cell adhesion complex and human cancer. Br J Surg 87:992-1005.
- Williams JM, Daniel CW. (1983). Mammary ductal elongation: differentiation of myoepithelium and basal lamina during branching morphogenesis. Dev Biol 97:274-290.

- Yamagami T, Kobayashi T, Takahashi K, Sugiyama M. (1996a). Influence of ovariectomy at the time of mastectomy on the prognosis for canine malignant mammary tumors. J Small Anim Pract 37:462-464.
- Yamagami T, Kobayashi T, Takahashi K, Sugiyama M. (1996b). Prognosis for canine malignant mammary tumors based on TNM and histologic classification. J Vet Med Sci 58:1079-1083.
- Yamaguchi K, Wu L, Caballero OL, Hibi K, Trink B, Resto V, Cairns P, Okami K, Koch WM, Sidransky D, Jen J. (2000). Frequent gain of the p40/p51/p63 gene locus in primary head and neck squamous cell carcinoma. Int J Cancer 86:684-689.
- Yamaguchi Y. (2007). Microenvironmental regulation of estrogen signals in breast cancer. Breast Cancer 14:175-181.
- Yamauchi H, Stearns V, Hayes DF. (2001). When is a tumor marker ready for prime time? A case study of c-erbB-2 as a predictive factor in breast cancer. J Clin Oncol 19:2334–2356.
- Yang A., Kaghad M, Wang Y, Gillet E, Fleming MD, Dotsch V, Andrews NC, Caput D, Mc Keon F. (1998). p63, a p53 homolog at 3q27-29, encodes multiple products with transactivating, death-inducing and dominant-negative activities. Mol Cell 2:305-316.
- Yang A, Schweitzer R, Sun D, Kaghad M, Walker N, Bronson RT, Tabin C, Sharpe A, Caput D, Crum C, Mc Keon F. (1999). p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. Nature 398:714-718.
- Yang A, McKeon F. (2000). p63 and p73: p53 mimics, menaces and more. Nature Rev Mol Cell Biol 1:194-207.
- Yarden Y, Sliwkowski MX. (2001). Untangling the ErbB signalling network. Nat Rev Mol Cell Biol 2:127–137.
- Yeh I-T, Mies C. (2008). Application of immunohistochemistry to breast lesions. Arch Pathol Lab Med 132:349–358.
- Yuzbasiyan-Gurkan V, Cao Y, Venta P, Yamini B. (1999). Loss of heterogenozygosity at the BRCA1 locus in canine mammary carcinoma. Proc Vet Cancer Soc 19:7.
- Zacchetti A, van Garderen E, Teske E, Nederbragt H, Dierendonck JH, Rutteman GR. (2003). Validation of the use of proliferation markers in canine neoplastic and non-neoplastic tissues: comparison of KI-67 and proliferating cell nuclear antigen (PCNA) expression versus in vivo bromodeoxyuridine labelling by immunohistochemistry. APMIS 111:430-438.
- Zaidan Dagli ML. (2008). The search for suitable prognostic markers for canine mammary tumors: A promising outlook. Vet J 177:3-5.

- Zhuang Z, Lininger RA, Man YG, Albuquerque A, Merino MJ, Tavassoli FA. (1997). Identical clonality of both components of mammary carcinosarcoma with differential loss of heterozygosity. Mod Pathol 10:354-362.
- Zschiesche W, Schonborn I, Behrens J, Herrenknecht K, Hartveit F, Lilleng P, Birchmeier W. (1997). Expression of E-cadherin and catenins in invasive mammary carcinomas. Anticancer Res 17:561-567.
- Zuccari DAPC, Santana AE, Cury PM, Cordeiro JA. (2004). Immunocytochemical study of Ki-67 as a prognostic marker in canine mammary neoplasia. Vet Clin Pathol 33:23-28.

3. AIMS AND OUTLINE OF THE THESIS

With the purpose of better elucidate canine mammary gland tumour biopathology, we have intended to provide new insights on their histogenesis/differentiation, prognosis and molecular classification. To accomplish this goal, we defined the following specific aims:

- . To perform an extensive clinicopathological characterization of canine mammary benign and malignant tumours. To study possible associations between host and tumour characteristics and biologic behaviour of canine mammary tumours.
- . To evaluate the immunohistochemical expression of the cell adhesion molecules E-cadherin, P-cadherin and β -catenin in a series of canine malignant mammary tumours and their relation to clinicopathological parameters, proliferation and survival.
- . To evaluate the immunohistochemical expression of EGFR in a series of benign and malignant canine mammary tumours. To evaluate its expression in relation to clinicopathological parameters and survival.
- . To evaluate the immunohistochemical expression of the luminal cell marker CK 19 and basal/myoepithelial cell markers (CK5, CK14, p63, calponin, smooth muscle actin and P-cadherin) in a series of canine malignant mammary tumours. To study CK19 prognostic significance and its relationship with clinicopathological parameters and basal/myoepithelial cell markers expression.
- . To identify molecular phenotypes in a series of canine mammary carcinomas based on the application of a human classification scheme, by using a surrogate panel of immunohistochemical markers (ER, HER-2, CK5, P63 and P-cadherin). To explore the relationship of these distinct phenotypes with clinicopathological parameters and survival

Chapter II

Canine mammary gland tumours: clinical and pathological parameters as predictors of overall and disease-free survival - a univariate and multivariate analysis

Gama A, Alves A, Schmitt F (submitted)

Canine mammary gland tumours: clinical and pathological parameters as predictors of overall

and disease-free survival - a univariate and multivariate analysis

Gama A^a, Alves A^a, Schmitt F^{b,c*}

^a Department of Veterinary Sciences, CECAV, University of Trás-os-Montes and Alto Douro

(UTAD), 5001-811 Vila Real, Portugal

^bInstitute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP),

4200-465 Porto, Portugal

^c Medical Faculty, University of Porto, Porto, Portugal

* Corresponding author:

Tel.: 351-22-557 07 00; fax: 351-22-557 07 99.

E-mail address: fernando.schmitt@ipatimup.pt (F. Schmitt).

Abstract

A hundred and fifty six canine mammary tumour specimens (46 benign and 110

malignant) were clinically and histopathologically characterized. In order to investigate

the prognostic value of clinical and pathological variables, a follow-up study was

performed in 69 female dogs for a minimum period of 12 months after surgical

procedure. Univariate analysis showed that tumour size, histological type, tumour

growth, differentiation grade, stromal and lymphovascular invasion, lymph node status,

mitotic and Ki-67 labelling indices were significantly associated with overall and

disease-free survival. Skin ulceration was only associated with poorer overall survival

rate. Cox regression multivariate analysis revealed lymph node status as the only

independent prognostic factor.

Keywords: canine; mammary tumour; prognosis

Introduction

Mammary tumours are the most common neoplasias in female dogs, representing a

serious problem worldwide (Misdorp et al., 1999; Zaidan Dagli, 2008). Malignant

tumours account for up to 50% of mammary neoplasms and the search for prognostic

markers has been increasing in the last decades, in order to better estimate the individual

51

risk of unfavourable clinical outcome (Misdorp, 2002; Sorenmo, 2003; Zaidan Dagli, 2008).

Although some clinicopathological factors have been recognized by several studies as reliable prognostic factors, a number of discrepancies and controversial results still exist concerning this subject. Tumour size (Bostock, 1975; Misdorp and Hart, 1976; Yamagami et al., 1996b; Chang et al., 2005; Martin de las Mulas et al., 2005), skin ulceration (Hellmén et al., 1993; Peña et al., 1998; Queiroga and Lopes, 2002), tumour type (Misdorp and Hart, 1976; Hellmén et al., 1993; Chang et al., 2005) and grade (Karayannopoulou et al., 2005; Martin de las Mulas et al., 2005) and presence of lymph node metastasis (Hellmén et al., 1993; Yamagami et al., 1996b; Queiroga and Lopes, 2002; Philibert et al., 2003; Chang et al., 2005) have been considered as good prognosticators by many investigators. However, clinical features such as animal age and breed, ovariohysterectomy status and tumour location (Schneider et al., 1969; Misdorp and Hart, 1976; Hellmén et al., 1993; Yamagami et al., 1996a, 1996b; Peña et al., 1998; Sorenmo et al., 2000; Rutteman et al., 2001; Queiroga and Lopes, 2002; Philibert et al., 2003; Chang et al., 2005) usually generate more controversial results. In the present study, we have characterized a series of benign and malignant canine mammary tumours, both at clinical and pathological level. Clinicopathological parameters were compared between benign and malignant tumours and a survival study was performed in the malignant group, in order to investigate the possible association of these clinicopathological features and clinical outcome.

Material and methods

Tumour specimens

Canine mammary gland tumour specimens were obtained from the archives of the Histopathology Laboratories of the University of Trás-os-Montes and Alto Douro, Vila Real and from the Institute of Biomedical Science at the University of Porto. Tumour samples were surgically removed from 153 female dogs by lumpectomy or mastectomy (regional or radical) in private clinical practices (the majority from the Northern region of Portugal) and in the hospitals of the above mentioned institutions. From the available archival material obtained between 1999 and 2007, selected benign (n=46) and

malignant (n=110) mammary tumours were studied. The material had been fixed in 10% neutral buffered formalin and embedded in paraffin wax. Sections (3 μ m) were cut and stained with haematoxylin and eosin (HE) for histological examination.

Clinicopathological parameters evaluation

Clinical gathered data included animal breed, age, reproductive status (intact; ovariohysterectomized prior to tumour development; ovariohysterectomized with mastectomy), previous administration of oestrous-prevention medications and tumour characteristics (location, size, skin ulceration). Tumour size was defined as the maximum diameter and tumours were grouped according to the TNM WHO staging of canine mammary tumours (Rutteman *et al.*, 2001) in: tumours with less than 3 cm; tumours with 3-5 cm and tumours larger than 5 cm.

All tumour samples were revised and reclassified independently by three observers from haematoxylin and eosin (HE) stained sections, according to the World Health Organization (WHO) criteria for canine mammary neoplasms (Misdorp *et al.*, 1999). Other histopathological parameters evaluated included: intra-tumoural necrosis (presence vs. absence), mode of growth (expansive vs. infiltrative), characterization of inflammatory cellular infiltrates (infiltrate type and extent), stromal/lymphovascular invasion (presence vs. absence) and lymph node metastases (presence vs. absence).

Histological grade was evaluated in malignant epithelial neoplasms, according to the Nottingham method for human breast tumours (Elston and Ellis, 1998), which is based on the assessment of three morphological features: tubule formation, nuclear pleomorphism and mitotic counts. Each of these features was scored as 1, 2 or 3 to indicate whether it was present in slight, moderate or marked degree, respectively, giving a putative total of 3-9 points. Grade was allocated by an arbitrary division of the total points as follows: grade I (well differentiated), 3, 4 or 5 points; grade II (moderately differentiated), 6 or 7 points; and grade III (poorly differentiated), 8 or 9 points. Mitotic counts were assessed as the number of mitoses per 10 high power fields (40x) at the tumour periphery, by using a Nikon Labophot microscope (area=0,152 mm²).

Proliferation indices

For Ki-67 immunostaining, a monoclonal antibody was used (MIB-1, 1:50, Dakocytomation) and the immunohistochemical technique was performed according to the streptavidin-biotin-peroxidase complex (ABC) method. Briefly, tissue sections were deparaffinized, rehydrated and antigen retrieval was carried out. Slides were incubated with 0.2 mg/mL trypsin (Merck) in phosphate buffered saline (PBS) for 10 min at 37 °C prior to microwave treatment (3 x 5 min) in a 10 mM citrate buffer, pH 6.0. After cooling 20 minutes at room temperature, the sections were immersed in 3% hydrogen peroxide (H₂O₂) and distillated water during 30 minutes to block endogenous peroxidase activity. Non-specific staining was eliminated by 5-minute incubation with Ultra V Block (Lab Vision). Excess serum was removed, replaced by the primary antibody, and the slides were incubated overnight in a humid chamber at 4°C. After incubation, the slides were washed and sections were incubated with biotinylated goat anti-polyvalent (Lab Vision) for 10 minutes followed by streptavidin peroxidase for 10 min (Lab Vision). Sections were rinsed thoroughly with PBS between each step of the procedure. Subsequently, the color was developed with 3,3-diaminobenzidine tetrahydrochloride (DAB) with H₂O₂ in PBS buffer for 10 minutes. Slides were counterstained with Gill's hematoxylin, dehydrated, and mounted. Adjacent normal mammary tissues were used as internal positive controls. Negative controls were carried out by replacing the primary antibody with PBS.

Ki-67 immunostaining was nuclear and considered positive regardless of the intensity. Mitotic and Ki-67 labelling indices were determined both on benign and malignant lesions, by counting 1,000 neoplastic cells in the most mitotically active areas, at high magnification (40x), with the help of a microscopic grid (Zeiss®). Mitotic and Ki-67 indices were calculated as the percentage of tumour cells that exhibited mitotic figures or had positive staining for Ki-67, respectively.

Follow up study

After the surgical procedure, dogs presenting malignant mammary tumours were submitted to a minimum follow-up period of 12 months (range 5-74 months). Follow-up was performed by the referring surgeons and it was possible in sixty nine malignant tumour cases. The remaining cases were excluded from follow-up due to a number of

reasons: dogs died immediately after surgery, others failed clinical examinations and some ancient cases just didn't have medical records anymore. Overall survival (OS) was defined as the period between surgery and animal natural death or euthanasia due to cancer. Disease-free survival (DFS) was defined as the period of time between surgery and recurrent or metastatic disease. One dog died due to causes unrelated to the mammary tumour and it was censored at the time of death (19 months).

Statistical Analysis

For statistical analysis, categorical variable studies were performed by using the chisquare test and Fisher's exact test (two-sided). Continuous variables (mitotic and Ki-67 indices) were evaluated with the Mann-Whitney or Kruskall Wallis test.

In order to determine the effect of studied clinicopathological variables on prognosis, survival curves were generated by the Kaplan-Meier method and the survival rates compared using the log-rank test. To analyze the effect of proliferation on survival, mitosis and Ki-67 labelling indices were dichotomized to low (minor than the median value) and high groups (≥ the median value).

The Cox proportional hazard model for multivariate analysis was performed to determine the effects of different co-variables on overall and disease-free survival. Variables that were found to be important in the Kaplan-Meier analyses were included in the multivariate analysis. The hazard ratios were estimated with their 95% confidence interval. All statistical analyses were performed using SPSS software (Statistical Package for the Social Sciences, Chicago, USA), 11.5 version. *P*<0.05 was considered statistically significant.

Results

Clinical characterization

The overall clinical characteristics of our tumour series are displayed in Table 1.

Clinical information regarding animal age and breed was possible in 145 and 146 cases, respectively. The mean age of dogs at the time of surgical removal of tumours was 9.3 ± 2.5 years (range 2–16 years of age). For statistical purposes, age groups were established based on the mean age of dogs, with animals divided in young (≤ 9 years)

and old (>9 years) dogs. Concerning animal breed, most affected bitches were mixed breed (n=48, 33.1%), followed by Poodles (n=25, 17.2%), Cocker spaniels (n=17, 11.7%), Boxers (n=9, 6.2%) and Labrador retrievers (n=6, 4.1%). For statistical purposes, we subdivided this variable in mixed breeds (n=48), poodles (n=25), cocker spaniels (n=17) and other breeds (n=48).

Table 1. Frequencies observed in the present series for clinical parameters.

Clinical parameter	Frequencies n (%)
Age (n=146)	
≤9 years	74 (50.7%)
>9 years	72 (49.3%)
Breed (n=145)	
Mixed breed	48 (33.1%)
Poodle	25 (17.2%)
Cocker spaniel	17 (11.7%)
Others	55 (37.9%)
Ovariohysterectomy (n=98)	
No	70 (71.4%)
Yes, prior to tumour development	13 (13.3%)
Yes, performed with mastectomy	15 (15.3%)
Contraception (n=98)	
No	72 (90%)
Yes	8 (10%)
Tumour size (n=149)	
<3 cm	80 (53.7%)
3-5 cm	41 (27.5%)
>5 cm	28 (18.8%)
Tumour location (n=89)	
Cranial glands (thoracic)	8 (9%)
Medial gland (cranial abdominal)	14 (15.7%)
Caudal glands (caudal abdominal and inguinal)	50 (56.2%)
Multiple	17 (19.1%)
Skin ulceration (n=154)	
No	132 (85.7%)
Yes	22 (14.3%)

Regarding reproductive status, from 98 cases with available clinical information, 13 dogs (13.3%) have been spayed prior to tumour development, whereas in 15 (15.3%) cases ovariohysterectomy was performed with mastectomy.

Out of 80 cases, 8 dogs (10%) have been previously medicated with contraceptives. All of them developed malignant mammary tumours, but no association was found between contraception and the parameters evaluated in our study. As for tumour size, it was obtained in 149 cases. The mean maximum tumour diameter was 3.38 ± 3.24 cm, with tumours ranging from 0.4 to 18cm. With regard to location, 89 cases had available information, with most tumours located in caudal abdominal and inguinal mammary glands (n=50, 56.2%) and in the right mammary chain (n=46, 51.7%). Thoracic mammary glands were involved only in 8 (9%) cases. Seventeen (19.1%) tumours presented a multiple location, usually affecting two mammary glands (n=15). As for skin ulceration, it was found in 22 (14.3%) out of 154 cases, always associated with malignancy (Fig.1).



Fig. 1. Canine mammary gland tumour exhibiting extensive skin ulceration.

Histopathological characterization

The histopathological characteristics of our series are presented in Table 2. Mammary gland tumours under study were subdivided in benign and malignant tumours, being classified according to the WHO criteria for canine mammary tumours. Next, we will provide a description of the histological tumour types observed in our series.

Benign tumours (n=46). Benign mixed tumours (n=22) were characterized by the proliferation of both glandular (luminal and myoepithelial) and mesenchymal elements like cartilage (n=13) or cartilage and bone (n=9) (Fig. 2). Distinct myoepithelial cell morphologies were observed, from spindle- to stellate-cells, located in a suprabasal or interstitial position. Complex adenomas (n=13) were composed of a mixed proliferation of luminal epithelial cells and cells resembling myoepithelial cells (Fig. 3). Distinct myoepithelial cell morphologies were observed, similarly to the ones described for benign mixed tumours. Basaloid adenomas (n=11) were characterized by the proliferation of uniform cords or clusters of monomorphic basaloid epithelial cells. Centrally located cells showed squamous differentiation in the majority of cases (n=10) (Fig. 4).

Malignant tumours (n=110). Complex carcinomas (n=31) were characterized by the dual proliferation of luminal epithelial and myoepithelial cells. These tumours were usually solid, with myoepithelial cell proliferations admixed with a solid or tubulopapillary proliferation of luminal cells (Fig. 5). Most tumours were well circumscribed, frequently showing moderate cellular atypia and an expansive tumour growth. Solid carcinomas (n=27) were characterized by the arrangement of epithelial tumour cells in solid sheets, cords or nests, usually associated with a scant stromal component. These solid proliferations showed marked cellular pleomorfism and high number of mitotic figures (Fig. 6). Tubulopapillary carcinomas (n=19) were composed of a proliferation of cells resembling luminal epithelial cells showing a tubular and/or papillary arrangement (Fig. 7). Yet, the use of myoepithelial cell markers confirmed an associated myoepithelial cell component in 8 cases, not easily identified on routine diagnosis. In addition, 3 cases displayed more than 50% of a micropapillary pattern, which defines a micropapillary carcinoma in human breast tumour classification. These tumours were characterized by papillary cell clusters surrounded by empty lacunar spaces (Fig. 8). Papillae lacked a true fibrovascular core and were lined by polygonal cells showing intermediate to high grade nuclei. Carcinosarcomas (n=16) were composed of both epithelial and mesenchymal malignant components (Fig. 9), characterized by marked cellular pleomorfism, abundant mitotic figures and infiltrative tumour growth. Carcinoma in benign tumour cases (n=7) were characterized by a benign proliferation of

a dual epithelial cell population, which presented large areas of malignant epithelial proliferation, associated with moderate or marked cellular pleomorphism, sometimes exhibiting metaplastic changes. Spindle cell carcinomas (n=5) were characterized by the proliferation of infiltrative spindle cells usually arranged in solid epithelial patterns, with the formation of bundles and nests (Fig. 10). All of these tumours were positive for at least one myoepithelial cell marker.

Anaplastic carcinomas (n=3) were characterized by the proliferation of pleomorphic epithelial cells, presenting a highly infiltrative behaviour usually in single cells (Fig 11). These cells lacked cohesion and appeared individually dispersed through a fibrous connective tissue, sometimes in single linear cords (n=2). This particular histopathological description overlaps the characteristics assigned to human pleomorphic lobular carcinoma, a variant of classical lobular carcinoma. As for sarcomas (n=3), we identified an osteosarcoma and 2 fibrosarcomas. Osteosarcoma was characterized by the presence of polyhedral and pleomorphic neoplastic cells, accompanied by osteoid formation. Fibrosarcomas were composed of highly infiltrative spindle cell proliferations, arranged in interlacing fascicles, associated with marked cellular pleomorphism and the presence of frequent mitotic figures.

For statistical purposes, we grouped malignant tumours in three main groups, considering cell differentiation: simple carcinomas (carcinomas composed of one cell type, which included solid carcinomas, tubulopapillary "simple" carcinomas, anaplastic carcinomas and spindle cell carcinomas), complex carcinomas (carcinomas composed of epithelial and myoepithelial cell proliferation, which included the above described complex carcinomas, in addition to carcinoma in benign tumour and tubulopapillary "complex" carcinoma types), and carcinosarcomas (tumours composed of a carcinoma and a sarcoma component). Sarcoma tumour type was excluded due to the small number of cases.

The presence of intra-tumoural necrosis, usually observed as large necrotic areas (Fig. 12), was observed in 117 (75%) tumour cases, being highly associated with malignancy (105/110 cases). Concerning mode of growth, seventy nine (50.6%) cases showed an infiltrative tumour growth, always found to be associated with a malignant diagnosis.

Table 2. Frequencies observed in the present series for histopathological parameters.

Histopathological parameter	Frequencies n (%)
Histological type (n=156)	
Benign tumours	
Benign mixed tumours	22 (14.1%)
Complex adenoma	13 (8.3%)
Basaloid adenoma	11 (7.1%)
Malignant tumours	
Complex carcinoma	31 (19.9%)
Solid carcinoma	27 (17.3%)
Tubulopapillary carcinoma	19 (12.2%)
Carcinosarcoma	15 (9.6%)
Carcinoma in benign tumour	7 (4.5%)
Spindle cell carcinoma	5 (3.2%)
Anaplastic carcinoma	3 (1.9%)
Sarcoma	3 (1.9%)
Necrosis (n=156)	
Absent	39 (25%)
Present	117 (75%)
Mode of growth (n=156)	
Expansive	77 (49.4%)
Infiltrative	79 (50.6%)
Inflammatory cellular infiltrates (n=156)	
Slight/Absent	58 (37.2%)
Moderate	58 (37.2%)
Marked	40 (25.6%)
Histological grade (n=107)	
Grade I	18 (16.8%)
Grade II	34 (31.8%)
Grade III	55 (51.4%)
Stromal Invasion (n=156)	
Absent	73 (46.8%)
Present	83 (53.2%)
Lymphovascular Invasion (n=156)	
Absent	91 (58.3%)
Present	65 (41.7%)
Lymph node metastasis (n=68)	
Absent	36 (52.9%)
Present	32 (47.1%)

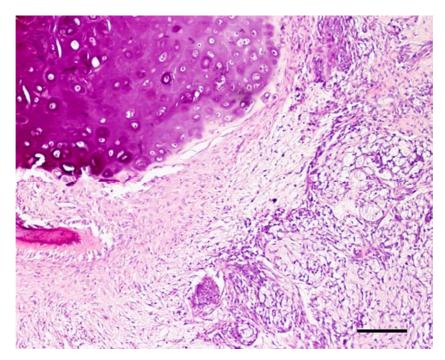


Fig. 2. Mammary gland; dog. Benign mixed tumour characterized by the proliferation of both epithelial and mesenchymal elements (cartilage and bone). HE. Bar=60μm.

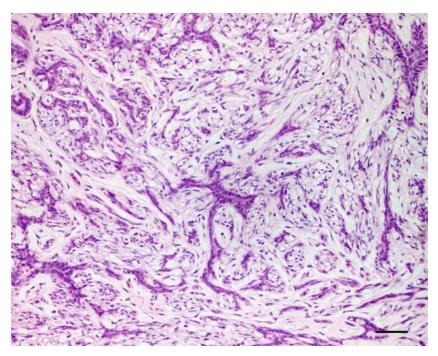


Fig. 3. Mammary gland; dog. Complex adenoma composed by the proliferation of both luminal epithelial and myoepithelial well differentiated cells. HE. Bar= $60\mu m$.

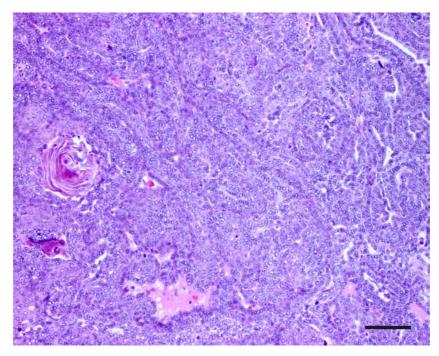


Fig. 4. Mammary gland; dog. Basaloid adenoma characterized by the proliferation of uniform cords or clusters of monomorphic basaloid epithelial cells, showing occasional squamous differentiation. HE. Bar= $60\mu m$.

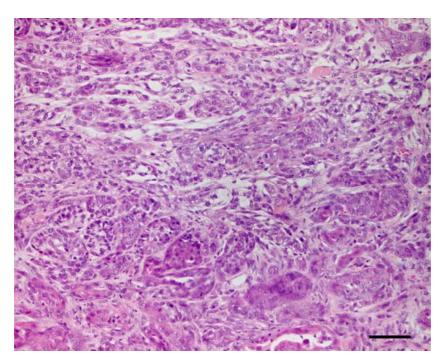


Fig. 5. Mammary gland; dog. Complex carcinoma characterized by the proliferation of both luminal epithelial and myoepithelial cells, exhibiting moderate cellular atypia HE. Bar=60μm.

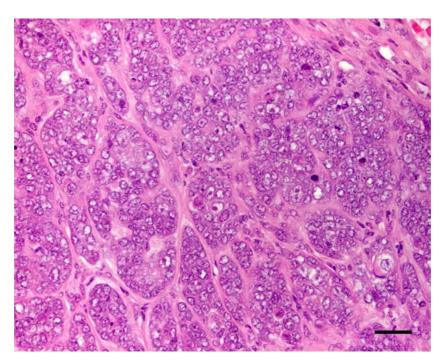


Fig. 6. Mammary gland; dog. Solid carcinoma characterized by the proliferation of epithelial cells arranged in solid nests, associated with marked cellular pleomorfism and high mitotic index. HE. Bar= $30\mu m$.

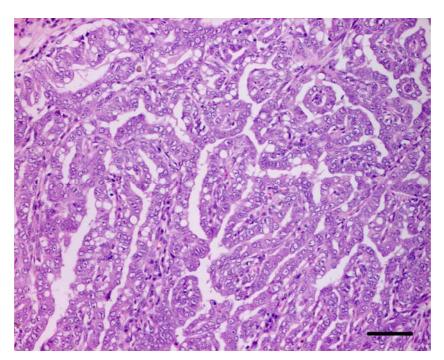


Fig. 7. Mammary gland; dog. Tubulopapillary carcinoma characterized by the proliferation of luminal epithelial cells showing a tubular and/or papillary arrangement. HE. Bar= $60\mu m$.

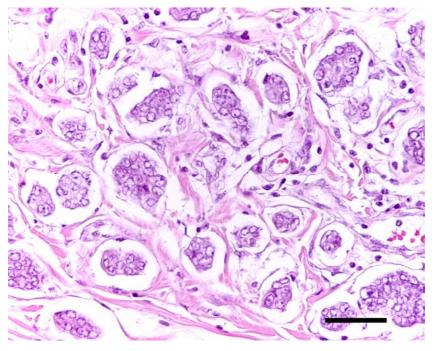


Fig. 8. Mammary gland; dog. Micropapillary carcinoma composed of small papillary cell clusters surrounded by empty lacunar spaces. HE. Bar= $40\mu m$.

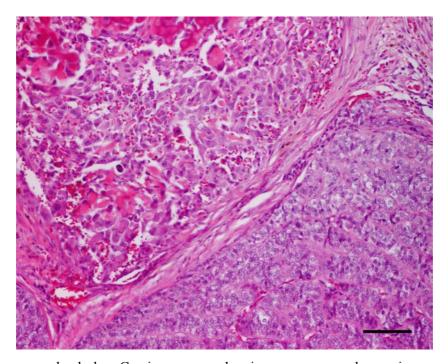


Fig. 9. Mammary gland; dog. Carcinosarcoma showing a sarcoma and a carcinoma component. HE. Bar= $60\mu m$.

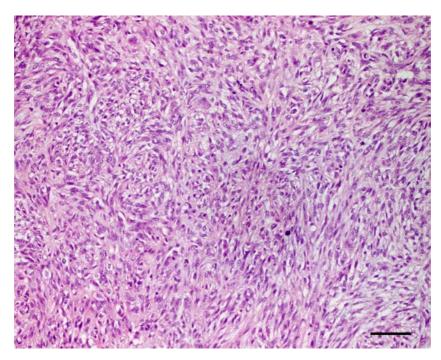


Fig. 10. Mammary gland; dog. Spindle cell carcinoma composed by spindle neoplastic cells. HE. Bar= $60\mu m$.

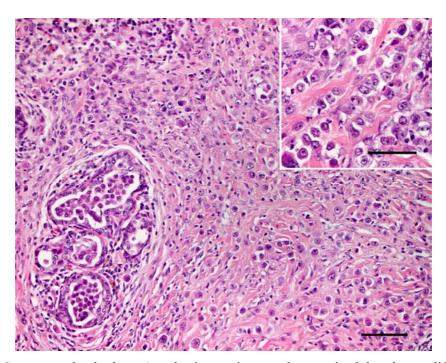


Fig. 11. Mammary gland; dog. Anaplastic carcinoma characterized by the proliferation of infiltrative non-cohesive epithelial cells. HE. Bar= $60\mu m$. Inset bar= $40 \mu m$.

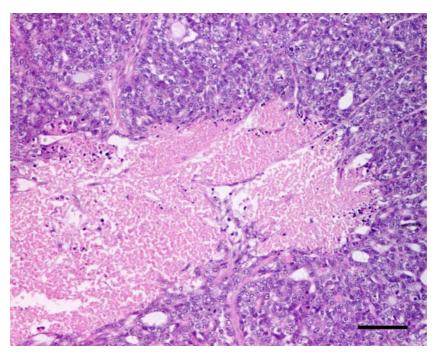


Fig. 12. Mammary gland; dog. Solid carcinoma showing the presence of intra-tumoural necrosis. HE. Bar= $60\mu m$.

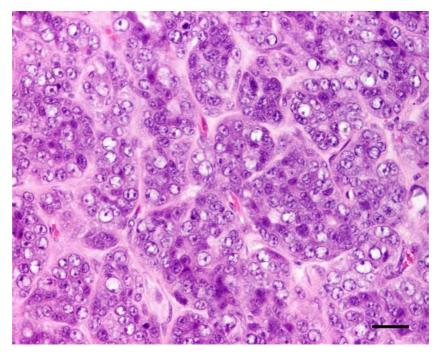


Fig. 13. Mammary gland; dog. Grade III mammary carcinoma, showing marked nuclear pleomorphism. HE. Bar=30 μ m.

As for inflammatory cellular infiltrates, most benign tumours presented a reduced or absent inflammatory cell infiltration (n=26, 56.5%), whereas malignant ones were usually associated with a moderate inflammatory response (n=43, 39%). When observed, inflammatory infiltrate was mainly characterized by the presence of lymphoid cells (mature lymphocytes and plasma cells).

According to the Nottingham method for histological grading of human breast carcinomas, canine mammary carcinomas (n=107) were classified as grade I (n=18, 16.8%), grade II (n=34, 31.8%) and grade III (n=55, 51.4%) (Fig. 13).

Eighty-three (53.2%) tumours showed stromal invasion, and 65 (41.7%) cases exhibited lymphovascular invasion, all diagnosed as malignant tumours. All carcinosarcoma, spindle cell carcinoma and anaplastic carcinoma cases showed stromal invasion. Anaplastic (n=3, 100%) and solid carcinoma (n=24, 88.9%) types showed the higher levels of vascular invasion.

In our series, we obtained lymph nodes in 68 tumour cases (Fig. 14). From these, 32 (47.1%) have revealed the presence of epithelial cancer cells by histological evaluation. Solid carcinoma (n=15, 78.9%) and anaplastic carcinoma (n=3, 100%) were the most frequent tumour types associated with positive lymph node metastasis.



Fig. 14. Female dog, mastectomy specimen. Regional lymph node metastasis (arrow).

Statistical analysis

Tables 3 and 4 elucidate the differences observed between benign and malignant tumours, with regard to the evaluated variables. Proliferation mitotic and Ki-67 labelling indices are presented in Table 4. Ki-67 evaluation was possible in 137 cases (Fig 15) (19 cases showed no immunostaining and were excluded of the analysis). Based on the median values of these indices, a significant difference was observed between benign and malignant tumours.

Table 3. Association between clinical parameters and histological diagnosis observed in the present series

Clinical parameter	Benign n (%)	Malignant n (%)
Age (n=146)	II (70)	II (70)
≤9 years	26 (35.1%)	48 (64.9%)
>9 years	16 (22.2%)	56 (77.8%)
>) years	P=0.1	30 (11.070)
Breed (n=145)		
Mixed breed	12 (25%)	36 (75%)
Poodle	7 (28%)	18 (72%)
Cocker spaniel	5 (29.4%)	12 (70.6%)
Others	17 (30.9%)	38 (69.1%)
	P=0.92	
Ovariohysterectomy (n=98)		
No	15 (21.4%)	55 (78.6%)
Yes, prior to tumour development	2 (15.4%)	11 (84.6%)
Yes, performed with mastectomy	2 (13.3%)	13 (86.7%)
	P=0.78	
Contraception (n=98)		
No	13 (18.1%)	59 (81.9%)
Yes	0 (0%)	8 (100%)
	P=0.34	
Tumour size (n=149)		
<3 cm	42 (52.5%)	38 (47.5%)
3-5 cm	4 (9.8%)	37 (90.2%)
>5 cm	0 (0%)	28 (100%)
	<i>P</i> <0.0001	
Tumour location (n=89)		
Cranial glands (thoracic)	1 (12.5%)	7 (87.5%)
Medial gland (cranial abdominal)	2 (14.3%)	12 (85.7%)
Caudal glands (caudal abdominal and inguinal)	11 (22%)	39 (78%)
Multiple	1 (5.9%)	16 (94.1%)
	P=0.47	
Skin ulceration (n=154)		
No	46 (34.6%)	87 (65.4%)
Yes	0 (0%)	22 (100%)
	P<0.0001	

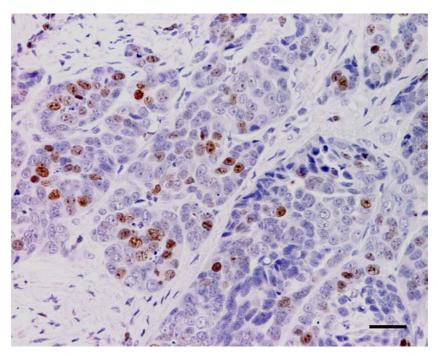


Fig. 15. Mammary gland; dog. Solid carcinoma showing nuclear Ki-67 positive immunostaining. Bar= $30\mu m$.

Malignant tumours were found to be associated with increased tumour size (P<0.0001), presence of skin ulceration (P<0.0001) and necrosis (P<0.0001), infiltrative tumour growth (P<0.0001) and a marked inflammatory cell response (P=0.006). Stromal/vascular invasion and lymph node metastases were restricted to malignant lesions. As for proliferation, these tumours also showed high mitotic (P<0.0001) and Ki-67 (P<0.0001) labelling indices.

Considering the malignant tumour group, several differences were observed across distinct histological types (Table 5). Simple carcinomas and carcinosarcomas showed a larger tumour size, whereas complex carcinomas were significantly smaller (P=0.044). In addition, the latter group was significantly associated with an expansive growth (P<0.0001), low differentiation grade (P<0.0001), reduced stromal (P<0.0001) and vascular invasion (P<0.0001). Simple carcinomas and carcinosarcomas were significantly associated with the presence of node metastases (P<0.0001). As for proliferation, complex carcinomas presented the lowest labelling indices, whereas the other histological types were significantly associated with increased proliferative mitotic and Ki-67 indices.

Table 4. Associations between histopathological parameters and histological diagnosis observed in the present series.

Histopathological parameter	Benign n (%)	Malignant n (%)	
Necrosis (n=156)	(, , ,	(, , ,	
Absent	34 (87.2%)	5 (12.8%)	
Present	12 (10.3%)	105 (89.7%)	
	<i>P</i> <0.0001		
Mode of growth (n=156)			
Expansive	46 (59.7%)	31 (40.3%)	
Infiltrative	0 (0%)	79 (100%)	
	P<0.0001	,	
Inflammatory cellular infiltrates (n=156)			
Slight/Absent	26 (44.8%)	32 (55.2%)	
Moderate	15 (25.9%)	43 (74.1%)	
Abundant	5 (12.5%)	35 (87.5%)	
	P=0.002	, ,	
Stromal Invasion (n=156)			
Absent	46 (63%)	27 (37%)	
Present	0 (0%)	83 (100%)	
	P<0.0001	, , ,	
Lymphovascular Invasion (n=156)			
Absent	46 (50.5%)	45 (49.5%)	
Present	0 (0%)	65 (100%)	
	P<0.0001	, , ,	
Lymph node metastasis (n=68)			
Absent	8 (22.2%)	28 (77.8%)	
Present	0 (0%)	32 (100%)	
	P=0.006	, ,	
Mitotic Index			
Median (Minimum-Maximum)	0.2 (0-1.47)	0.7 (0-2.99)	
,	P<0.0001	, ,	
Ki-67 Index			
Median (Minimum-Maximum)	11.86 (6.81-26.93)	23.5 (5.39-56.36)	
,	P<0.0001	, ,	

Kaplan-Meier univariate analysis

Follow-up data concerning OS was available in 69 bitches with malignant tumours. During the follow-up period, according to the referring surgeons, 37 animals died or were euthanized due to metastatic disease and/or local recurrence (25 with distant metastases, 7 with local recurrence and 5 with both recurrence and distant metastases).

Table 5. Significant differences found between distinct malignant groups in the present series.

Clinicopathological parameter	Simple carcinoma	Complex carcinoma	Carcinosarcoma
Tumour size (n=100)			
<3 cm	13 (35.1%)	22 (59.5%)	2 (5.4%)
3-5 cm	18 (51.4%)	13 (37.1%)	4 (11.4%)
>5 cm	15 (53.6%) P=0.044	7 (25%)	6 (21.4%)
Mode of growth (n=110)			
Expansive	4 (12.9%)	27 (87.1%)	0 (0%)
Infiltrative	43 (56.6%)	18 (23.7%)	15 (19.7%)
	P < 0.0001	,	,
Histological grade (n=107)			
Grade I	1 (5.6%)	16 (88.9%)	1 (5.6%)
Grade II	9 (26.5%)	23 (67.6%)	2 (5.9%)
Grade III	37 (27.3%)	6 (10.9%)	12 (21.8%)
	P<0.0001	. ()	(,
Stromal Invasion (n=110)			
Absent	4 (14.8%)	23 (85.2%)	0 (0%)
Present	43 (53.8%) P<0.0001	22 (27.5%)	15 (18.8%)
Lymphovascular Invasion (n=110)			
Absent	6 (13.3%)	35 (77.8%)	4 (8.9%)
Present	41 (66.1%) P<0.0001	10 (16.1%)	11 (17.7%)
Lymph node metastasis (n=57)			
Absent	7 (26.9%)	18 (69.2%)	1 (3.8%)
Present	25 (80.6%)	1 (3.2%)	5 (16.1%)
	P<0.0001	` ,	, ,
Mitotic Index			
Median (Min-Max)	0.96 (0.1-2.99) <i>P</i> <0.0001	0.46 (0.0-1.6)	0.8 (0.1-1.90)
Ki-67 Index			
Median (Min-Max)	27.5 (12.10- <i>P</i> <0.0001	17.84 (5.39-	27.2 (10.2-40.9)

Distant metastases were predominantly found in the lung (n=28) and liver (n=8) (Fig. 16 and 17). At 12 months after mastectomy, 33 (47.83%) animals have died whereas 36 of the 69 (52.17%) dogs enrolled in the follow-up study were alive; the median overall survival was 15 months. Tables 6 and 7 present those factors significantly associated with OS and DFS, respectively.

Statistically significant associations were achieved between OS and size (P=0.0042), skin ulceration (P=0.0322), tumour growth (P<0.0001), stromal and lymphovascular invasion (P<0.0001 and P<0.0001, respectively), lymph node status (P<0.0001),

mitosis (P=0.0143) and Ki-67 (P=0.0001). Histological type and grade were also significantly related to OS. Dogs with simple or carcinosarcoma histotype showed poorer OS times (P<0.0001) than did the ones affected by complex carcinomas. Survival was significantly worse in grade III cases (P<0.0001), compared to grade I/II.

Table 6. Factors significantly associated to overall survival in malignant tumours.

		Overall survival		
Variable		Mean survival (months)	Average 1-year survival rate (n[%])	Р
Tumour size				
<3 cm	27	48.63	19 (70.37)	0.0042
3-5 cm	17	19.59	9 (52.94)	
>5 cm	19	10.21	5 (26.32)	
Skin ulceration				
No	55	37.41	31 (56.36)	0.032
Yes	13	10.62	5 (38.46)	
Histological type				
Simple carcinoma	24	8.21	6 (25)	< 0.0001
Complex carcinoma	36	49.68	28 (77.78)	
Carcinosarcoma	9	7.33	2 (22.22)	
Mode of growth				
Expansive	24	58.14	22 (91.67)	< 0.0001
Infiltrative	45	9.08	14 (31.11)	
Histological grade				
Grade I/II	36	47.43	28 (77.78)	< 0.0001
Grade III	33	9.38	8 (24.24)	
Stromal Invasion				
Absent	20	57.88	19 (95)	< 0.0001
Present	49	13.91	17 (34.69)	
Lymphovascular Invasion				
Absent	32	56.09	29 (90.63)	< 0.000
Present	37	6.57	7 (18.92)	
Lymph node metastasis				
Absent	15	39.74	13 (86.67)	< 0.0001
Present	15	6.53	3 (20)	
Mitotic index				
< 0.7	35	39.24	25 (71.43)	0.0143
≥0.7	29	17.86	9 (31.03)	
Ki-67 index				
<23.5	35	47.62	26 (74.29)	0.0001
≥23.5	29	13.35	8 (27.59)	

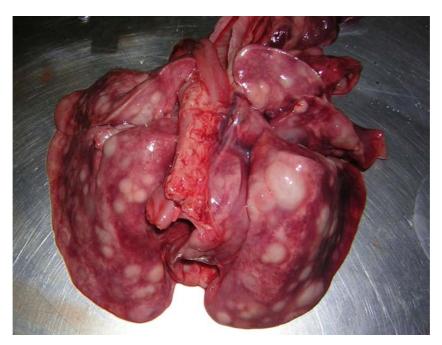


Fig. 16. Female dog. Lung showing multiple metastases of a mammary carcinosarcoma.

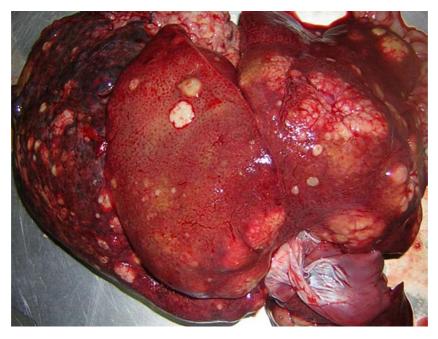


Fig. 17. Female dog. Liver showing multiple metastases of a mammary carcinoma.

Data for DFS was available in 68 cases. One case was excluded from this study because the bitch developed oncologic disease within 10 to 30 days after surgery. At 12 months after mastectomy, 33.8% (23/68) of dogs were free of oncological disease; the median disease-free survival was 9 months. Statistically significant differences were achieved between DFS and tumour size (P=0.04), histological type (P=0.0037), tumour growth (P<0.0001), grade (P=0.002), stromal and lymphovascular invasion (P=0.0018 and P<0.0001, respectively), lymph node invasion on clinical presentation (P=0.0017), mitosis (P=0.0134) and Ki-67 labelling index (P=0.0005).

Table 7. Factors significantly associated to disease-free survival in malignant tumours.

		Disease-free survival		
Variable	n	Mean survival (months)	Average 1-year survival rate (n[%])	P
Tumour size				
<3 cm	27	14.56	13 (48.15)	0.04
3-5 cm	17	17.82	8 (47.06)	
>5 cm	18	6.22	2 (11.11)	
Histological type				
Simple carcinoma	23	7.22	6 (26.09)	0.0037
Complex carcinoma	36	17.88	17 (47.22)	
Carcinosarcoma	9	5.78	0 (0)	
Mode of growth				
Expansive	24	22.33	15 (62.5)	< 0.0001
Infiltrative	44	6.86	8 (18.18)	
Histological grade				
Grade I/II	36	17.31	16 (44.44)	0.002
Grade III	32	7.97	7 (21.88)	
Stromal Invasion				
Absent	20	18.46	12 (60)	0.0018
Present	48	10.05	11 (22.92)	
Lymphovascular Invasion				
Absent	32	22.05	19 (59.38)	< 0.0001
Present	36	4.97	4 (11.11)	
Lymph node metastasis				
Absent	15	17.93	9 (60)	0.0017
Present	14	4.93	2 (14.29)	
Mitotic index				
< 0.7	35	17.29	15 (42.86)	0.0134
≥0.7	28	8.56	7 (25)	
Ki-67 index				
<23.5	35	18.35	17 (48.57)	0.0005
≥23.5	28	7.17	5 (17.86)	

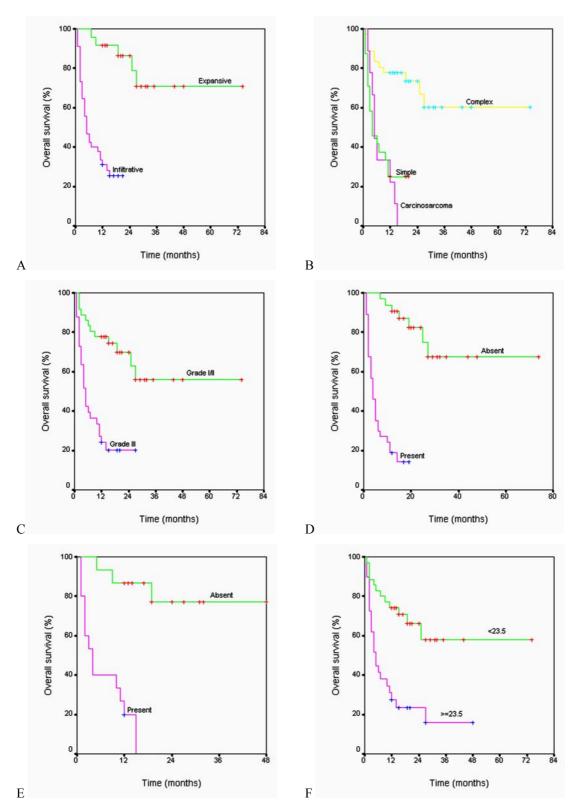


Fig. 18. Overall Kaplan-Meier survival curves for dogs with malignant mammary tumours. A - Tumour growth (expansive vs. infiltrative), P<0.0001; B - Histological type (complex vs. simple vs. carcinosarcoma), P<0.0001; C- Histological grade (Grade I/II vs. Grade III), P<0.0001; D-Lymphovascular invasion (Absent vs. present), P<0.0001; E- Lymph node metastases (Absent vs. present), P<0.0001; F- Ki-67 labelling index (high vs. low index), P=0.0001.

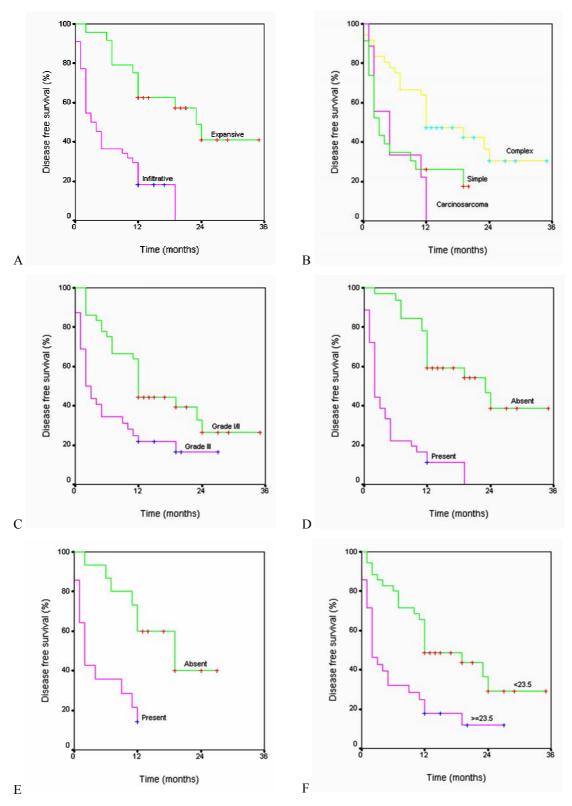


Fig. 19. Disease-free Kaplan-Meier survival curves for dogs with malignant mammary tumours. A - Tumour growth (expansive vs. infiltrative), P<0.0001; B - Histological type (complex vs. simple vs. carcinosarcoma), P=0.0037; C- Histological grade (Grade I/II vs. Grade III), P=0.002; D-Lymphovascular invasion (Absent vs. present), P<0.0001; E- Lymph node metastases (Absent vs. present), P=0.0017; F- Ki-67 labelling index (high vs. low index), P=0.0005.

Multivariate analysis, proportional hazard model

Multivariate analysis including factors significantly associated with survival by the log-rank test was performed using forward Cox regression method. Multivariate analysis disclosed lymph node status as significantly associated with overall and disease-free survival (Table 8). The inclusion of this variable reduced the number of available observations considerably because of missing data, and therefore models without this variable were also estimated, confirming lymphovascular invasion as an independent prognostic factor for both overall (HR: 19.33; 95% CI: 5.65-66.15; P<0.0001) and disease-free survival (HR: 6.75; 95% CI: 3.19-14.25; P<0.0001).

Table 8. Significant prognostic factors in multivariate analysis for dogs with malignant tumours.

Dependent variable	Independent variables	Hazard ratio (HR)	95% CI	P
Overall survival	Lymph node metastases			
	Absent	Referent		
	Present	10.924	2.38-50.12	0.002
Disease-free survival	Lymph node metastases			
	Absent	Referent		
	Present	4.494	1.50-13.46	0.007

Discussion

In this study we have characterized a series of benign and malignant tumours at the clinical and pathological level and we performed an evaluation of proliferation labelling indices. These variables were compared across benign and malignant tumour types and a survival analysis was conducted in order to improve understanding of prognostic factors in canine mammary tumours. Determining the prognosis of a canine patient with a malignant mammary tumour is very important for the clinician, but it is often difficult because the biologic behaviour of these tumours varies widely. The major challenge is to find those prognostic variables that allow the prediction of disease behaviour in the individual case (Sarli *et al.*, 2002).

A diagnosis of malignancy was significantly associated with clinical and pathological aggressive features. In our series, increased tumour size and ulceration were significantly associated with tumour malignancy, with all tumours larger than 5cm and presenting skin ulceration classified as malignant. Univariate analysis of survival

showed that these clinical variables were of prognostic value on OS, confirming previous studies (Bostock, 1975; Misdorp and Hart, 1976; Shofer *et al.*, 1989; Yamagami *et al.*, 1996b; Matos, 2007). Tumour size was also significantly associated with DFS, which was also described by others (Peña *et al.*, 1998; Martin de las Mulas *et al.*, 2005). In our study, ulceration was not found of prognostic value on disease-free survival, which is contrast with previous studies (Hellmén *et al.*, 1993; Peña *et al.*, 1998; Queiroga and Lopes, 2002).

We have found no differences between benign and malignant tumours with regard to animal age at the time of surgical procedure. Also, no statistical significant association was observed with clinical outcome. The available literature shows similar mean age values to our study (around 9-10 years), but opposing results exists concerning its prognostic information; although several investigations described animal age as a prognostic factor, with old animals associated with more aggressive tumours (Schneider *et al.*, 1969; Hellmén *et al.*, 1993; Peña *et al.*, 1998; Nieto *et al.*, 2000), others have not confirmed these findings (Philibert *et al.*, 2003; Martin de las Mulas *et al.*, 2005; Matos, 2007). These contradictory results might be related with differences in statistical approaches (some studies have considered age as a continuous and others as a categorical variable) or with animal age variations between studies.

No significant associations were found between other clinical variables and tumour diagnosis or outcome. However, an association was recently described between small animal breeds and a lower rate of tumour malignancy (Itoh *et al.*, 2005), when comparing small vs. other breeds. Although ovariohysterectomy has been decribed as having a protective effect on the development of mammary tumours and some studies have reported its association with increased survival times (Sorenmo *et al.*, 2000; Chang *et al.*, 2005), the prognostic value of this procedure is still under debate. Similarly to the present findings, other researchers found no association between ovariohysterectomy before or at the time of mastectomy and patient prognosis (Morris *et al.*, 1998; Philibert *et al.*, 2003). In addition, no association between the administration of contraceptives and malignancy or outcome was observed, as previously reported by other studies (Hellmén *et al.*, 1993; Peña *et al.*, 1998; Nieto *et al.*, 2000; Martin de las Mulas *et al.*, 2005).

It is obvious from the available literature the controversy around histological classification of canine mammary tumours. Several classification systems have been

proposed in the last decades (Fowler et al., 1974; Bostock, 1975; Moulton, 1990) and the published studies parallel these distinct criteria. Diverse histological categorizations have been used, rendering a comparison between studies difficult. With the recent proposed WHO classification, a prognostic element has been introduced, separating complex and simple carcinomas, the latter characterized by higher malignancy (Misdorp et al., 1999). Considering this classification system and given that we had a small number of samples in some histological types, we have grouped carcinomas into these two main groups of carcinomas and we have considered a third group, composed only by those tumours fulfilling malignant criteria in both epithelial and mesenchymal tumour components (carcinosarcoma group). When we considered these 3 major groups of malignant tumours, we found that complex carcinomas were associated with less aggressive pathological features such as small size, expansive tumour growth, low to moderate histological grade, lack of stromal and vascular invasion and reduced number of lymph node metastases. Accordingly, this tumour group was significantly associated with the lowest proliferation indices. These characteristics were reflected on survival analysis, which revealed better overall and disease-free survival times for this type of neoplasms. Despite some differences in tumour categorization, our results are in conformity with other studies, who also found complex carcinomas as the ones showing a more favourable clinical behaviour (Misdorp and Hart, 1976; Yamagami et al., 1996b; Matos, 2007). However, recent studies did not found a statistical significant association between histological type and disease-free survival (Martin de las Mulas et al., 2005), probably because of differences in tumour categories.

Complex lesions of the present series were characterized by a myoepithelial component, usually readily identified on HE stained sections, admixed with the proliferation of luminal epithelial cells. The participation of myoepithelial cells has been associated with the better prognosis assigned to complex carcinomas (Yamagami *et al.*, 1996b), giving that these cells have been described as having a tumour/invasive suppressor function in several human studies (Sternlicht *et al.*, 1997; Barsky, 2003; Jones *et al.*, 2003; Adriance *et al.*, 2005). However, although these biphasic tumours (myoepithelial and epithelial differentiation) are associated with low malignancy in canine (Yamagami *et al.*, 1996b), feline (Seixas *et al.*, 2008) and human (Foschini and Eusebi, 1998) species, pure malignant myoepithelioma show a distinct clinical behaviour, related with poorer outcome (Foschini and Eusebi, 1998). Accordingly, the spindle cell carcinoma of

our series submitted to follow up (n=3) showed an aggressive behaviour, with overall survival times ranging from 4 to 10 months. Although several recent studies have drawn attention to mammary myoepithelial cells, this cell type has been largely neglected and our understanding of the functions of this second major mammary cell population in mammary gland tumorigenesis remains very limited (Faraldo *et al.*, 2005).

In contrast to complex carcinomas, simple carcinoma and carcinosarcoma types showed very aggressive features, as described by previous studies (Misdorp and Hart, 1976; Misdorp et al., 1999; Sorenmo, 2003; Matos, 2007). Simple carcinoma group comprised a variety of histological types, but a small number of cases per type had available follow-up in order to perform a consistent statistical analysis. Future studies are needed in order to compare their biological behaviour. One histological type that deserves further attention is the micropapillary invasive carcinoma, which have been associated with a very aggressive behaviour in canine (Cassali et al., 2002b; Gama et al., 2008) and feline (Seixas et al., 2007) species, as well as in humans (Siriaunkgul and Tavassoli, 1993; Kuroda et al., 2004; Kim et al., 2005; Nassar, 2004; Putti et al., 2005). In the present series, 3 cases were described, all associated with rapid progression of the oncologic disease. Only one case was included in the follow-up study, since the other 2 dogs died immediately after surgery. Another rare histological type found in our tumour series (n=2) was the pleomorphic lobular carcinoma, a recognized subtype of invasive lobular carcinoma described in the human species (Eusebi et al., 1992; Radhi, 2000). One case of canine pleomorphic lobular carcinoma has been reported previously by Cassali et al. (2002), which described similar cytomorphologic features to the ones found in our cases (Cassali et al., 2002a).

With regard to histological grade, grade III tumours were found significantly associated with simple carcinoma and carcinosarcoma types, shorter overall and disease-free survival rates, when compared to grade I and II tumours. Our results are in accordance to previous reports on canine (Karayannopoulou *et al.*, 2005; Martin de las Mulas *et al.*, 2005; Matos, 2007) and human (Elston and Ellis, 1991) tissues.

In our study, the presence of tumour necrosis was associated with a malignant phenotype but it was not associated with decreased survival intervals. This result is similar to a recent study (Matos, 2007) but contradicts another, in which necrosis was associated with a shorter disease-free survival (Martin de las Mulas *et al.*, 2005).

An infiltrative growth pattern and stromal invasion were restricted to the malignant tumour group, being significantly associated with simple and carcinosarcoma histological types, as well as with poorer survival times. This result is in accordance to some studies (Misdorp and Hart, 1976; Sarli *et al.*, 2002), but contradicts others who failed to find differences between tumour growth pattern or stromal invasion and prognosis (Itoh *et al.*, 2005; Martin de las Mulas *et al.*, 2005).

Lymphoid cellular infiltration was associated with malignancy, but no other associations were found. It was suggested that lymphoid cellular reactivity could indicate an anti-tumour immune response and it was associated with a better prognosis (Gilbertson *et al.*, 1983), but several studies have also failed in finding such an association (Martin de las Mulas *et al.*, 2005; Matos, 2007).

In univariate analysis, lymphovascular invasion was significantly associated with simple and carcinosarcoma tumour types, and strongly associated with local recurrence/distant metastases and decreased overall survival rates. Dogs affected by tumours showing no lymphovascular invasion had a significant survival advantage and a reduced risk of relapse. Our findings confirm previous reports both on canine (Gilbertson *et al.*, 1983; Yamagami *et al.*, 1996b; Martin de las Mulas *et al.*, 2005) and human breast cancer (Pinder *et al.*, 1994; Elston *et al.*, 1998). However, other authors failed to find such an association (Misdorp and Hart, 1976).

Our results also showed that tumour growth fraction, as assessed by mitotic and Ki-67 labelling indices, is an important predictor of survival. Ki-67 labelling index values of more than 23.5% were significantly associated with shorter overall and disease-free survival times. This comes in accordance with reported findings in canine (Peña *et al.*, 1998; Nieto *et al.*, 2000; Sarli *et al.*, 2002; de Matos *et al.*, 2006) and human (Bouzubar *et al.*, 1989; Brown and Gatter, 1990; Veronese and Gambacorta, 1992; Pinder *et al.*, 1995) tissues, which described Ki-67 as a valuable prognostic factor.

Lymph node status was of statistically significant prognostic value, confirming several previous canine (Gilbertson *et al.*, 1983; Nieto *et al.*, 2000; Itoh *et al.*, 2005; Martin de las Mulas *et al.*, 2005) and human breast cancer studies (Elston *et al.*, 1998; Ellis *et al.*, 2003). Despite the identification of several parameters with prognostic value on univariate analysis, their prognostic power was not retained in the Cox regression multivariate analysis, which considered lymph node status as the only independent variable. As already confirmed by previous studies, this parameter is obviously of great

importance in predicting clinical outcome of bitches with malignant mammary tumours, but several studies failed on proving its prognostic value in multivariate analysis (Misdorp and Hart, 1976; Hellmén *et al.*, 1993; Martin de las Mulas *et al.*, 2005). A number of reasons have been listed to explain these results: the small number of lymph nodes evaluated (Martin de las Mulas *et al.*, 2005), the inclusion of sarcomas (which probably attenuate the effect of node status, giving that sarcomas are usually associated with haematogeneous spreading, rather than lymphatic) (Hellmén *et al.*, 1993) or even the particular characteristics of the lymph node network associated with the canine mammary gland (Misdorp and Hart, 1976). Additional studies are warranted evaluating a larger series in order to confirm our present results.

If lymph node status was not included in the multivariate analysis, lymphovascular invasion replaced it as the most important predictive factor for survival in this group of dogs. Sarli *et al.* (2002) also found lymphovascular invasion as an independent prognosticator, but the study performed by Yamagami and co-workers (1996b) did not revealed this significance, despite describing this variable as a prognostic factor in univariate analysis (Yamagami *et al.*, 1996b). The most likely explanation for such discrepancies is related to problems in the distinction of true vessels, especially lymphatics, from artefactual soft tissue spaces due to fixation shrinkage artefact (Elston *et al.*, 1998; Martin de las Mulas *et al.*, 2005).

Survival time is considered a useful criterion for evaluating prognosis in both man and animals (Misdorp, 1987). Although, in our study, several parameters were considered as useful prognostic factors, the present results should be interpreted with caution given the relatively small number of cases with follow-up. All cases were treated by surgery alone and followed for a minimum period of 12 months. During this time, all occurring deaths (n=33) were tumour-related and no dog was lost to follow-up. Since a number of cases are still being submitted to follow-up (12 dogs with a follow-up period inferior to 24 months), we have presented the survival rates observed one year after surgical treatment. However, it is our objective to extend the post-surgical evaluation in order to confirm the validity of our first results.

Canine mammary tumours with apparent signs of malignancy do not present any diagnostic problems for experienced clinicians. However, because many canine tumours are not at an advanced stage of development when first detected by owners or clinicians, the potential biologic behaviour of these tumours is rather difficult to predict (Hellmén

et al., 1993). Pathologists certainly represent a signicant role in the management of dogs with mammary cancer, by providing reliable prognostic information to clinicians. From our preliminary study, lymph node status was the only independent prognostic factor in canine mammary malignant tumours; unfortunately, a considerable number of mastectomy and lumpectomy specimens received in the histopathology laboratories are not accompanied by regional lymph nodes, fact that was reflected on our and other studies. We believe that it is of major importance to encourage veterinary surgeons to routinely remove lymph nodes whenever they perform a mastectomy, even if no clinical signs of metastases are observed.

Acknowledgements

The authors thank Prof. Fátima Gärtner (Institute of Biomedical Science at the University of Porto, Portugal) for the contribution of some cases included in this study. We also thank Mrs Lígia Bento for expert technical assistance. This work was supported by the Centro de Ciência Animal e Veterinária (CECAV), University of Trás os Montes e Alto Douro (UTAD), Vila Real, Portugal, and by Portuguese Science and Technology Foundation, project POCTI/CVT/57795/2004.

References

- Adriance MC, Inman JL, Petersen OW, Bissell MJ. (2005). Myoepithelial cells: good fences make good neighbors. Breast Cancer Res 7:190-197.
- Barsky SH. (2003). Myoepithelial mRNA expression profiling reveals a common tumor-suppressor phenotype. Exp Mol Pathol 74:113-122.
- Bostock DE. (1975). The prognosis following the surgical excision of canine mammary neoplasms. Eur J Cancer 11:389-396.
- Bouzubar N, Walker KJ, Griffiths K, Ellis IO, Elston CW, Robertson JF, Blamey RW, Nicholson RI. (1989). Ki67 immunostaining in primary breast cancer: pathological and clinical associations. Br J Cancer 59:943-947.
- Brown DC, Gatter KC. (1990). Monoclonal antibody Ki-67: its use in histopathology. Histopathology 17:489-503.
- Cassali GD, Gärtner F, Schmitt FC. (2002a). Pleomorphic lobular carcinoma of the canine mammary gland: histopathologic and immunohistochemical features. Arq Bras Med Vet Zootec 54:592-594.
- Cassali GD, Serakides R, Gärtner F, Schmitt FC. (2002b). Invasive micropapillary carcinoma of the dog mammary gland. A case report. Arq Bras Med Vet Zootec 24:366-369.
- Chang SC, Chang CC, Chang TJ, Wong ML. (2005). Prognostic factors associated with survival two years after surgery in dogs with malignant mammary tumours: 79 cases (1998-2002). J Am Vet Med Ass 227:1625-1629.
- de Matos AJ, Lopes CC, Faustino AM, Carvalheira JG, Dos Santos MS, Rutteman GR, Gärtner MF. (2006). MIB-1 labelling indices according to clinico-pathological variables in canine mammary tumours: a multivariate study. Anticancer Res 26:1821–1826.
- Ellis IO, Schnitt SJ, Sastre-Garau X, Bussolati G, Tavassoli FA, Eusebi V, Peterse JL, Mukai K, Tabár L, Jacquemier J. (2003). Invasive breast carcinoma. In: Pathology and Genetics of tumours of the breast and female genital organs. FA Tavassoli e P Deville (Eds). World Health Organization Classification of Tumours. IARC Press, Lyon, France, pp. 13-59.
- Elston CW, Ellis IO. (1991). Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term follow-up. Histopathology 19:403–410.
- Elston CW, Ellis IO. (1998). Assesment of histological grade. In: Systemic Pathology The Breast. CW Elston and IO Ellis (Eds.), 3rd Ed., vol. 13. Churchill and Livingstone, London, pp. 365–384.

- Elston CW, Ellis IO, Goulding H, Pinder SE. (1998). Role of pathology in the prognosis and management of breast cancer. In: Systemic Pathology The Breast. CW Elston and IO Ellis (Eds.), 3rd Ed., vol. 13., Churchill and Livingstone, London, pp. 385-400.
- Eusebi V, Magalhães F, Azzopardi JG. (1992). Pleomorphic lobular carcinoma of the breast: an aggressive tumor showing apocrine differentiation. Hum Pathol 23:655-662.
- Faraldo MM, Teulière J, Deugnier M-A, Hosseraye IT, Thiery JP, Glukhova MA. (2005). Myoepithelial cells in the control of mammary development and tumorigenesis: data from genetically modified mice. J Mammary Gland Biol Neoplasia 10:211-219.
- Foschini MP, Eusebi V. (1998). Carcinomas of the breast showing myoepithelial cell differentiation. A review of the literature. Virchows Arch 432:303-310.
- Fowler EH, Wilson GP, Koester A. (1974). Biologic behaviour of canine mammary neoplasms based on a histogenetic classification. Vet Pathol 11:212-229.
- Gama A, Alves A, Schmitt FC. (2008). Letter to the Editor: Invasive micropapillary carcinoma of canine mammary gland: an under recognized entity? Vet Pathol 45:600-601.
- Gilbertson SR, Kurzman ID, Zachrau RE, Hurvitz AI, Black MM. (1983). Canine mammary epithelial neoplasms: biologic implications of morphologic characteristics assessed in 232 dogs. Vet Pathol 20:127-142.
- Hellmén E, Bergstrom R, Holmberg L, Spangberg I-B, Hansson K, Lindgren A. (1993). Prognostic factors in canine mammary tumors: a multivariate study of 202 consecutive cases. Vet Pathol 30:20-27.
- Itoh T, Uchida K, Ishikawa K, Kushima K, Kushima E, Tama H, Moritake T, Nakao H, Shii H. (2005). Clinicopathological survey of 101 canine mammary gland tumors: differences between small-breed dogs and others. J Vet Med Sci 67:345-347.
- Jones JL, Shaw JA, Pringle JH, Walker RA. (2003). Primary breast myoepithelial cells exert an invasion-suppressor effect on breast cancer cells via paracrine down-regulation of MMP expression in fibroblasts and tumour cells. J Pathol 201:562-572.
- Karayannopoulou M, Kaldrymidou E, Constantinidis TC, Dessiris A. (2005). Histological grading and prognosis in dogs with mammary carcinomas: application of a human grading method. J Comp Pathol 133:246–252
- Kim MJ, Gong G, Joo HJ, Ahn SH, Ro JY. (2005). Immunohistochemical and clinicopathologic characteristics of invasive ductal carcinoma of breast with micropapillary carcinoma component. Arch Pathol Lab Med 129:1277-1282.
- Kuroda H, Sakamoto G, Ohnisi K, Itoyama S. (2004). Clinical and pathologic features of invasive micropapillary carcinoma. Breast Cancer 11:169-174.
- Martin de las Mulas J, Millán Y, Dios R. (2005). A prospective analysis of immunohistochemically determined Estrogen Receptor α and Progesterone Receptor

- Expression and host and tumor factors as predictors of disease-free period in mammary tumors of the dog. Vet Pathol 42:200–212.
- Matos AJF. (2007). Prognostic factors in canine mammary tumours. PhD Thesis, University of Oporto, Portugal.
- Misdorp W. (1987). The impact of pathology on the study and treatment of cancer. In: Veterinary Cancer Medicine, GH Theilen and BR Madewell (Eds), Lea and Febiger, Philadelphia, pp. 53–70.
- Misdorp W. (2002). Tumours of the mammary gland. In: Tumors in Domestic Animals, DJ Meuten (Ed), 4th Edn. Iowa State Press, Blackwell Publishing Company, pp. 575-606.
- Misdorp W, Else RW, Hellmén E, Lipscomb TP. (1999). Histological classification of mammary tumors of the dog and the cat. 2nd series. Vol VII. Washington DC, Armed Forces Institute of Pathology, American Registry of Pathology and the World Health Organization Collaborating Center for Worldwide reference on Comparative Oncology, pp. 1-59.
- Misdorp W, Hart AA. (1976). Prognostic factors in canine mammary cancer. J Natl Cancer Inst 56:779-786.
- Morris JS, Dobson JM, Bostock DE, O'Farrell E. (1998). Effect of ovariohysterectomy in bitches with mammary neoplasms. Vet Rec 142:656-658.
- Moulton JE. (1990). Tumors of the mammary gland. In: Tumors in domestic animals. JE Moulton Ed., 3rd Ed., California Press, Berkeley, California, pp. 518-553.
- Nassar H. (2004). Carcinomas with micropapillary morphology: clinical significance and current concepts. Adv Anat Pathol 11:297-303.
- Nieto A, Peña L, Perez-Alenza MD, Sanchez MA, Flores JM, Castaño M. (2000). Immunohistologic detection of estrogen receptor alpha in canine mammary tumors: clinical and pathologic associations and prognostic significance. Vet Pathol 37:239–247.
- Peña L, Nieto A, Pérez-Alenza D, Cuesta P, Castano M. (1998). Immunohistochemical detection of Ki-67 and PCNA in canine mammary tumors: relationship to clinical and pathologic variables. J Vet Diagn Invest 10:237-246.
- Philibert JC, Snyder PW, Glickman N, Glickman LT, Knapp DW, Waters DJ. (2003). Influence of host factors on survival in dogs with malignant mammary gland tumours. J Vet Intern Med 17:102-106.
- Pinder SE, Ellis IO, Galea M, O'Rouke S, Blamey RW, Elston CW. (1994). Pathological prognostic factors in breast cancer. III. Vascular invasion: relationship with recurrence and survival in a large study with long-term follow-up. Histopathology 24:41-47.
- Pinder SE, Wencyk P, Sibbering DM, Bell JA, Elston CW, Nicholson R, Roertson JF, Blamey RW, Ellis IO. (1995). Assessment of the new proliferation marker MIB1 in breast

- carcinoma using image analysis: associations with other prognostic factors and survival. Br J Cancer 71:146-149.
- Putti TC, Pinder SE, Elston CW, Lee AHS, Ellis IO. (2005). Breast pathology practice: most common problems in a consultation service. Histopathology 47:445-457.
- Queiroga F, Lopes C. (2002). Canine mammary tumours, research on new prognostic factors. Rev Port Ciências Veterinárias 97:119-127.
- Radhi JM. (2000). Immunohistochemical analysis of pleomorphic lobular carcinoma: higher expression of p53 and chromogranin and lower expression of ER and PgR. Histopathology 36:156-160.
- Rutteman GR, Withrow SJ, MacEwen EG. (2001). Tumors of the mammary gland. In: Small Animal Clinical Oncology, SJ Withrow and BR MacEwen (Ed.), WB Saunders Company, Philadelphia, pp. 455–477.
- Sarli G, Preziosi R, Benazzi C, Castellani G, Marcato PS. (2002). Prognostic value of histologic stage and proliferative activity in canine malignant mammary tumors. J Vet Diagn Invest 14:25–34.
- Schneider R, Dorn CR, Taylor DON. (1969). Factors influencing canine mammary cancer development and postsurgical survival. J Natl Cancer Inst 43:1249-1261.
- Seixas F, Palmeira C, Pires MA, Lopes C. (2007). Mammary invasive micropapillary carcinoma in cats: clinicopathologic features and nuclear DNA content. Vet Pathol 44:842-848.
- Seixas F, Pires MA, Lopes C. (2008). Complex carcinomas of the mammary gland in cats: pathological and immunohistochemical features. Vet J 176:210-215.
- Shofer FS, Sonnenschein EG, Goldschmidt MH, Laster LL, Glickman LT. (1989). Histopathologic and dietary prognostic factors for canine mammary carcinoma. Breast Cancer Res Treat 13:49-60.
- Siriaunkgul S, Tavassoli FA. (1993). Invasive micropapillary carcinoma of the breast. Modern Pathol 6:660-662.
- Sorenmo K. (2003). Canine mammary gland tumors. Vet Clin North Am 33:573-596.
- Sorenmo K, Shofer FS, Goldschmidt MH. (2000). Effect of spaying and timing of spaying on survival of dogs with mammary carcinoma. J Vet Intern Med 14:266-270.
- Sternlicht MD, Kedeshian P, Shao ZM, Safarians S, Barsky SH. (1997). The human myoepithelial cell is a natural tumor suppressor. Clin Cancer Res 3:1949-1958.
- Veronese SM, Gambacorta M. (1992). Detection of Ki-67 proliferation rate in breast cancer-correlation with clinical and pathologic features. Am J Clin Pathol 95:30-34.
- Yamagami T, Kobayashi T, Takahashi K, Sugiyama M. (1996a). Influence of ovariectomy at the time of mastectomy on the prognosis for canine malignant mammary tumors. J Small Anim Pract 37:462-464.

- Yamagami T, Kobayashi T, Takahashi K, Sugiyama M. (1996b). Prognosis for canine malignant mammary tumors based on TNM and histologic classification. J Vet Med Sci 58:1079-1083.
- Zaidan Dagli ML. (2008). The search for suitable prognostic markers for canine mammary tumors: A promising outlook. Vet J 177:3-5.

Chapter III

 $\label{eq:catherin} \text{Expression of E-cadherin, P-cadherin and } \beta\text{-catenin in canine malignant mammary tumours in} \\ \text{relation to clinicopathological parameters, proliferation and survival.}$

Gama A, Paredes J, Gärtner F, Alves A, Schmitt F.

The Veterinary Journal 177: 49-53, 2008



Available online at www.sciencedirect.com



The Veterinary Journal 177 (2008) 45-53



Expression of E-cadherin, P-cadherin and β-catenin in canine malignant mammary tumours in relation to clinicopathological parameters, proliferation and survival

Adelina Gama ^a, Joana Paredes ^{b,c}, Fátima Gärtner ^{b,d}, Anabela Alves ^a, Fernando Schmitt ^{b,c,e,*}

Accepted 27 May 2007

Abstract

Cadherin–catenin complexes play a critical role in intercellular adhesion, and their altered expression has been implicated in tumour progression. In this study, the expression of E-cadherin, P-cadherin and β -catenin was analysed in 65 canine malignant mammary tumours and correlated with clinicopathological parameters, proliferation and survival. Reduction in E-cadherin expression was significantly associated with increased tumour size, high histological and invasion grades, lymph node metastasis and high mitotic index. Reduced β -catenin expression was associated with high histological and invasion grades. Anomalous expression of P-cadherin was only associated with invasion. In 39 cases for which follow-up data were available, reduced E-cadherin and β -catenin expression was significantly associated with shorter overall survival and disease free survival. Abnormal expression of adhesion molecules is a common phenomenon in canine mammary malignant tumours and may play a central role in tumour progression.

Keywords: Canine; Mammary tumours; E-cadherin; P-cadherin; β-Catenin

Introduction

Cadherins are calcium-dependent cell–cell adhesion molecules that play critical roles during embryogenesis and in the maintenance of normal adult tissue architecture (Takeichi, 1991; Gumbiner, 1996). Cadherins interact with several proteins termed catenins, including α -, β - and γ -catenin, which link cadherins to the actin cytoskeleton and mediate signal-transduction mechanisms that control

E-mail address: fernando.schmitt@ipatimup.pt (F. Schmitt).

cellular events, including cell polarity, differentiation, growth and migration (Knudsen et al., 1998).

The best characterised and most widely distributed members of the family are the classical cadherins, namely epithelial (E-) and placental (P-) cadherins (Nose and Takeichi, 1986). E-cadherin is found in almost all human epithelial tissues, whereas P-cadherin is restricted to the basal layers of stratified epithelium (Nose and Takeichi, 1986; Shimoyama et al., 1989). In normal human breast and canine mammary tissue, these molecules show a distinct pattern of expression; E-cadherin is expressed by luminal epithelial cells, whereas expression of P-cadherin is restricted to myoepithelial cells (Shimoyama et al., 1989; Palacios et al., 1995; Gama et al., 2002, 2004).

Department of Veterinary Sciences, University of Trás-os-Montes and Alto Douro (UTAD), 5001-911 Vila Real, Portugal
 Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP), 4200-465 Porto, Portugal
 Life and Health Science Research Institute (ICVS), Health Science School, University of Minho, Braga, Portugal
 Instituto de Ciências Biomédicas de Abel Salazar, University of Porto, 4099-003 Porto, Portugal
 Medical Faculty, University of Porto, Portugal

^{*} Corresponding author. Address: Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP), 4200-465 Porto, Portugal. Tel.: +351 22 5570700; fax: +351 22 5570799.

Loss or down-regulation of E-cadherin/β-catenin complexes is associated with oncogenic progression in human breast cancer (Gamallo et al., 1993; Yoshida et al., 2001). In addition, anomalous epithelial expression of P-cadherin in human breast cancer is associated with aggressive biological behaviour and poor outcome (Palacios et al., 1995; Peralta Soler et al., 1999; Paredes et al., 2002, 2005).

The role of cadherins and catenins in canine mammary tumours is still poorly understood (Restucci et al., 1997; Reis et al., 2003; Brunetti et al., 2005; De Matos et al., 2007). Brunetti et al. (2005) showed that reduced E-cadherin/β-catenin expression influences invasion of canine mammary tumours, but not proliferation or survival. Loss of E-cadherin in canine mammary tumours was correlated with tumour size, ulceration, lymph node metastasis, necrosis and infiltrative growth (Matos et al., 2006). Recently, we found a significant association between Pcadherin expression in canine mammary tumours and histological type (Gama et al., 2004). In the present study, we correlated the expression of E- and P-cadherin and βcatenin in a series of malignant canine mammary tumours with clinicopathological parameters, proliferation and survival to study their possible role in canine mammary tumorigenesis.

Materials and methods

Source of tumours

Sixty-five malignant canine mammary tumours were selected from the histopathological files of the University of Trás-os-Montes and Alto Douro, Vila Real and from the Institute of Biomedical Science at the University of Porto, Portugal. The material had been fixed in 10% neutral buffered formalin and embedded in paraffin wax. Sections (3 μm) were cut and stained with haematoxylin and eosin (HE) for histological examination and immunohistochemistry.

Case follow-up

Follow-up data were available for 39 cases for a mean of 15 months (range 1–36 months) after surgical treatment. Disease-free survival (DFS) and overall survival (OS) were calculated from the day of the surgery until the time of recurrence/metastasis or death, respectively. The cause of death was confirmed at post-mortem examination.

Histopathological examination

Tumours were classified according to the World Health Organization (WHO) criteria for canine mammary neoplasms (Misdorp et al., 1999) by three pathologists. Each tumour was assessed for size, skin ulceration, necrosis and mode of growth (expansile vs. infiltrative). Regional lymph nodes were available in 52 cases and assessed for the presence of metastases.

Malignant epithelial neoplasms were graded according to the Nottingham method for human breast tumours (Elston and Ellis, 1998). Tubule formation, nuclear pleomorphism and mitotic index were scored on a scale of 1–3 (slight, moderate or marked degree) and grades were based on the total score: grade I (well differentiated): 3–5 points; grade II (moderately differentiated): 6–7 points; and grade III (poorly differentiated): 8–9 points. Tumours were also graded for invasion according to Gilbertson et al. (1983): stage 0 (non-infiltrating); stage I (stromal inva-

sion); and stage II (neoplastic emboli in vessels and/or lymph node involvement).

Immunohistochemistry

Monoclonal antibodies used in the present study were anti-E-cadherin (4A2C7, 1:100, Zymed Laboratory), anti-P-cadherin (clone 56, 1:50, BD Transduction Laboratories), anti- β -catenin (CAT-5H10, 1:100, Zymed Laboratory) and anti-Ki-67 (MIB-1, 1:50, Dakocytomation). A streptavidin–biotin-peroxidase complex method was used with a commercial detection system (Ultra Vision Detection System, Lab Vision Corporation) following the manufacturer's instructions.

Antigen retrieval for E-cadherin and β -catenin was carried out by microwave treatment in a 0.05% detergent solution (Extran, Merck) and for P-cadherin with ethylene diamine tetraacetic acid (EDTA) buffer pH 8.0 (Lab Vision Corporation) in a boiling water bath for 20 min. For Ki-67 antigen retrieval, slides were incubated with 0.2 mg/mL trypsin (Merck) in phosphate buffered saline (PBS) for 10 min at 37 °C prior to microwave treatment (3 × 5 min) in 10 mM citrate buffer, pH 6.0. Adjacent normal mammary tissues were used as internal positive controls. The primary antibody was replaced with PBS for negative controls.

Quantification of immunolabelling

P-cadherin expression in canine mammary tissues was assessed semi-quantitatively according to the percentage of immunoreactive cells (cells showing a membranous and/or cytoplasmic expression pattern) in negative (0: <10%) and positive (1: 10–25%; 2: 26–50%; 3: >50%) tumours (Gama et al., 2004). E-cadherin and β -catenin immunoreactivity was classified as membranous (localised at cell–cell boundaries) or cytoplasmic (uniformly distributed through the cytoplasm, with no recognisable distinction between membrane and cytoplasm). Nuclear expression of β -catenin was only detected in normal mammary gland epithelial cells adjacent to the tumour in two cases and therefore was not scored. Cases were grouped according to Brunetti et al. (2005) as "preserved", when positivity was membranous and higher than 75% of neoplastic epithelial cells and as "reduced" in all remaining samples, including negative tumours.

Combined variables were created for immunoreactivity patterns of E-cadherin/ β -catenin (E-cad/ β -cat), E-cadherin/P-cadherin (E-cad/P-cad) and β -catenin/P-cadherin (β -cat/P-cad) to investigate a possible relationship between any reduction/lack of expression of E-cadherin/ β -catenin and over-expression of P-cadherin. Tumours were classified into four categories (+/+, +/-, -/+, -/-) for each combined variable.

Proliferative indices

Mitotic index and Ki-67 index were determined by counting 1000 neoplastic cells in the most mitotically active areas or areas with the highest Ki-67 positivity and were calculated as the percentage of tumour cells that exhibited mitotic figures or had positive staining for Ki-67, respectively.

Statistical analysis

For statistical analysis, associations between the expression of the different adhesion molecules and continuous variables (mitotic and Ki-67 indices) were assessed by analysis of variance (ANOVA). Associations between adhesion molecule expression and clinicopathological parameters (categorical variables), such as tumour size, histological type, histological grade and invasion, were performed using the χ^2 test. Fisher's exact test was performed when compared variables had exactly two groups (2 × 2 table), such as E-cadherin expression (preserved and reduced groups) versus ulceration (absent and present groups). Survival curves were generated by the Kaplan–Meyer method and the survival rates were compared using the log-rank test. All statistical analysis was performed using SPSS 11.5 statistical software. P values <0.05 were considered statistically significant.

A. Gama et al. | The Veterinary Journal 177 (2008) 45-53

Results

The 65 malignant tumours examined in this study were classified as solid carcinomas (22 cases), tubulopapillary carcinomas (12), complex carcinomas (12), carcinosarcomas (12), spindle cell carcinomas (four) and sarcomas (three) according to WHO criteria. Epithelial tumours were classified as grade I (four cases), grade II (18) and grade III (40) according to the Nottingham method.

Expression of P-cadherin was evident in all histological types, whereas expression of E-cadherin and β -catenin was negative in all sarcomas. Reduction/lack of E-cadherin expression was evident in 27 malignant tumours (41.5%) and reduction/lack of β -catenin expression in 35 (53.8%). Aberrant cytoplasmic/membranous expression of P-cadherin was found in 56 (86.2%) malignant tumours. In 13 (20%) of these 56 tumours, 10–25% of neoplastic cells were

positive; in 17 (26.2%), 26–50% were positive; and in 26 (40%) >50% were positive (Fig. 1).

Immunohistochemical expression of E-cadherin and β -catenin was significantly correlated (P < 0.0001): 26 cases were positive for both proteins, there was reduced expression of both proteins in 23 cases and 16 cases were discordant. We found no significant association between aberrant P-cadherin expression and reduced E-cadherin or β -catenin expression.

The relationship between cadherins and β -catenin expression and several clinicopathological variables is shown in Table 1. There was a significant difference across histological types in expression of E-cadherin (P=0.01). Reduced E-cadherin expression was significantly related to infiltrative tumour growth (P=0.02), higher histological grade (P<0.0001), higher degree of invasion (P=0.002) and lymph node metastasis (P=0.047). Reduced β -catenin

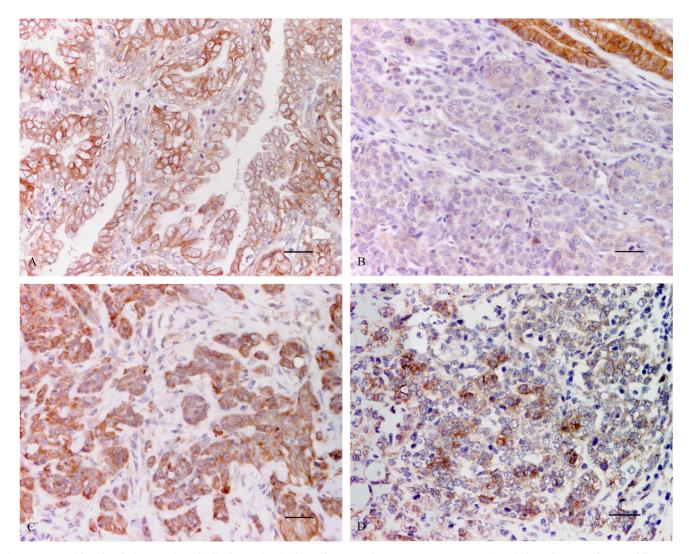


Fig. 1. Immunohistochemical expression of adhesion molecules in malignant canine mammary tumours. (A) Tubulopapillary carcinoma with preserved membranous E-cadherin expression. (B) Solid carcinoma. Neoplastic cells are negative for E-cadherin expression, whereas non-neoplastic epithelial cells are positive. (C) Solid carcinoma, with strong P-cadherin immunoreactivity. (D) Complex carcinoma showing reduced membranous β -catenin expression (Bar = 30 μ m).

Table 1 Association between E-cadherin, P-cadherin and β-catenin expression and clinicopathological parameters

Clinicopathological	n	E-cadherin		β-Catenin		P-cadherin			
parameter		Pr ^a	Rd ^b	Pr	Rd	0	1	2	3
Tumour size ^c									
<3 cm	22	14 (63.6%)	8 (36.4%)	8 (36.4%)	14 (63.6%)	4 (18.2%)	5 (22.7%)	7 (31.8%)	6 (27.3%)
3-5 cm	26	16 (61.5%)	10 (38.5%)	16 (61.5%)	10 (38.5%)	3 (11.5%)	5 (19.2%)	7 (26.9%)	11 (42.3%)
>5 cm	16	7 (43.8%)	9 (56.3%)	5 (31.3%)	11 (68.8%)	2 (12.5%)	2 (12.5%)	3 (18.8%)	9 (56.3%)
P		0.43		0.1		0.77			
Ulceration									
Absent	55	32 (58.2%)	23 (41.8%)	26 (47.3%)	29 (52.7%)	7 (12.7%)	12 (21.8%)	15 (27.3%)	21 (38.2%)
Present	10	6 (60%)	4 (40%)	4 (40%)	6 (60%)	2 (20%)	1 (10%)	2 (20%)	5 (50%)
P		0.98		0.74		0.73			
Histological type									
Solid carcinoma	22	9 (40.9%)	13 (59.1%)	8 (36.4%)	14 (63.6%)	6 (27.3%)	6 (27.3%)	5 (22.7%)	5 (22.7%)
Tubulopapillary carcinoma	12	10 (83.3%)	2 (16.7%)	8 (66.7%)	4 (33.3%)	1 (8.3%)	2 (16.7%)	3 (25%)	6 (50%)
Complex carcinoma	12	10 (83.3%)	2 (16.7%)	6 (50%)	6 (50%)	1 (8.3%)	4 (33.3%)	3 (25%)	4 (33.3%)
Carcinosarcoma	12	6 (50%)	6 (50%)	6 (50%)	6 (50%)	0	1 (8.3%)	3 (25%)	8 (66.7%)
Spindle cell carcinoma	4	3 (75%)	1 (25%)	2 (50%)	2 (50%)	0	0	2 (50%)	2 (50%)
Sarcoma	3	0	3 (100%)	0	3 (100%)	1 (33.3%)	0	1 (33.3%)	1 (33.3%)
P		0.01		0.38		0.44			
Mode of growth									
Expansile	11	10 (90.9%)	1 (9.1%)	7 (63.6%)	4 (36.4%)	2 (18.2%)	5 (45.5%)	3 (27.3%)	1 (9.1%)
Infiltrative	54	28 (51.9%)	26 (48.1%)	23 (42.6%)	31 (57.4%)	7 (13%)	8 (14.8%)	14 (25.9%)	25 (46.3%)
P		0.02		0.32		0.04			
Necrosis									
Absent	6	5 (83.3%)	1 (16.7%)	4 (66.7%)	2 (33.3%)	0	1 (16.7%)	4 (66.7%)	1 (16.7%)
Present	59	33 (55.9%)	26 (44.1%)	26 (44.1%)	33 (55.9%)	9 (15.3%)	12 (20.3%)	13 (22%)	25 (42.4%)
P		0.38		0.40		0.1			
Histological grade ^d									
Grade I	4	4 (100%)	0	4 (100%)	0	1 (25%)	1 (25%)	1 (25%)	1 (25%)
Grade II	18	17 (94.4%)	1 (5.6%)	12 (66.7%)	6 (33.3%)	2 (11.1%)	4 (22.2%)	5 (27.8%)	7 (38.9%)
Grade III	40	17 (42.5%)	23 (57.5%)	14 (35%)	26 (65%)	6 (15%)	8 (20%)	10 (25%)	16 (40%)
P		< 0.0001		0.006		0.99			
Invasion ^e									
Stage 0	11	11 (100%)	0	9 (81.8%)	2 (18.2%)	1 (9.1%)	5 (45.5%)	3 (27.3%)	2 (18.2%)
Stage I	11	8 (72.7%)	3 (27.3%)	5 (45.5%)	6 (54.5%)	1 (9.1%)	1 (9.1%)	5 (45.5%)	4 (36.4%)
Stage II	43	19 (44.2%)	24 (55.8%)	16 (37.2%)	27 (62.8%)	7 (16.3%)	7 (16.3%)	9 (20.9%)	20 (46.5%)
P		0.002		0.03		0.22			
Lymph node metastasis ^f									
Absent	27	20 (74.1%)	7 (25.9%)	13 (48.1%)	14 (5.9%)	3 (11.1%)	8 (29.6%)	9 (33.3%)	7 (25.9%)
Present	25	11 (44%)	14 (56%)	13 (52%)	12 (48%)	3 (12%)	3 (12%)	5 (20%)	14 (56%)
P		0.047		0.99		0.13			

Probability (P) values in bold are significant.

expression was also associated with high histological grade (P=0.006) and degree of invasion (P=0.03). Anomalous expression of P-cadherin was associated only with invasion (P=0.04).

When proliferation indices (mitotic index and Ki-67 index) were compared with expression patterns of the cadherins and β -catenin, there were significant differences only between higher mitotic counts and reduced expression of

E-cadherin (P=0.03) and β -catenin (P=0.003) (Table 2). No significant differences were found between P-cadherin groups.

Tumours with reduced expression of E-cadherin and β -catenin were associated with significantly shorter survival times for both DFS (E-cadherin: P = 0.0263; β -catenin: P = 0.0095) and OS (E-cadherin: P = 0.0245; β -catenin: P = 0.0113) (Fig. 2). There was no significant association

^a Preserved.

^b Reduced.

^c Tumour size was available in 64 cases.

^d According to the Nottingham method for human breast tumours (Elston and Ellis, 1998).

^e According to Gilbertson et al. (1983).

f Lymph nodes were available in 52 cases.

A. Gama et al. | The Veterinary Journal 177 (2008) 45-53

Table 2 Association between E-cadherin, P-cadherin and β -catenin expression and proliferation indices

	n	Mitotic index	Ki-67 index
			${\text{Mean} \pm \text{standard}}$ ${\text{deviation}}$
E-cadherin			
Preserved	33	0.88 ± 0.58	24.98 ± 9.84
Reduced	27	1.27 ± 0.75	29.69 ± 9.31
P		0.03	0.064
β-Catenin			
Preserved	25	0.75 ± 0.49	24.54 ± 10.59
Reduced	35	1.27 ± 0.49	28.92 ± 8.93
P		0.003	0.088
P-cadherin			
<10%	8	1.45 ± 0.84	26.85 ± 8.75
10-25%	12	0.76 ± 0.42	24.1 ± 7.52
26-50%	15	1.02 ± 0.68	27.97 ± 9.93
>50%	25	1.09 ± 0.72	28.09 ± 11.18
P		0.179	0.693

Probability (P) values in bold are significant.

between increased expression of P-cadherin and DFS or OS (Fig. 3).

When tumours were grouped according to combined patterns of expression of E-cadherin/P-cadherin, β-catenin/P-cadherin and E-cadherin/β-catenin, there were significant differences in histological grade III (P = 0.005), invasion grades I and II (P = 0.045) and lymph node metastasis (P = 0.029) for E-cad/P-cad (-/+) cases, but no significant differences for E-cad/P-cad (+/-) or E-cad/ P-cad (+/+) tumours (Supplementary Table 1). There was a significant association between β -cat/P-cad (-/+) immunoreactivity and less differentiated tumours (P =0.016) when compared to other groups (Supplementary Table 2). Loss/reduction of immunoreactivity for E-cadherin, β-catenin, or both, was significantly associated with less differentiated tumours (P = 0.003), higher invasive grade (P = 0.02) and lymph node metastases (P = 0.043) (Supplementary Table 3). No statistical differences were found with the remaining variables.

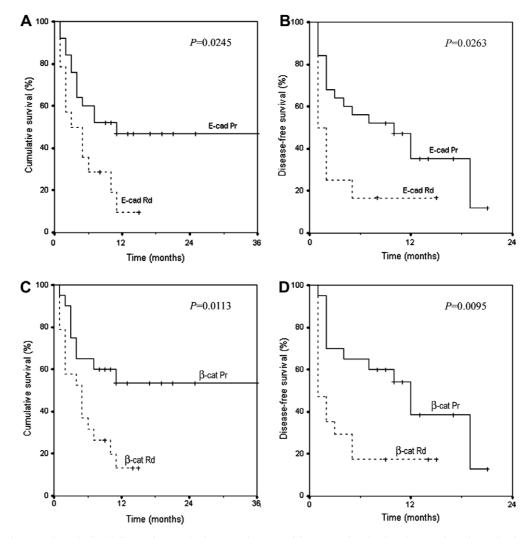
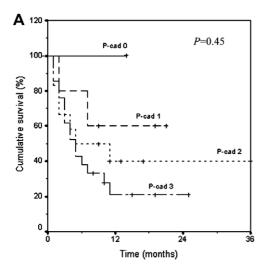


Fig. 2. Kaplan–Meier overall survival and disease-free survival curves of groups with preserved and reduced expression of E-cadherin (A, B) and β -catenin (C, D).



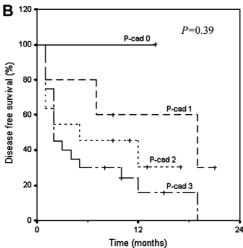


Fig. 3. Kaplan–Meier overall survival (A) and disease-free survival (B) curves of cases with <10% (0), 10-25% (1), 25-50% (2) or >50% (3) P-cadherin positive cells.

Discussion

Altered expression or absence of expression of the cadherin–catenin complex results in decreased adhesion of cells and has been implicated in tumorigenesis, particularly tumour cell invasion (Wijnhoven et al., 2000). Several previous reports have described the expression of cadherins and/or associated proteins in canine mammary gland tumours, but the patterns of expression of E-cadherin, P-cadherin and β -catenin have not been correlated with a wide range of clinicopathological features, including survival (Restucci et al., 1997; Reis et al., 2003; Gama et al., 2004; Brunetti et al., 2005; De Matos et al., 2007).

The present study demonstrated reduced membranous expression of E-cadherin and β -catenin in malignant canine mammary tumours compared to the normal mammary gland, suggesting that down-regulation of these molecules is a common event in canine mammary tumours. This finding is supported by other recent studies on canine mammary tumours (Brunetti et al., 2005; De Matos et al.,

2007) and human breast cancer (Park et al., 2007). Although P-cadherin is not expressed in normal mammary epithelial cells, a subset of canine mammary tumours exhibited aberrant P-cadherin expression, as previously reported (Gama et al., 2004).

A relationship between E-cadherin and β-catenin expression was observed in canine mammary tumours in this study, corroborating previous reports of co-expression of these adhesion molecules in canine mammary tissue (Brunetti et al., 2005; De Matos et al., 2007) and human breast tissue (Gillet et al., 2001; Yoshida et al., 2001). This finding is consistent with the formation of adhesion complexes on the cell membrane. Some human breast cancer studies have reported an inverse correlation between loss of E-cadherin and aberrant P-cadherin immunoreactivity (Palacios et al., 1995; Peralta Soler et al., 1999; Gamallo et al., 2001), whereas we found no association between P-cadherin and other adhesion molecules.

In this study, reduced membranous expression of E-cadherin was significantly associated with histological type, poor differentiation, high invasiveness, high index of proliferation and lymph node metastasis. Previous studies on canine mammary tumours have made similar observations (Restucci et al., 1997; Reis et al., 2003; Brunetti et al., 2005; De Matos et al., 2007). Together with our results, this suggests a possible role for E-cadherin-mediated adhesion in preventing invasion and metastasis in canine mammary tumours, corroborating some studies in human breast cancer (Bankfalvi et al., 1999; Madhavan et al., 2001).

However, other studies on human breast cancer have not confirmed such a relationship (Palacios et al., 1995; Bukholm et al., 1998; Kovacs et al., 2003) or have associated preservation of expression of E-cadherin with lymph node metastasis (Gillet et al., 2001; Howard et al., 2005). In contrast to the present study, one previous report on canine mammary tumours showed no association between reduced E-cadherin expression and high histological grade (Matos et al., 2006). Our study identified an association between expression of E-cadherin and mitotic index, whereas no such association was identified by Brunetti et al. (2005). In human cancer studies, we also find opposing results for differentiation (Siitonen et al., 1996; Kovacs et al., 2003; Howard et al., 2005) and proliferation (Charpin et al., 1999; Fricke et al., 2003). Sample selection (histological type, stage, tumour grade), number of cases analysed and differences in staining evaluation may individually or in combination be held responsible for the observed discrepancies between different studies.

Reduced membranous β -catenin expression was found to be significantly associated with high grade and highly invasive tumours. These findings confirm a previous study (Brunetti et al., 2005) but contradict another (De Matos et al., 2007). In the present study, we found no association between the loss of β -catenin expression and the presence of lymph node metastases, which supports similar findings in canine (De Matos et al., 2007) and human studies (Bukholm et al., 1998; Gonzalez et al., 1999; Yoshida et al.,

2001), but we did observe a significant association between β -catenin reduction and high mitotic index, in contrast to a previous study in dogs (Brunetti et al., 2005).

The prognostic significance of E-cadherin and β -catenin expression in terms of survival of dogs with mammary carcinoma is unclear. Our data show that loss/reduction of E-cadherin and β -catenin expression is significantly associated with shorter OS and DFS, in contrast to previous findings by Brunetti et al. (2005). In human breast cancer studies, assessment of down-regulation of E-cadherin and catenins as prognostic markers of breast carcinoma has also proven problematic. Some authors claim that E-cadherin and β -catenin represent valuable prognostic markers (Siitonen et al., 1996; Dollet-Filhard et al., 2006; Park et al., 2007), whereas others have found no association between these molecules and survival time (Peralta Soler et al., 1999; Yoshida et al., 2001).

These conflicting results may reflect the current poor understanding of the dynamics of cell–cell adhesion. This process seems to be regulated at various levels, including gene transcription, protein stability and post-translational modification of the cadherin/catenin complex, in particular by phosphorylation of β -catenin. Besides its function in establishing tight cell adhesion, β -catenin plays a major role in cell signalling through interactions with receptor tyrosine kinases and transcription factors of the Lef/Tcf family, suggesting a dual role as a tumour suppressor and as an oncogene in human cancers (Wijnhoven et al., 2000).

Aberrant expression of P-cadherin was associated with invasion, but not with a higher invasion grade, which also takes into account vessel and/or lymph node involvement. Our results with canine mammary malignant tumours do not reflect some in vitro studies in human breast (Paredes et al., 2004) and pancreatic (Taniuchi et al., 2005) cancer cell lines, which suggest a proinvasive role for P-cadherin, through its interaction with signalling molecules such as p120^{ctn} (Taniuchi et al., 2005).

In the present series, we did not find further associations between P-cadherin expressions and other clinicopathological variables or proliferation labelling indices. In our previous study, we described an association with tumour type (Gama et al., 2004), which was not confirmed in this larger series. In human cancer studies we also find contradictory results, probably related with sample selection. Paredes et al. (2005) found no significant correlation with histological type, although some authors suggested that P-cadherin was related with some special tumour types, such as medullary and metaplastic carcinomas (Palacios et al., 1995).

The present work supports our previous study, which did not find a statistically significant difference between P-cadherin aberrant expression and differentiation grade (Gama et al., 2004). However, in recent studies on human breast cancer, P-cadherin expression was significantly associated with increased histological grade (Palacios et al., 1995; Peralta Soler et al., 1999; Gamallo et al., 2001; Paredes et al., 2002, 2005; Kovacs et al., 2003). The small num-

ber of grade I tumours in our study may not have provided sufficient statistical power to establish an association.

Some human breast cancer studies (Palacios et al., 1995; Kovacs et al., 2003; Paredes et al., 2005) also failed to find a correlation between anomalous expression of P-cadherin and the presence of lymph node metastases. However, other studies have described an association with highly proliferative tumours (Paredes et al., 2005), lymph node metastases (Gamallo et al., 2001) and poor prognosis (Peralta Soler et al., 1999; Gamallo et al., 2001; Paredes et al., 2005). Although several authors suggested a possible role for P-cadherin in promoting aggressive tumour cell behaviour (Peralta Soler et al., 1999; Gamallo et al., 2001; Paredes et al., 2002, 2005), the biological significance of the anomalous P-cadherin in breast cancer is still poorly understood. As P-cadherin is expressed only by myoepithelial cells in normal breast tissue, the presence of this molecule might indicate a basal/myoepithelial differentiation (Peralta Soler et al., 1999), which has been associated with a poor outcome in human breast cancer (van de Rijn et al., 2002).

Despite some similarities with human breast cancer, canine mammary tumours are frequently associated with myoepithelial differentiation (Misdorp, 2002), which might explain the high percentage of P-cadherin positive tumours in our series. Although no correlation was found in the present study, we propose additional studies including a large series of simple carcinomas in order to investigate if P-cadherin expression is able to identify a subset of carcinomas with a particularly poor prognosis.

It is important to note that alterations in any component may lead to disrupted function of adhesion complexes. When we studied E-cadherin/β-catenin combinations, we found that the loss or reduction of at least one of these molecules was associated with less differentiated, highly invasive tumours, frequently with lymph node metastasis, supporting previous canine (Brunetti et al., 2005) and human studies (Bukholm et al., 1998; Wijnhoven et al., 2000). However, future studies are needed with a larger series and a longer follow up to investigate these interrelationships and correlate them with distinct tumour behaviours in the canine mammary gland.

Conclusions

The study has demonstrated altered expression of intercellular adhesion molecules in canine mammary malignant tumours. There were significant associations between reduced E-cadherin expression and some known prognostic parameters, such as tumour type, histological grade, invasiveness and lymph node metastasis and between reduced β -catenin expression and histological grade and invasiveness. Preserved expression of these molecules was associated with tumours of low mitotic index and with a better clinical outcome. We were also able to confirm aberrant P-cadherin expression in malignant mammary tumours,

which was associated with an infiltrative tumour growth, but with no relation to other clinicopathological variables.

Acknowledgements

The authors thank Mrs Lígia Bento for expert technical assistance. This work was supported by the Centro de Ciência Animal e Veterinária (CECAV) – University of Trás os Montes e Alto Douro (UTAD), Vila Real, Portugal and by Portuguese Science and Technology Foundation, Project POCTI/CVT/57795/2004.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tvjl.2007. 05.024.

References

- Bankfalvi, A., Terpe, H.J., Breukelmann, D., Bier, B., Rempe, D., Pschadka, G., Krech, R., Lelle, R.J., Boecker, W., 1999. Immunophenotypic and prognostic analysis of E-cadherin and β-catenin expression during breast carcinogenesis and tumour progression: a comparative study with CD44. Histopathology 34, 25–34.
- Brunetti, B., Sarli, G., Preziosi, R., Monari, I., Benazzi, C., 2005. E-cadherin and β-catenin reduction influence invasion but not proliferation in canine malignant mammary tumors. Veterinary Pathology 42, 781–787.
- Bukholm, I.K., Nesland, J.M., Karesen, R., Jacobsen, U., Borresen-Dale, A.L., 1998. E-cadherin and α-, β-, and γ-catenin protein expression in relation to metastasis in human breast carcinoma. Journal of Pathology 185, 262–266.
- Charpin, C., Bonnier, P., Garcia, S., Andrac, L., Crebassa, B., Dorel, M., Lavaut, M.N., Allasia, C., 1999. E-cadherin and β-catenin expression in breast medullary carcinomas. International Journal of Oncology 15, 285–292.
- De Matos, A.J., Lopes, C.C., Faustino, A.M., Carvalheira, J.G., Rutteman, G.R., Gärtner Mde, F., 2007. E-cadherin, β-catenin, invasion and lymph node metastases in canine malignant mammary tumours. Acta Pathologica, Microbiologica et Immunologica Scandinavica 115, 327–334.
- Dollet-Filhard, M., McCabe, A., Giltnane, J., Cregger, M., Cam, R.L., Rimm, D.L., 2006. Quantitative in situ analysis of β-catenin expression in breast cancer shows decreased expression is associated with poor outcome. Cancer Research 66, 5487–5494.
- Elston, C.W., Ellis, I.O., 1998. Assessment of histological grade. In: Elston, C.W., Ellis, I.O. (Eds.), Systemic Pathology. The Breast., third ed. Churchill Livingstone, London, UK, pp. 365–384.
- Fricke, E., Keller, G., Becker, I., Rosivatz, E., Schott, C., Plaschke, S., Rudelius, M., Hermannstadter, C., Busch, R., Hofler, H., Becker, K.F., Luber, B., 2003. Relationship between E-cadherin gene mutation and p53 gene mutation, p53 accumulation, Bcl-2 expression and Ki-67 staining in diffuse-type gastric carcinoma. International Journal of Cancer 104, 60–65.
- Gama, A., Paredes, J., Milanezi, M.F., Reis-Filho, J.S., Gartner, F., Schmitt, F.C., 2002. P-cadherin expression in canine lactating mammary gland. Journal of Cellular Biochemistry 86, 420–421.
- Gama, A., Paredes, J., Albergaria, A., Gartner, F., Schmitt, F.C., 2004. P-cadherin expression in canine mammary tissues. Journal of Comparative Pathology 130, 13–20.
- Gamallo, C., Moreno-Bueno, G., Sarrio, D., Calero, F., Hardisson, D., Palacios, J., 2001. The prognostic significance of P-cadherin in infiltrating ductal breast carcinoma. Modern Pathology 14, 650–654.

- Gamallo, C., Palacios, J., Suarez, A., Pizarro, A., Navarro, P., Quintanilla, M., 1993. Correlation of E-cadherin expression with differentiation grade and histological grade in breast carcinoma. American Journal of Pathology 142, 987–993.
- Gilbertson, S.R., Kurzman, I.D., Zachrau, R.E., Hurvitz, A.I., Black, M.M., 1983. Canine mammary epithelial neoplasms: biologic implications of morphologic characteristics assessed in 232 dogs. Veterinary Pathology 20, 127–142.
- Gillet, C.E., Miles, D.W., Ryder, K., Skilton, D., Liebman, R.D., Springall, R.J., Barnes, D.M., Hanby, A.M., 2001. Retention of the expression of E-cadherin and catenins is associated with shorter survival in grade III ductal carcinoma of the breast. Journal of Pathology 193, 433–441.
- Gonzalez, M.A., Pinder, S.E., Wencyk, P.M., Bell, J.A., Elston, C.W., Nicholson, R.I., Robertson, J.F., Blamey, R.W., Ellis, I.O., 1999. An immunohistochemical examination of the expression of E-cadherin, α and β / γ -catenins, and α 2- and β 1-integrins in invasive breast cancer. Journal of Pathology 187, 523–529.
- Gumbiner, B.M., 1996. Cell adhesion: the molecular basis of tissue architecture and morphogenesis. Cell 84, 345–357.
- Howard, E.M., Lau, S.K., Lyles, R.H., Birdsong, G.G., Umbreit, J.N., Kochhar, R., 2005. Expression of E-cadherin in high-risk breast cancer. Journal of Cancer Research and Clinical Oncology 131, 14–18.
- Knudsen, K.A., Frankowski, C., Johnson, K.R., Wheelock, M.J., 1998. A role for cadherins in cellular signaling and differentiation. Journal of Cellular Biochemistry 30–31 (Suppl.), 168–176.
- Kovacs, A., Dhillon, J., Walker, R.A., 2003. Expression of P-cadherin, but not E-cadherin or N-cadherin, relates to pathological and functional differentiation of breast carcinomas. Molecular Pathology 56, 318–322.
- Madhavan, M., Srinivas, P., Abraham, E., Ahmed, I., Mathew, A., Vijayalekshmi, N.R., Balaram, P., 2001. Cadherins as predictive markers of nodal metastasis in breast cancer. Modern Pathology 14, 423–427.
- Matos, A.J.F., Lopes, C., Carvalheira, J., Santos, M., Rutteman, G.R., Gartner, F., 2006. E-cadherin expression in canine malignant mammary tumours: relationship to other clinico-pathological variables. Journal of Comparative Pathology 134, 182–189.
- Misdorp, W., Else, R.W., Hellmén, E., Lipscomb, T.P., 1999. Histological Classification of Mammary Tumors of the Dog and the Cat, 2nd series, Vol VII, Armed Forces Institute of Pathology, American Registry of Pathology, Washington D.C. and the World Health Organization Collaborating Center for Worldwide Reference on Comparative Oncology, pp. 1–59.
- Misdorp, W., 2002. Tumours of the mammary gland. In: Meuten, D.J. (Ed.), Tumors in Domestic Animals, fourth ed. Iowa State Press, Blackwell Publishing Company, pp. 575–606.
- Nose, A., Takeichi, M., 1986. A novel cadherin cell adhesion molecule: its expression patterns associated with implantation and organogenesis of mouse embryos. Journal of Cell Biology 103, 2649–2658.
- Palacios, J., Benito, N., Pizarro, A., Suarez, A., Espada, J., Cano, A., Gamallo, C., 1995. Anomalous expression of P-cadherin in breast carcinoma: correlation with E-cadherin expression and pathological features. American Journal of Pathology 146, 605–612.
- Paredes, J., Milanezi, F., Viegas, L., Amendoeira, I., Schmitt, F., 2002. P-cadherin expression is associated with high-grade ductal carcinoma in situ of the breast. Virchows Archiv 440, 16–21.
- Paredes, J., Stove, C., Stove, V., Milanezi, F., van Marck, V., Derycke, L., Mareel, M., Bracke, M., Schmitt, F., 2004. P-cadherin is up-regulated by the antiestrogen ICI 182,780 and promotes invasion of human breast cancer cells. Cancer Research 64, 8309–8317.
- Paredes, J., Albergaria, A., Oliveira, J., Jerónimo, C., Milanezi, F., Schmitt, F., 2005. P-Cadherin overexpression is an Indicator of clinical outcome in invasive breast carcinomas and is associated with CDH3 promoter hypomethylation. Clinical Cancer Research 11, 5869–5877.
- Park, D., Kåresen, R., Axcrona, U., Noren, T., Sauer, T., 2007. Expression pattern of adhesion molecules (E-cadherin, α-, β-, γ-catenin and claudin-7), their influence on survival in primary breast

- carcinoma, and their corresponding axillary lymph node metastasis. Acta Pathologica, Microbiologica et Immunologica Scandinavica 115, 52–65.
- Peralta Soler, A., Knudsen, K.A., Salazar, H., Han, A.C., Keshgegian, A.A., 1999. P-cadherin expression in breast carcinoma indicates poor survival. Cancer 86, 1263–1272.
- Reis, A.L., Carvalheira, J., Schmitt, F.C., Gärtner, F., 2003. Immunohistochemical study of E-cadherin expression in canine mammary tumours. Veterinary Record 152, 621–624.
- Restucci, B., Papparella, S., De Vico, G., Maiolino, P., 1997. E cadherin expression in normal and neoplastic canine mammary gland. Journal of Comparative Pathology 116, 191–202.
- Shimoyama, Y., Hirohashi, S., Hirano, S., Noguchi, M., Shimosato, Y., Takeichi, M., Abe, O., 1989. Cadherin cell adhesion molecules in human epithelial tissues and carcinomas. Cancer Research 49, 2128– 2133.
- Siitonen, S.M., Kononen, J.T., Helin, H.J., Rantala, I.S., Holli, K.A., Isola, J.J., 1996. Reduced E-cadherin expression is associated with invasiveness and unfavourable prognosis in breast cancer. American Journal of Clinical Pathology 105, 394–402.

- Taniuchi, K., Nakagawa, H., Hosokawa, M., Nakamura, T., Eguchi, H.,
 Ohigashi, H., Ishikawa, O., Katagiri, T., Nakamura, Y., 2005.
 Overexpressed P-cadherin/CDH3 promotes motility of pancreatic cancer cells by interacting with p120^{ctn} and activating rho-family GTPases. Cancer Research 65, 3092–3099.
- Takeichi, M., 1991. Cadherin cell adhesion receptors as a morphogenetic regulator. Science 251, 1451–1455.
- Yoshida, R., Kimura, N., Harada, Y., Ohuchi, N., 2001. The loss of E-cadherin, α- and β-catenin expression is associated with metastasis and poor prognosis in invasive breast cancer. International Journal of Oncology 18, 513–520.
- van de Rijn, M., Perou, C.M., Tibshirani, R., Haas, P., Kallioniemi, O., Kononen, J., Torhorst, J., Sauter, G., Zuber, M., Köchli, O.R., Mross, F., Dieterich, H., Seitz, R., Ross, D., Botstein, D., Brown, P., 2002.
 Expression of cytokeratins 17 and 5 identifies a group of breast carcinomas with poor clinical outcome. American Journal of Pathology 161, 1991–1996.
- Wijnhoven, B.P.L., Dinjens, W.N.M., Pignatelli, M., 2000. E-cadherin– catenin cell–cell adhesion complex and human cancer. British Journal of Surgery 87, 992–1005.

Supplementary Table 1 Association between E-cad/P-cad combined expression and clinicopathological parameters.

Clinicopathological parameter	n		E-c	ad/P-cad	
	11	+/+	+/-	_/+	-/-
Tumour size ^a					
<3 cm	22	12 (54.5%)	2 (9.1%)	6 (27.3%)	2 (9.1%)
3-5 cm	26	15 (57.7%)	1 (3.8%)	8 (30.8%)	2 (7.7%)
>5 cm	16	6 (37.5%)	1 (6.3%)	8 (50%)	1 (6.3%)
P		0.83			
Ulceration					
Absent	55	29 (52.7%)	3 (5.5%)	19 (34.5%)	4 (7.3%)
Present	10	5 (50%)	1 (10%)	3 (30%)	1 (10%)
P		0.99			
Histological type					
Solid carcinoma	22	7 (31.8%)	2 (9.1%)	9 (40.9%)	4 (18.2%)
Tubulopapillary carcinoma	12	9 (75%)	1 (8.3%)	2 (16.7%)	0
Complex carcinoma	12	9 (75%)	1 (8.3%)	2 (16.7%)	0
Carcinosarcoma	12	6 (50%)	0	6 (50%)	0
Spindle cell carcinoma	4	3 (75%)	0	1 (25%)	0
Sarcoma	3	0	0	2 (66.7%)	1 (33.3%)
P		0.13		, ,	· · ·
Mode of growth					
Expansive	11	9 (81.8%)	1 (9.1%)	0	1 (9.1%)
Infiltrative	54	25 (46.3%)	3 (5.6%)	16 (40.7%)	4 (7.4%)
P		0.07	, , ,	•	
Necrosis					
Absent	6	5 (83.3%)	0	1 (16.7%)	0
Present	59	29 (49.2%)	4 (6.8%)	21 (35.6%)	5 (8.5%)
P		0.34	,	,	,
Histological grade ^b					
Grade I	4	3 (75%)	1 (25%)	0	0
Grade II	18	16 (88.9%)	1 (5.6%)	0	1 (5.6%)
Grade III	40	15 (37.5%)	2 (5%)	19 (47.5%)	4 (10%)
P		0.005	` '	, ,	, ,
Invasion ^c					
Stage 0	11	10 (90.9%)	1 (9.1%)	0	0
Stage I	11	7 (63.6%)	1 (9.1%)	3 (27.3%)	0
Stage II	43	17 (39.5%)	2 (4.7%)	19 (44.2%)	5 (11.6%)
P		0.045	()	· · · · · ·	(/*)
Lymph node metastasis ^d					
Absent	27	17 (63%)	3 (11.1%)	7 (25.9%)	0
Present	25	11 (44%)	0	11 (44%)	3 (12%)
P		0.029	-	(''')	- (,0)

^a Tumour size was available in 64 cases; ^b According to the Nottingham method for human breast tumours (Elston and Ellis, 1998); ^c According to Gilbertson *et al.* (1983); ^d Lymph nodes were available in 52 cases

Supplementary Table 2 Association between β -cat/P-cad combined expression and clinicopathological parameters.

Clinicopathological			β-cat/	P-cad	
parameter	n	+/+	+/-	_/+	-/-
Tumour size ^a					
<3 cm	22	8 (36.4%)	0	10 (45.5%)	4 (18.2%)
3-5 cm	26	14 (53.8%)	3 (11.5%)	8 (30.8%)	1 (14.3%)
>5 cm	16	5 (31.3%)	0	9 (56.3%)	2 (12.5%)
P		0.13			
Ulceration					
Absent	55	24 (43.6%)	3 (5.5%)	23 (41.8%)	5 (9.1%)
Present	10	4 (40%)	0	4 (40%)	2 (20%)
P		0.76			
Histological type					
Solid carcinoma	22	7 (31.8%)	1 (4.5%)	9 (40.9%)	5 (22.7%)
Tubulopapillary carcinoma	12	7 (58.3%)	1 (8.3%)	4 (33.3%)	0
Complex carcinoma	12	6 (50%)	0	5 (41.7%)	1 (8.3%)
Carcinosarcoma	12	6 (50%)	0	6 (50%)	0
Spindle cell carcinoma	4	2 (50%)	0	2 (50%)	0
Sarcoma	3	0	1 (33.3%)	1 (33.3%)	1 (33.3%)
P		0.26			
Mode of growth					
Expansive	11	7 (63.6%)	0	2 (18.2%)	2 (18.2%)
Infiltrative	54	21 (38.9%)	3 (5.6%)	25 (46.3%)	5 (9.3%)
P		0.23			
Necrosis					
Absent	6	4 (66.7%)	0	2 (33.3%)	0
Present	59	24 (40.7%)	3 (5.1%)	25 (42.4%)	7 (11.9%)
P		0.69			
Histological grade ^b					
Grade I	4	3 (75%)	1 (25%)	0	0
Grade II	18	12 (66.7%)	0	4 (22.2%)	2 (11.1%)
Grade III	40	13 (32.5%)	1 (2.5%)	21 (52.5%)	5 (12.5%)
P		0.016			
Invasion ^c					
Stage 0	11	9 (81.8%)	0	1 (9.1%)	1 (9.1%)
Stage I	11	4 (36.4%)	1 (9.1%)	6 (54.5%)	0
Stage II	43	15 (34.9%)	2 (4.7%)	20 (46.5%)	6 (14%)
P		0.095			
Lymph node metastasis ^d					
Absent	27	12 (44.4%)	1 (3.7%)	12 (44.4%)	2 (7.4%)
Present	25	12 (48%)	2 (8%)	9 (42.9%)	2 (8%)
P		0.89			

^a Tumour size was available in 64 cases; ^b According to the Nottingham method for human breast tumours (Elston and Ellis, 1998); ^c According to Gilbertson *et al.* (1983); ^d Lymph nodes were available in 52 cases

Supplementary Table 3 Association between E-cad/ β -cat combined expression and clinicopathological parameters.

Clinicopathological	n		E-cad	/β-cat	
parameter	n	+/+	+/-	_/+	-/-
Tumour size ^a					
<3 cm	22	8 (36.4%)	6 (27.3%)	0	8 (36.4%)
3-5 cm	26	13 (50%)	3 (11.5%)	3 (11.5%)	7 (26.9%)
>5 cm	16	4 (25%)	3 (18.8%)	1 (6.3%)	8 (50%)
P		0.31			
Ulceration					
Absent	55	22 (40%)	10 (18.2%)	4 (7.3%)	19 (34.5%)
Present	10	4 (40%)	2 (20%)	0	4 (40%)
P		0.92			
Histological type					
Solid carcinoma	22	5 (22.7%)	4 (18.2%)	3 (13.6%)	10 (45.5%)
Tubulopapillary carcinoma	12	8 (66.7%)	2 (16.7%)	0	2 (16.7%)
Complex carcinoma	12	6 (50%)	4 (33.3%)	0	2 (16.7%)
Carcinosarcoma	12	5 (41.7%)	1 (8.3%)	1 (8.3%)	5 (41.7%)
Spindle cell carcinoma	4	2 (50%)	1 (25%)	0	1 (25%)
Sarcoma	3	0	0	0	3 (100%)
P		0.23			
Mode of growth					
Expansile	11	7 (63.6%)	3 (27.3%)	0	1 (9.1%)
Infiltrative	54	19 (35.2%)	9 (16.7%)	4 (7.4%)	22 (40.7%)
P		0.11			
Necrosis					
Absent	6	4 (66.7%)	1 (16.7%)	0	1 (16.7%)
Present	59	22 (37.3%)	11 (18.6%)	4 (6.8%)	22 (37.3%)
P		0.56			
Histological grade ^b					
Grade I	4	4 (100%)	0	0	0
Grade II	18	12 (66.7%)	5 (27.8%)	0	1 (5.6%)
Grade III	40	10 (25%)	7 (17.5%)	4 (10%)	19 (47.5%)
P		0.003			
Invasion ^c					
Stage 0	11	9 (81.8%)	2 (18.2%)	0	0
Stage I	11	5 (45.5%)	3 (27.3%)	0	3 (27.3%)
Stage II	43	12 (27.9%)	7 (16.3%)	4 (9.3%)	20 (46.5%
P		0.02			
Lymph node metastasis ^d					
Absent	27	13 (48.1%)	7 (25.9%)	0	7 (25.9%)
Present	25	9 (42.9%)	2 (8%)	4 (16%)	10 (40%)
P		0.043			

^a Tumour size was available in 64 cases. ^b According to the Nottingham method for human breast tumours (Elston and Ellis, 1998); ^c According to Gilbertson *et al.* (1983); ^d Lymph nodes were available in 52 cases.

Chapter IV

Immunohistochemical expression of Epidermal Growth Factor Receptor (EGFR) in canine mammary tissues. Gama A, Gärtner F, Alves A, Schmitt F (submitted)

Immunohistochemical expression of Epidermal Growth Factor Receptor (EGFR) in canine mammary tissues

Gama A^a, Gärtner F^{b,c}, Alves A^a, Schmitt F^{b,d*}

^a Department of Veterinary Sciences, CECAV, University of Trás-os-Montes and Alto Douro

(UTAD), 5001-811 Vila Real, Portugal

^b Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP),

4200-465 Porto, Portugal

^c Instituto de Ciências Biomédicas de Abel Salazar, University of Porto, 4099-003 Porto,

Portugal

^d Medical Faculty, University of Porto, Porto, Portugal

* Corresponding author:

Tel.: 351-22-557 07 00; fax: 351-22-557 07 99.

E-mail address: fernando.schmitt@ipatimup.pt (F. Schmitt).

Abstract

Epidermal Growth Factor Receptor (EGFR) has been extensively studied in human breast cancer; however, systematic studies of EGFR protein expression in canine mammary gland tumours are lacking. Therefore, we evaluated its immunohistochemical

expression in a series of 136 canine mammary tumours and representative areas of

adjacent normal and hyperplastic mammary tissue and investigated a possible

correlation between EGFR overexpression and several clinicopathological parameters

and survival. In normal and hyperplastic canine mammary glands, EGFR expression

was consistently observed in myoepithelial cells, with luminal cells usually negative. In

tumour tissues, EGFR overexpression was found in 9 benign (19.6%) and 38 malignant

(42.2%) lesions, with EGFR positivity significantly related with malignancy. Besides

animal age and tumour size, there were no significant associations between other

clinicopathological parameters and EGFR overexpression. On survival analysis,

tumours with EGFR overexpression showed a reduced disease-free and overall survival;

however these associations failed to reach statistically significant levels.

Keywords: canine; mammary tumours; EGFR; immunohistochemistry

107

Introduction

Epidermal Growth Factor Receptor-1 (EGFR) is a member of the human epidermal growth factor receptor (HER) family. This family includes four closely related tyrosine kinase receptors (EGFR or erbB1, HER-2/neu or erbB2, HER-3 or erbB3, and HER-4 or erbB4) and has been receiving great attention in recent human literature (Hynes and Lane, 2005; Park *et al.*, 2007; Sassen *et al.*, 2008).

EGFR overexpression has been found in 16-48% of human breast cancer, generally associated with poor clinical outcome and aggressive biological properties (Sainsbury *et al.*, 1985; Lewis *et al.*, 1990; Klijn *et al.*, 1992; Toi *et al.*, 1994; Tsutsui *et al.*, 2002). The interest in EGFR is further enhanced by the availability and US Food and Drug Administration approval of specific EGFR tyrosine kinase inhibitors, which are currently being tested in human patients with lung and breast cancer (Baselga and Arteaga, 2005).

Mammary gland tumors are the most commonly occurring neoplasm in the female dog (Nerurkar *et al.*, 1989; Misdorp, 2002) but despite their high incidence and clinical importance, a few number of studies are available with respect to EGFR. In addition, although EGFR has been identified by several methodologies in canine mammary gland tissues such as radioligand binding or immunoenzymatic assays, to the best of our knowledge there are currently no data addressing the analysis of EGFR expression by immunohistochemistry. Therefore, the aim of the present study was to assess EGFR immunohistochemical expression in normal, benign and malignant canine mammary gland tissues and correlate its expression with clinicopathological parameters and survival, in order to provide some information on its biological significance and potential diagnostic use.

Material and methods

Tumour specimens

A hundred and thirty six cases of canine benign (n=46) and malignant (n=90) mammary tumours were selected from the histopathological files of the University of Trás-os-Montes and Alto Douro, Vila Real, and from the Institute of Biomedical Science of the

University of Porto, Portugal. The material was fixed in 10% neutral formalin and embedded in paraffin wax. Sections (3 µm) were cut and stained with haematoxylin and eosin (HE) for histological examination, or used to perform immunohistochemistry. Sixty four malignant tumour cases had available follow up data. Dogs were followed after surgical treatment, with a mean follow-up period of 13 months (range, 5-74 months). Disease-free survival (DFS) and overall survival (OS) were calculated from the day of the surgery until the time of recurrence/metastasis or death, respectively.

Histological Examination

Tumours were diagnosed according to the World Health Organization criteria for canine mammary neoplasms (Misdorp *et al.*, 1999). Clinicopathological variables included in the present study were: age, breed, presence of ovariohysterectomy, contraceptive administration, tumour size, tumour location, presence of skin ulceration, tumour histological type, presence of intra-tumoral necrosis, mode of growth (expansive vs. infiltrative), presence of stromal and vascular invasion and presence of lymph node metastasis.

Malignant tumours were also graded histologically in accordance with the Nottingham method for human breast tumours (Elston and Ellis, 1998), based on the assessment of three morphological features: tubule formation, nuclear pleomorphism and mitotic counts. Each of these features was scored on a scale of 1 to 3 to indicate whether it was present in slight, moderate or marked degree, giving a putative total of 3-9 points. Grade was allocated by an arbitrary division of the total points as follows: grade I (well differentiated), 3, 4 or 5 points; grade II (moderately differentiated), 6 or 7 points; and grade III (poorly differentiated), 8 or 9 points.

Immunohistochemistry

Immunohistochemistry was performed with a mouse monoclonal antibody raised against EGFR (Clone 31G7, 1:50, Zymed Laboratories, San Francisco, California, USA). Antigen retrieval was carried out by enzyme digestion: sections were incubated with 0.4% pepsin (Dako, Denmark) in HCl 0.01 N solution (pH=2) for 30 minutes at 37°C. Slides were incubated overnight with EGFR antibody in a humid chamber at 4°C.

A polymeric labelling methodology was used as a detection system (Novolink Polymer Detection System, Novocastra, Newcastle, United Kingdom), following the manufacturer's instructions. Adjacent normal mammary tissues were used as internal positive controls. Negative controls were carried out by replacing the primary antibody with PBS.

Quantification of Immunolabelling

To evaluate EGFR expression in canine mammary tissues Herceptest scoring system was applied (0=no membrane staining or <10% of cells stained; 1+=incomplete membrane staining in >10% of cells; 2+=>10% of cells with weak to moderate complete membrane staining; and 3+=strong and complete membrane staining in >10% of cells) (Reis Filho *et al.*, 2005), with 2+ and 3+ cases considered positive.

Statistical Analysis

To compare variables of interest χ^2 and Fisher's exact test (two-sided) were used when appropriate. The Kaplan-Meier method was used to examine OS and DFS curves, and comparisons of these curves were assessed by using the log-rank test. Analysis was performed by SPSS 11.5 software statistics and two values were considered significant when P < 0.05.

Results

In this study, the 46 benign tumours examined were classified as: benign mixed tumours (22 cases), complex adenomas (13 cases) and basaloid adenomas (11 cases) and the 90 malignant tumours were classified as: complex carcinomas (30 cases), solid carcinomas (21 cases), carcinosarcomas (13 cases), tubulopapillary carcinomas (12 cases), carcinoma in benign tumours (7 cases), spindle cell carcinomas (4 cases) and anaplastic carcinomas (3 cases). According to the Nottingham method, malignant tumours were classified as grade I (17 cases), grade II (30 cases) and grade III (43 cases).

In normal and hyperplastic canine mammary ducts and lobules, a strong EGFR expression was consistently observed in the myoepithelial cell layer. Luminal epithelial

cells were usually negative, with the exception of some normal and hyperplastic ducts showing EGFR expression in both epithelial layers, results which are in accordance to previous human studies (Santini *et al.*, 2002). The surrounding stroma was usually EGFR negative; however, perilobular stroma was commonly positive.

In benign tumours, EGFR expression was observed in both epithelial cell components; however, luminal epithelial cells usually showed a reduced level of expression (score 0 and 1+). On the other hand, the malignant tumours analysed showed a significant higher proportion of EGFR epithelial expression (Fig. 1). In fact, a complete membrane EGFR immunostaining was observed in more than 10 % of neoplastic cells (score 2+ and 3+) in 38 out of 90 (42.2%) malignant tumours, versus 9 out of 46 (19.6%) benign lesions.

The relationship between EGFR immunoexpression and clinicopathological parameters are shown in Tables 1 and 2, respectively. Benign and malignant lesions differed significantly (P=0.013) when considering EGFR expression. It was also found a significant association between EGFR expression and animal age (P=0.028) and tumour size (P=0.013). There was no significant association with all other tested variables (P>0.05).

Malignant tumours with EGFR overexpression (score 2+ and 3+) were usually related with shorter survival times for both disease free survival and overall survival. However, despite the differences observed, these associations failed to reach statistically significant levels (Disease free survival: P=0.08; Overall survival: P=0.09) (Fig. 2).



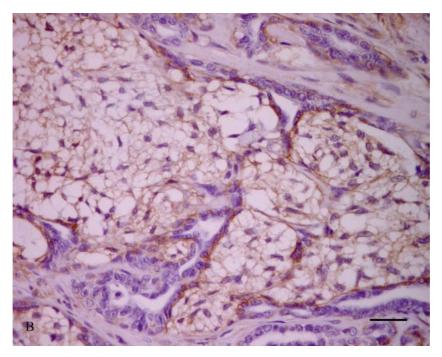
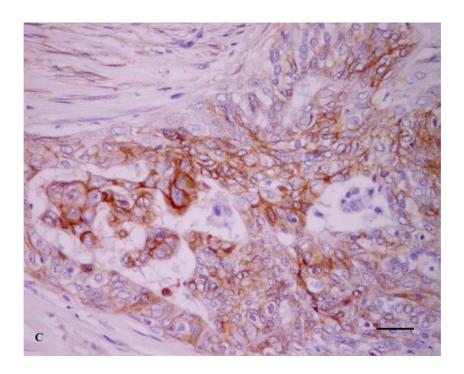


Fig. 1. Immunohistochemical expression of EGFR in canine mammary tissues. A. Normal mammary gland showing EGFR immunoexpression in the myoepithelial cell layer. Epithelial cells were negative; bar=60 μ m; B. Complex adenoma with myoepithelial spindle and stellate-shaped cells showing EGFR expression. Epithelial cells are unreactive (0 score); bar=30 μ m.



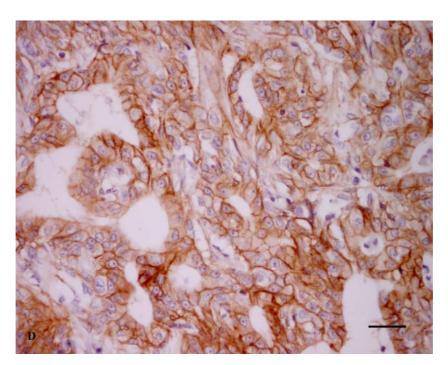


Fig. 1. (cont.) Immunohistochemical expression of EGFR in canine mammary tissues. C. Complex carcinoma, with epithelial neoplastic cells EGFR positive (2+ score); D. Tubulopapillary carcinoma showing epithelial neoplastic cells strongly positive for EGFR (3+ score); bar=30 μ m.

Table 1: Association between EGFR expression and clinical parameters.

Clinical parameters	n	EC	FR
The Property of the Property o		Negative	Positive
Age			
≤ 9 years old	69	51 (73.9%)	18 (26.1%)
> 9 years old	60	33 (55%)	27 (45%)
P		0.028	
Breed			
Mixed breed	42	27 (64.3%)	15 (35.7%)
Poodle	25	19 (76%)	6 (24%)
Cocker spaniel	15	11 (73.3%)	4 (26.7%)
Boxer	6	5 (83.3%)	1 (16.7%)
Labrador retriever	6	4 (66.7%)	2 (33.3%)
Others	34	19 (55.9%)	15 (44.1%)
P		0.588	
Tumour size			
<3 cm	74	57 (77%)	17 (23%)
3-5 cm	33	19 (57.6%)	14 (42.4%)
>5 cm	25	12 (48%)	13 (52%)
P		0.013	
Tumour location			
Cranial glands (1 and 2)	6	4 (66.7%)	2 (33.3%)
Medial gland (3)	9	5 (55.6%)	4 (44.4%)
Caudal glands (4 and 5)	45	30 (66.7%)	15 (33.3%)
Multiple	15	8 (53.3%)	7 (46.7%)
P		0.837	
Skin ulceration			
Absent	120	81 (67.5%)	39 (32.5%)
Present	13	6 (46.2%)	7 (53.8%)
P		0.137	
Ovariohysterectomy			
No	60	34 (56.7%)	26 (43.3%)
Yes, prior to tumour development	12	8 (66.7%)	4 (33.3%)
Yes, performed with mastectomy	14	10 (71.4%)	4 (28.6%)
P		0.554	
Contraception			
No	62	39 (62.9%)	23 (37.1%)
Yes	8	4 (50%)	4 (50%)
P		0.702	

Table 2: Association between EGFR expression and pathological parameters.

Pathological parameters	n	E	GFR
		Negative	Positive
Histological diagnoses			
Benign lesions	46	37 (80.4%)	9 (19.6%)
Malignant lesions	90	52 (57.8%)	38 (42.2%)
P		0.013	
Histological type			
Benign mixed tumour	22	18 (81.8%)	4 (18.2%)
Complex adenoma	13	10 (76.9%)	3 (23.1%)
Basaloid adenoma	11	9 (81.8%)	2 (18.2%)
Solid carcinoma	21	14 (66.7%)	7 (33.3%)
Tubulopapillary carcinoma	12	6 (50%)	6 (50%)
Complex carcinoma	30	16 (53.3%)	14 (46.7%)
Carcinosarcoma	13	9 (69.2%)	4 (30.8%)
Spindle cell carcinoma	4	1 (25%)	3 (75%)
Carcinoma in benign tumour	7	5 (71.4%)	2 (28.6%)
Anaplastic carcinoma	3	1 (33.3%)	2 (66.7%)
P		0.192	
Necrosis			
Absent	39	29 (74.4%)	10 (25.6%)
Present	97	60 (61.9%)	37 (38.1%)
P		0.231	
Mode of growth			
Expansive	75	52 (69.3%)	23 (30.7%)
Infiltrative	61	37 (60.7%)	24 (39.3%)
P		0.365	
Histological grade			
Grade I	17	10 (58.8%)	7 (41.2%)
Grade II	30	16 (53.3%)	14 (46.7%)
Grade III	43	26 (60.5%)	17 (39.5%)
P		0.85	
Stromal Invasion			
Absent	24	12 (50%)	12 (50%)
Present	65	39 (60%)	26 (40%)
P		0.472	
Lymphovascular Invasion			
Absent	41	22 (53.7%)	19 (46.3%)
Present	47	29 (61.7%)	18 (38.3%)
P		0.519	
Lymph node metastasis			
Absent	23	12 (52.2%)	11 (47.8%)
Present	23	13 (56.5%)	10 (43.5%)
P	•	0.99	

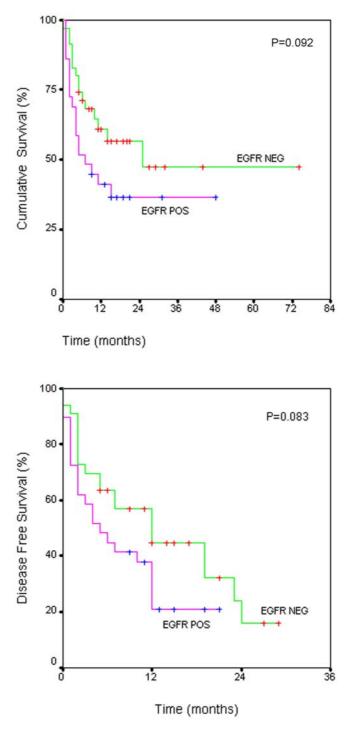


Fig. 2. Kaplan-Meier overall survival (A) and disease-free survival (B) curves of groups with negative (0 and 1+ scores) and positive (2+ and 3+ scores) EGFR expression.

Discussion

EGFR was the first tyrosine kinase transmembrane receptor to be directly linked with human cancer (Hynes and Lane, 2005). EGFR is a member of the human epidermal growth factor receptor family that consists of an extracellular domain binding EGF or TGF-α, a short transmembrane domain and an intracellular domain carrying tyrosine kinase activity (Carpenter and Cohen, 1979). EGFR pathway contributes to several processes involved in tumour survival and growth, including cell proliferation/differentiation, angiogenesis and metastasis, which make this molecule an attractive target for cancer prevention and treatment (Shien et al., 2005).

While the expression of EGFR gene has been extensively investigated in human breast cancer (Klijn *et al.*, 1992; Bhargava *et al.*, 2005; Reis Filho *et al.*, 2005; Reis-Filho *et al.*, 2006; Park *et al.*, 2007), there are only a limited number of reports available in canine literature demonstrating its presence in mammary gland tissues (Nerurkar *et al.*, 1987; Donnay *et al.*, 1993; Rutteman *et al.*, 1994; Donnay *et al.*, 1996; Matsuyama *et al.*, 2001). These previous reports describe EGFR concentration or expression in normal and tumorous canine mammary gland based on distinct methodologies, with a lack of information about its immunohistochemical pattern of cellular distribution and on the possible correlation between EGFR expression and a wide range of clinicopathological features, including survival.

The present study demonstrates that myoepithelial cells in normal, hyperplastic and benign lesions constantly express EGFR, similarly to previous studies in human breast (Moller *et al.*, 1989; Santini *et al.*, 2002). Thus, alike human breast tissues, EGFR immunoreactivity appears to be of diagnostic use for myoepithelial cell identification in canine mammary gland, in addition to specific immunohistochemical markers such as p63, P-cadherin, α-smooth muscle actin, CK14 or calponin (Destexhe *et al.*, 1993; Espinosa de Los Monteros *et al.*, 2002; Gama *et al.*, 2003; Gama *et al.*, 2004). According to several authors, EGFR myoepithelial expression in human breast tissues can be related to the recently recognized paracrine function by which myoepithelial cells exert their mechanical and functional barrier role in the juxtaposition between epithelium and stroma (Moller *et al.*, 1989; Santini *et al.*, 2002).

Nevertheless, despite our and previous demonstration of EGFR presence in the normal canine mammary gland (Donnay *et al.*, 1993; Rutteman *et al.*, 1994; Donnay *et al.*, 1996), the research performed by Matsuyama and coworkers (2001) failed to find EGFR mRNA in normal canine mammary tissues, discrepancy that might be related with the different techniques used in each study.

In the present series, EGFR overexpression (2+ and 3+ scores) was found in 9/46 (19.6%) benign and 38/90 (42.2%) malignant tumours, with its expression significantly related with malignancy (*P*=0.013). These results are in contradiction with previous reports, which found no significant differences on EGFR concentration between normal, benign and malignant canine mammary tissues (Nerurkar *et al.*, 1987; Rutteman *et al.*, 1994; Donnay *et al.*, 1996). However, because the methods used in those studies were different from ours, a direct comparison is difficult.

EGFR overexpression was only found to be associated with old aged animals and large sized tumours. No association was observed with other clinicopathological parameters, which is at some point in accordance to Donnay *et al.* (1993), who described no significant differences between EGFR concentrations and the clinicopathological parameters evaluated, including animal age, tumour location and histology. Similarly, there are contradictory results in human literature with respect to EGFR relationship with known prognostic factors (Fox *et al.*, 1994; Toi *et al.*, 1994; Pirinen *et al.*, 1995; Reis-Filho *et al.*, 2006).

The underlying mechanisms of EGFR protein overexpression are not completely understood. Gene amplification has been recently described in human breast carcinomas and *EGFR* activating mutations were also found, although uncommon (Al-Kuraya *et al.*, 2004; Bhargava *et al.*, 2005; Reis-Filho *et al.*, 2005; Reis-Filho *et al.*, 2006). It is likely that, in the majority of cases, EGFR up-regulation happens at the transcriptional level (Kersting *et al.*, 2004). Given that this gene is consistently expressed in normal myoepithelial cells (Santini *et al.*, 2002) and in human breast tumours with basal and/or myoepithelial differentiation (Nielsen *et al.*, 2004; Shien *et al.*, 2005; Reis-Filho *et al.*, 2006), some authors argue that EGFR overexpression would constitute the maintenance of a myoepithelial phenotype or would be part of a transcriptomic programme of myoepithelial/"basal-like" differentiation (Reis-Filho *et al.*, 2005). Canine mammary gland tumours are frequently associated with myoepithelial differentiation (Misdorp,

2002), and this fact is probably related with the relatively high percentage of EGFR positive tumours in our series.

EGFR is a recent tumour marker whose prognostic value has been well studied in humans, but still remains controversial (Klijn *et al.*, 1992). Although some studies involving immunohistochemical analysis of EGFR expression in human breast tumours have shown a significant correlation of EGFR positivity with a shorter disease-free and overall survival (Tsutsui *et al.*, 2002), others did not found such a correlation, and no consensus has been reached on the role of EGFR as a prognostic indicator (Ciardiello and Tortora, 2003).

In the present series, there was no association between EGFR overexpression and DFS or OS, whereas there was a tendency toward shorter DFS and OS in dogs with positive EGFR expression. In order to make definitive conclusions about the relationships and prognostic value of EGFR status in canine mammary tumours, larger studies with long term follow-up periods are warranted. In addition, studies designed to unravel the mechanism associated with EGFR overexpression in canine malignant tumours are also justified, considering that EGFR tyrosine kinase inhibitors might be used as potential therapeutic agents.

Acknowledgments

The authors thank Mrs Lígia Bento for expert technical assistance. This work was supported by the Centro de Ciência Animal e Veterinária (CECAV) – University of Trás-os-Montes e Alto Douro (UTAD), Vila Real, Portugal and by Portuguese Science and Technology Foundation, project POCTI/CVT/57795/2004.

References

- Al-Kuraya K, Schraml P, Torhorst J, Tapia C, Zaharieva B, Novotny H, Spichtin H, Maurer R, Mirlacher M, Köchli O, Zuber M, Dieterich H, Mross F, Wilber K, Simon R, Sauter G. (2004). Prognostic relevance of gene amplifications and coamplifications in breast cancer. Cancer Res 64:8534-8540.
- Baselga J, Arteaga CL. (2005). Critical update and emerging trends in epidermal growth factor receptor targeting in cancer. J Clin Oncol 23:2445-2459.
- Bhargava R, Gerald WL, Li AR, Pan Q, Lal P, Ladanyi M, Chen B. (2005). EGFR gene amplification in breast cancer: correlation with epidermal growth factor receptor mRNA and protein expression and HER-2 status and absence of EGFR-activating mutations. Modern Pathol 18:1027-1033.
- Carpenter G, Cohen S. (1979). Epidermal growth factor. Annu Rev Biochem 48:193-216.
- Ciardiello F, Tortora G. (2003). Epidermal growth factor receptor (EGFR) as a target in cancer therapy: understanding the role of receptor expression and other molecular determinants that could influence the response to anti-EGFR drugs. Eur J Cancer 39:1348-1354.
- Destexhe E, Lespagnard L, Degeyter M, Heymann R, Coignoul F. (1993). Immunohistochemical identification of myoepithelial, epithelial, and connective tissue cells in canine mammary tumors. Vet Pathol 30:146-154.
- Donnay I, Devleeschouwer N, Wouters-Ballman P, Leclercq G, Verstegen J. (1996). Relationship between receptors for epidermal growth factor and steroid hormones in normal, dysplastic and neoplastic canine mammary tissues. Res Vet Science 60:251-254.
- Donnay I, Rauis J, Wouters-Ballman P, Devleeschouwer N, Leclercq G, Verstegen JP. (1993). Receptors for oestrogen, progesterone and epidermal growth factor in normal and tumorous canine mammary tissues. J Repr Fertil Suppl 47:501-512.
- Elston CW, Ellis IO. (1998). Assessment of histological grade. In: Systemic Pathology The Breast, CW Elston and IO Ellis (Eds.), third ed., vol. 13, Churchill and Livingstone, London, pp. 365–384.
- Espinosa de Los Monteros A, Millán MY, Ordás J, Carrasco L, Reymundo C, Martín de Las Mulas J. (2002). Immunolocalization of the smooth muscle-specific protein calponin in complex and mixed tumors of the mammary gland of the dog: assessment of the morphogenetic role of the myoepithelium. Vet Pathol 39:247-256.
- Fox SB, Smith K, Hollyer J, Greenall M, Hastrich D, Harris AL. (1994). The epidermal growth factor receptor as a prognostic marker: results of 370 patients and review of 3009 patients. Breast Cancer Res Treat 29:41-49.

- Gama A, Alves A, Gartner F, Schmitt FC. (2003). P63: a novel myoepithelial cell marker in canine mammary tissues. Vet Pathol 40:412-420.
- Gama A, Paredes J, Albergaria A, Gartner F, Schmitt FC. (2004). P-cadherin expression in canine mammary tissues. J Comp Pathol 130:13-20.
- Hynes NE, Lane HA. (2005). ERBB receptors and cancer: the complexity of targeted inhibitors. Nat Rev Cancer 5:341-354.
- Kersting C, Tidow N, Schmidt H, Liedtke C, Neumann J, Boecker W, van Diest PJ, Brandt B, Buerger H. (2004). Gene dosage PCR and fluorescence in situ hybridization reveal low frequency of EGFR amplifications despite protein overexpression in invasive breast carcinoma. Lab Invest 84:582-587.
- Klijn JGM, Berns PMJJ, Schmitz PIM, Foekens JA. (1992). The clinical significance of epidermal growth factor receptor (EGFR) in human breast cancer: a review on 5232 patients. Endocr Rev 13:3-17.
- Lewis S, Locker A, Todd JH, Bell JA, Nicholson R, Elston CW, Blamey RW, Ellis IO. (1990). Expression of epidermal growth factor receptor in breast carcinoma. J Clin Pathol 43:385-89.
- Matsuyama S, Nakamura M, Yonezawa K, Shimada T, Ohashi F, Takamori Y, Kubo K. (2001). Expression patterns of the erbB subfamily mRNA in canine benign and malignant mammary tumours. J Vet Med Sci 63:949-954.
- Misdorp W, Else RW, Hellmén E, Lipscomb TP. (1999). Histological Classification of Mammary Tumors of the Dog and the Cat, 2nd series, Armed Forces Institute of Pathology, American Registry of Pathology, Washington D.C., and the World Health Organization Collaborating Center for Worldwide Reference on Comparative Oncology, pp. 11-56.
- Misdorp W. (2002). Tumours of the mammary gland. In: Tumors in Domestic Animals, DJ Meuten Ed., 4th Edition, Iowa State Press, Blackwell Publishing Company.
- Moller P, Mechtersheimer G, Kaufmann M, Moldenhauer G, Momburg F, Mattfeldt T, Otto HF. (1989). Expression of epidermal growth factor receptor in benign and malignant primary tumours of the breast. Virchows Arch A Pathol Anat Histopathol 414:157-164.
- Nerurkar VR, Chitale AR, Jalnakurpar BV, Naik SN, Lalitha VS. (1989). Comparative pathology of canine mammary tumours. J Comp Pathol 101:389-397.
- Nerurkar VR, Seshadri R, Mulherkar R, Ishwad CS, Lalitha VS, Naik SN. (1987). Receptors for epidermal growth factor and estradiol in canine mammary tumors. Int J Cancer 40:230-232.
- Nielsen TO, Hsu FD, Jensen K, Cheang M, Karaca G, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslen LA, Ragaz J, Gown AM, Gilks CB, van de Rijn M, Perou

- CM. (2004). Immunohistochemical and clinical characterization of the basal- like subtype of invasive breast carcinoma. Clin Cancer Res 10:5367-5374.
- Park K, Han S, Shin E, Kim HJ, Kim JY. (2007). EGFR gene and protein expression in breast cancers. Eur J Surg Oncol 33:956-960.
- Pirinen R, Lipponen P, Syrjänen K. (1995). Expression of epidermal growth factor receptor (EGFR) in breast cancer as related to clinical, prognostic and cytometric factors. Anticancer Res 15:2835-2840.
- Reis Filho JS, Milanezi F, Carvalho S, Simpson PT, Steele D, Savage K, Lambros MB, Pereira EM, Nesland JM, Lakhani SR, Schmitt FC. (2005). Metaplastic breast carcinomas exhibit EGFR, but not HER2, gene amplification and overexpression: immunohistochemical and chromogenic in situ hybridization analysis. Breast Cancer Res 7:RA1028-1035.
- Reis-Filho JS, Pinheiro C, Lambros MB, Milanezi F, Carvalho S, Savage K, Simpson PT, Jones C, Swift S, Mackay A, Reis RM, Hornick JL, Pereira EM, Baltazar F, Fletcher CD, Ashworth A, Lakhani SR, Schmitt FC. (2006). EGFR amplification and lack of activating mutations in metaplastic breast carcinomas. J Pathol 209:445-453.
- Reis-Filho JS, Milanezi F, Steele D, Savage K, Simpson PT, Nesland JM, Pereira EM, Lakhani SR, Schmitt FC. (2006). Metaplastic breast carcinomas are basal-like tumours. Histopathology 49:10-21.
- Rutteman GR, Foekens JA, Portengen H, Vos JH, Blankenstein MA, Teske E, Cornelisse CJ, Misdorp W. (1994). Expression of epidermal growth factor receptor (EGFR) in non-affected and tumorous mammary tissue of female dogs. Breast Cancer Res Treat 30:139-146.
- Sainsbury JR, Malcolm AI, Appleton DR, Farndom JR, Harris AL. (1985). Presence of epidermal growth factor receptor as an indicator of poor prognosis in patients with breast cancer. J Clin Pathol 38:1225-1228.
- Santini D, Ceccarelli C, Tardio ML, Taffurelli M, Marrano D. (2002). Immunocytochemical expression of epidermal Growth factor receptor in myoepithelial cells of the breast. Appl Immunohistochem Mol Morphol 10:29-33.
- Sassen A, Rochon J, Wild PJ, Hartmann A, Hofstaedter F, Schwarz S, Brockhoff G. (2008). Cytogenetic analysis of HER1/EGFR, HER2, HER3, and HER4 in 278 breast cancer patients. Breast Cancer Res 10:R2.
- Shien T, Tashiro T, Omatsu T, Masuda T, Furuta K, Sato N, Akashi-Tanaka S, Uehara M, Iwamoto E, Kinoshita T, Fukutomi T, Tsuda H, Hasegawa T. (2005). Frequent overexpression of epidermal growth factor receptor (EGFR) in mammary high grade ductal carcinomas with myoepithelial differentiation. J Clin Pathol 58:1299-1304.

- Toi M, Tominaga T, Osaki A, Toge T. (1994). Role of epidermal growth factor receptor expression in primary breast cancer: results of a biochemical study and an immunocytochemical study. Breast Cancer Res Treat 29:51-58.
- Tsutsui S, Ohno S, Murakami S, Hachitanda Y, Oda S. (2002). Prognostic value of epidermal growth factor receptor (EGFR) and its relationship to the estrogen receptor status in 1029 patients with breast cancer. Breast Cancer Res Treat 71:67-75.

Chapter V

Expression and prognostic significance of CK19 in canine malignant mammary tumours

Gama A, Alves A, Schmitt F (submitted)

Expression and prognostic significance of CK19 in canine malignant mammary tumours

Gama A^a, Alves A^a, Schmitt F^{b,c*}

^a Department of Veterinary Sciences, CECAV, University of Trás-os-Montes and Alto Douro

(UTAD), 5001-811 Vila Real, Portugal

^b Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP),

4200-465 Porto, Portugal

^c Medical Faculty, University of Porto, Porto, Portugal

* Corresponding author:

Tel.: 351-22-557 07 00; fax: 351-22-557 07 99.

E-mail address: fernando.schmitt@ipatimup.pt (F. Schmitt).

Abstract

The prognostic significance of the expression of cytokeratins (CK) in canine mammary

malignant tumours is still unclear. We have examined the expression pattern of the

luminal cytokeratin CK19 in a series of 102 canine mammary carcinomas by

immunohistochemical analysis, and associated its expression with known prognostic

markers, proliferation and survival. We also compared CK19 with the presence of

basal/myoepithelial cell markers. Reduced/absent CK19 was significantly associated

with histological type, invasiveness, high histological grade and an elevated Ki-67

index. CK19 positive expression was significantly associated with the presence of ER,

whereas its reduced immunostaining was associated with basal/myoepithelial cell

markers positive expression. Survival analysis demonstrated that down-regulation of

this luminal CK is significantly associated with shorter overall and disease-free survival

rates; however CK19 was not an independent prognostic factor in multivariate analysis.

Thus, CK19 down-regulation in canine mammary carcinomas is related to an aggressive

phenotype and seems to play a role in tumour progression.

Keywords: Canine; Mammary tumours; Cytokeratin 19; Prognosis

127

Introduction

Mammary gland tumours are the most commonly occurring neoplasm in the female dog and are known for their biological and histomorphological heterogeneity (Ferguson, 1985; Nerurkar *et al*, 1989; Moulton, 1990). Despite several immunohistochemical studies on the diagnostic value of specific luminal and basal/myoepithelial cell markers in canine mammary tumours (Hellmén and Lindgren, 1989; Griffey *et al.*, 1993; Vos *et al.*, 1993a, b; Espinosa de los Monteros, 2002; Gama *et al.*, 2003, 2004), the increasing prevalence of canine neoplasia compels the veterinary pathologist not only to determine a precise diagnosis but also to assess prognosis (Zaidan Dagli, 2008).

Cytokeratin (CK) is one of the three types of intermediate filaments that constitute the cytoskeleton of epithelial mammalian cells (Moll *et al.*, 1982; Chu and Weiss, 2002). CKs comprise a family of related proteins encoded by different genes, which are expressed in various epithelial cells in a developmentally regulated and differentiation-dependent manner (Romano *et al.*, 1988; Bocker *et al.*, 2002; Chu and Weiss, 2002). Simple epithelia express cytokeratins of low molecular weight, whereas stratified epithelia contain high molecular weight keratins.

In the normal human breast, the majority of luminal cells express CK7, CK8, CK18 and CK19, while basal/myoepithelial cells express CK5, CK14, CK15 and CK17 (Bocker *et al.*, 2002). Considering that all of these cells can undergo malignant change, breast carcinomas can be classified as expressing a luminal or a basal phenotype (Sorlie *et al.*, 2001; Abd El-Rehim *et al.*, 2004), with several studies describing a significant association between basal phenotype and poor prognosis (van de Rijn *et al.*, 2002; Abd El-Rehim *et al.*, 2004).

As for the canine species, we and others have described the expression of a number of cell differentiation markers, such as CK (Hellmén and Lindgren, 1989; Destexhe *et al.*, 1993; Griffey *et al.*, 1993; Vos *et al.*, 1993a, b and c; Rabanal and Else, 1994), p63 (Gama *et al.*, 2003; Ramalho *et al.*, 2004), P-cadherin (Gama *et al.*, 2004, 2007); SMA (Destexhe *et al.*, 1993; Vos *et al.*, 1993a, b and c) and calponin (Espinosa de los Monteros *et al.*, 2002) in mammary gland tissues. CK 8, 18 and 19 have been considered luminal cell markers (Griffey *et al.*, 1993; Vos *et al.*, 1993a, b and c; Rabanal and Else, 1994), whereas CK5, 14 and 17 (Griffey *et al.*, 1993; Vos *et al.*, 1993a, b and c), p63 and P-cadherin (Gama *et al.*, 2003, 2004, 2007) were found to be expressed by basal/myoepithelial cells. SMA and calponin were exclusively expressed

by myoepithelial cells (Destexhe *et al.*, 1993; Vos *et al.*, 1993a, b and c; Espinosa de los Monteros *et al.*, 2002). Based on the immunohistochemical expression of CK14, Griffey *et al.* (1993) applied the "basal carcinoma" nomenclature to canine carcinomas. Similarly to humans, these carcinomas showed aggressive clinical behaviour (Griffey *et al.*, 1993) but, to the best of our knowledge, no investigations have been performed associating CK expression and prognosis in canine mammary tumours.

Recently, down-regulation of luminal CKs 18 and 19 has been identified as a significant predictor of aggressive disease in breast cancer patients (Woelfle *et al.*, 2004; Parikh *et al.*, 2008). We sought to evaluate the immunohistochemical expression of the CK19 luminal cell marker in a series of malignant canine mammary tumours and its possible correlation with some relevant clinicopathological parameters, namely proliferation, oestrogen receptor status, other cell-specific cell markers (basal/myoepithelial) and survival, in order to investigate its possible value as a prognostic marker in canine mammary cancer.

Material and methods

Tumour specimens

The present study is based on a series of a 102 cases of canine malignant mammary tumours selected from the histopathological files of the University of Trás-os-Montes and Alto Douro, Vila Real, and from the Institute of Biomedical Science at the University of Porto, Portugal. The material was fixed in 10% neutral buffered formalin and embedded in paraffin wax. Sections (3 µm) were cut and stained with haematoxylin and eosin (HE) for histological examination or immunohistochemistry.

Case follow-up

Sixty nine dogs (n = 69) were followed post-surgically by the referring surgeons and presented a median overall survival time of 15 months (range 5-74 months). Overall survival (OS) was defined as the period between surgery and animal natural death or euthanasia due to cancer. Disease-free survival (DFS) was defined as the period of time between surgery and recurrent or metastatic disease. During the follow-up period, according to the referring surgeons, 35 animals died or were euthanized due to

metastatic disease and/or local recurrence. One dog died due to causes unrelated to the mammary tumour and it was censored at the time of death (19 months).

Histological Examination

Tumours were diagnosed according to the World Health Organization (WHO) criteria for canine mammary neoplasms (Misdorp *et al.*, 1999). Clinicopathological variables included in the present study were: age, breed, ovariohysterectomy status, contraceptive administration, tumour size, tumour location, presence of skin ulceration, tumour histological type, presence of intra-tumoral necrosis, mode of growth (expansive vs. infiltrative), presence of stromal and vascular invasion and presence of lymph node metastasis.

Tumours were evaluated for grade in accordance with the Nottingham method for human breast tumours (Elston and Ellis, 1998), based on the assessment of three morphological features: tubule formation, nuclear pleomorphism and mitotic counts. Each of these features was scored on a scale of 1-3 to indicate whether it was present in slight, moderate or marked degree, giving a putative total of 3-9 points. Grade was allocated by an arbitrary division of the total points as follows: grade I (well differentiated): 3-5 points; grade II (moderately differentiated): 6-7 points; and grade III (poorly differentiated): 8-9 points.

Immunohistochemistry

Tissue sections were incubated with primary monoclonal antibodies against CK19, Ki-67, oestrogen receptor (ER), CK5, CK14, calponin, p63, P-cadherin and α-smooth muscle actin (SMA). Table 1 summarises the antibodies used and the staining procedures adopted for each antibody. Antigen retrieval was carried out by microwave treatment in 10 mM citrate buffer, pH 6.0, with the exception of P-cadherin, which was treated with an ethylene diamine tetraacetic acid (EDTA) buffer, pH 8.0 (LabVision) in a boiling bath for 20 min. For Ki-67 antigen retrieval, slides were incubated with 0.2 mg/mL trypsin (Merck) in phosphate buffered saline (PBS) for 10 min at 37 °C. After cooling (20 min at room temperature), the sections were immersed in 3% hydrogen peroxide and distilled water for 30 min to block endogenous peroxidase activity. All slides were incubated with a blocking serum (LabVision) for 10 min and then incubated with the specific antibody.

After incubation, slides were incubated with biotinylated secondary antibody, followed by streptavidin-conjugated peroxidase (LabVision), except for ER. For this antibody, a polymeric labelling methodology was used as a detection system (Novolink Polymer Detection System, Novocastra), following the manufacturer's instructions. Subsequently, the colour was developed with 3,3-diaminobenzidine tetrahydrochloride (DAB) and slides were counterstained with Gill's haematoxylin, dehydrated, and mounted for evaluation by light microscopy.

Adjacent normal mammary tissues were used as internal positive controls. Negative controls were carried out by replacing the primary antibody with PBS.

Table 1. Primary monoc	lonal antibodies and	immunostaining prote	ocols used
3		\mathcal{L}_{1}	

Antibody	Origin	Clone	Dilution	Pretreatment	Incubation
CK19	Neomarkers, USA	BA17	1:150	Microwave	Overnight
Ki-67	Dako, Denmark	Mib1	1:50	Trypsin + Microwave	Overnight
ER	Novocastra, UK	NCL-LH2	1:40	Microwave	2 hours
CK5	Neomarkers, USA	XM26	1:25	Microwave	Overnight
CK14	Novocastra, UK	NCL-LL002	1:20	Microwave	2 hours
CALP	Dako, Denmark	CALP	1:400	Microwave	2 hours
P63	Neomarkers, USA	4A4	1:150	Microwave	Overnight
PCAD	BD Transduction, USA	56	1:50	Water bath, 98°C	Overnight
SMA	Novocastra, UK	NCL-SMA	1:50	Microwave	2 hours

Evaluation of the immunohistochemical data

CK19 positivity was indicated by the presence of cytoplasmic staining in neoplastic cells. For the evaluation of CK19 expression, we adopted a scoring method based on the estimation of the staining intensity in combination with an estimation of the percentage of immunoreactive cells (Parikh *et al.*, 2008). As for staining intensity, CK19 expression was scored as: 0: no staining; 1: weak to moderate intensity; and 2: strong intensity. As for percentage, tumours were evaluated in <10% positive cells and \geq 10% positive cells. For statistical purposes, CK19 status was considered positive if a tumour presented \geq 10% positive epithelial cells, with a moderate or strong immunoreactivity. Ki-67 immunostaining was evaluated as described previously (Gama *et al.*, 2008). Immunoreactivities were classified by estimating the percentage of tumour cells showing characteristic staining. Nuclear ER immunoreactivity was considered positive when more than 10% of the neoplastic cells expressed this marker. A semi-quantitative analysis was performed for calponin, CK5, CK14, SMA and p63: 0: <10% positive

cells; 1: 10-50% positive cells; and 2: >50% positive cells, with a cytoplasmic (calponin, CK5, CK14, SMA) or nuclear (p63) pattern of cellular distribution. P-cadherin immunostaining was evaluated semi-quantitatively as previously described (Gama *et al.*, 2004, 2008): 0: <10% positive cells; 1: 10-25% positive cells; 2: 26-50% positive cells and 3: >50% positive cells, with cells showing a membranous and/or cytoplasmic expression pattern.

Statistical Analysis

Associations between expression of CK19 and continuous variables (mitotic and Ki-67 indices) were assessed by the non parametric Mann-Whitney test. Associations between CK19 expression and categorical variables, such as tumour size, histological type, histological grade and invasion, were performed using the χ^2 test. Fisher's exact test was performed when compared variables had exactly two groups (2 x 2 table). Survival curves were generated by the Kaplan-Meier method and the survival rates were compared using the log-rank test. The combined effects of CK19 with previously recognised prognostically relevant variables were examined via Cox proportional hazards model. All statistical analysis was performed using SPSS 11.5 statistical software. A P value <0.05 was considered to be statistically significant.

Results

Patients and tumour characteristics

The mean age of dogs at the time of surgical removal of tumours was 9.6 ± 2.4 years (range 4-16 years). Most dogs in our series were mixed breeds (n = 34, 34.7%), followed by Toy poodles (n = 18, 18.4%) and Cocker spaniels (n = 11, 11.2%). The mean maximum tumour diameter was 4.24 ± 3.4 cm (range 0.5-18 cm), with tumours more frequently located in caudal mammary glands (n = 39; 57.4%). Skin ulceration was present in 19 cases (19%). In 11/74 (14.9%) female dogs with available clinical information, ovariohysterectomy was performed prior to the removal of mammary tumours. There was a history of contraceptive administration in 8/63 (12.7%) cases. Histologically, tumour types comprised 45/102 (44.1%) simple carcinomas, 45/102

(44.1%) complex carcinomas and 12/102 (11.8%) carcinosarcomas. According to the Nottingham method, tumours were classified as grade I (n = 17, 16.7%), grade II (n = 34, 33.3%) and grade III (n = 51, 50%).

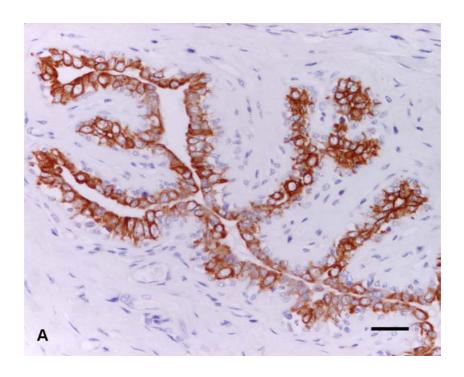
CK19 expression in canine mammary malignant tumours

CK19 was consistently expressed in the normal adjacent canine mammary gland, being exclusively observed in luminal epithelial cells (Fig. 1A). In contrast, CK19 expression was variable in malignant tumour tissues. Cytoplasmic expression was evident in more than 10% of epithelial cells in 78/102 (76.5%) malignant tumours, whereas 24/102 (23.5%) presented with no staining or staining in less than 10% neoplastic cells (Fig. 1B and C).

The relationships between CK19 status and several clinicopathological variables are shown in Tables 2 and 3. The proportion of CK19 positive and negative tumours differed significantly with gland location (P = 0.04), histological type (P = 0.002), tumour growth pattern (P = 0.001), histological grade (P < 0.0001) and stromal and lymphovascular invasion (P = 0.003 and P < 0.0001, respectively), with most CK19 negative tumours exhibiting aggressive phenotypical features, such as high histological grade and stromal/vascular invasion.

The associations found between CK19 expression and the presence of other molecular cell markers are summarised in Table 4. With regard to these cell markers, CK19 expression showed significant differences across the different staining groups for ER, CK5, P-cadherin, p63 and calponin. Eighteen out of 22 (81.8%) CK19 negative tumours were ER negative (P<0.0001), 15/24 (62.5%) CK19 negative tumours were CK5 positive (P = 0.015), 19/23 (82.6%) CK19 negative tumours were P-cadherin positive (P = 0.039), 21/24 (87.5%) CK19 negative tumours were p63 positive and 13/24 (54.2%) CK19 negative tumours were calponin positive. No significant differences were observed between CK19 expression and CK14 or SMA staining patterns.

As shown in Table 4, compared with positive tumours, the CK19 negative group of tumours showed higher mitotic and Ki-67 labelling indices, being significantly associated with Ki-67 proliferative index (P = 0.017).



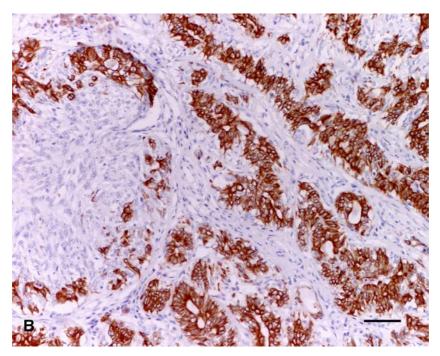


Fig. 1. Immunohistochemical expression of CK19 in canine mammary tissues. A. Normal mammary duct, showing CK19 expression restricted to luminal epithelial cells. Bar = 30 μ m. B. Complex carcinoma (grade II) with strong cytoplasmic CK19 expression in more than 10% of epithelial cells. Bar = 60 μ m.

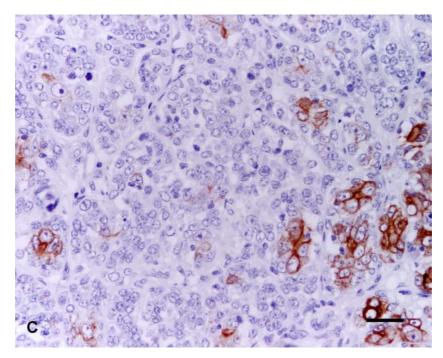


Fig. 1. (cont.) Immunohistochemical expression of CK19 in canine mammary tissues. C. Solid carcinoma (grade III) negative for CK19 immunostaining, with less than 10% positive neoplastic cells. Bar = $30 \mu m$.

Prognostic significance of CK19 expression

Follow up data is summarised in Table 5. Kaplan-Meier survival curves revealed that a down-regulated expression of CK19 was significantly associated with lower overall (P = 0.0001, Fig. 2A) and disease-free (P = 0.0004, Fig. 2B) survival. To investigate whether CK19 expression represents an independent prognostic factor for canine mammary cancer, a multivariate analysis, including the histopathological parameters lymph node status and grade of differentiation, was performed; however the data showed that CK19 expression was not an independent prognostic parameter.

Table 2. Association between CK19 expression and clinical parameters

Clinical parameters	n	CK19	CK19 expression		
Clinical parameters		Negative	Positive		
Age	97ª				
\leq 9 years old	47	10 (21.3%)	37 (78.7%)		
> 9 years old	50	12 (24%)	38 (76%)		
P		0.81			
Breed	98ª				
Mixed breed	34	9 (26.5%)	25 (73.5%)		
Poodle	18	5 (27.8%)	13 (72.2%)		
Cocker spaniel	11	2 (18.2%)	9 (81.8%)		
Others	35	7 (20%)	28 (80%)		
P		0.87			
Tumour size	95ª				
<3 cm	36	6 (16.7%)	30 (83.3%)		
3-5 cm	33	10 (30.3%)	23 (69.7%)		
>5 cm	26	7 (26.9%)	19 (73.1%)		
P		0.42	, ,		
Tumour location	68 ^a				
Cranial glands (1 and 2)	5	2 (40%)	3 (60%)		
Medial gland (3)	10	1 (10%)	9 (90%)		
Caudal glands (4 and 5)	39	13 (33.3%)	26 (66.7%)		
Multiple	14	0 (0%)	14 (100%)		
P		0.04			
Skin ulceration	100 ^a				
Absent	81	18 (22.2%)	63 (77.8%)		
Present	19	4 (21.1%)	15 (78.9%)		
P		0.99			
Ovariohysterectomy	74ª				
No	51	14 (27.5%)	37 (72.5%)		
Yes, prior to tumour development	11	0 (0%)	11 (100%)		
Yes, performed with mastectomy	12	4 (33.3%)	8 (66.7%)		
P		0.13	<u> </u>		
Contraception	63ª				
No	55	13 (23.6%)	42 (76.4%)		
Yes	8	2 (25%)	6 (75%)		
P		0.99			

^a Total number of cases for which clinical information was available

Table 3. Association between CK19 expression and pathological parameters

Pathological parameters	n	CK19		
ratifological parameters	n	Negative	Positive	
Histological type				
Simple carcinoma	45	17 (37.8%)	28 (62.2%)	
Complex carcinoma	45	3 (6.7%)	42 (93.3%)	
Carcinosarcoma	12	4 (33.3%)	8 (66.7%)	
P		0.002		
Necrosis				
Absent	5	0 (0%)	5 (100%)	
Present	97	24 (24.7%)	73 (75.3%)	
P		0.58		
Mode of growth				
Expansive	31	1 (3.2%)	30 (96.8%	
Infiltrative	71	23 (32.4%)	48 (67.6%	
P		0.001		
Histological grade				
Grade I	17	0 (0%)	17 (100%)	
Grade II	34	1 (2.9%)	33 (97.1%	
Grade III	51	23 (45.1%)	28 (54.9%	
P		<0.0001		
Stromal invasion				
Absent	27	1 (3.7%)	26 (96.3%)	
Present	75	23 (30.7%)	52 (69.3%	
P		0.003		
Lymphovascular invasion				
Absent	43	2 (4.7%)	41 (95.3%	
Present	59	22 (37.3%)	37 (62.7%	
P		< 0.0001		
Lymph node metastasis ^a				
Absent	24	4 (16.7%)	20 (83.3%	
Present	30	7 (23.3%)	23 (76.7%)	
P		0.73		

^a Lymph nodes were available in 54 cases

Table 4. Association between CK19 expression and other molecular markers and proliferation indices

Molecular markers ^{a, b}	n		CK19
		Negative	Positive
ER			
Negative	40	18 (45%)	22 (55%)
Positive	56	4 (7.1%)	52 (92.9%)
P		<0.0001	
CK5			
0	20	9 (45%)	11 (55%)
1	49	6 (12.2%)	43 (87.8%)
2	33	9 (27.3%)	24 (72.7%)
P		0.015	
CK14			
0	17	5 (29.4%)	12 (70.6%)
1	56	12 (21.4%)	44 (78.6%)
2	29	7 (24.1%)	22 (75.9%)
P		0.77	
P-cadherin			
0	9	1 (11.1%)	8 (88.9%)
1	23	2 (8.7%)	21 (93.3%)
2	22	4 (18.2%)	18 (81.8%)
3	42	16 (38.1%)	26 (61.9%)
D		0.039	
P63			
0	29	3 (10.3%)	26 (89.7%)
1	40	4 (10%)	36 (90%)
2	33	17 (51.5%)	16 (48.5%)
0		<0.0001	
Calponin			
0	37	11 (29.7%)	26 (70.3%)
1	47	4 (8.5%)	43 (91.5%)
2	18	9 (50%)	9 (50%)
D		0.001	
SMA			
0	54	13 (24.1%)	41 (75.9%)
1	35	5 (14.3%)	30 (85.7%)
2	11	5 (45.5%)	6 (54.5%)
D		0.09	
Median Mitotic index		0.98	0.59
(Min-Max)		(0.1-1.9)	(0-2.99)
P		0.05	
Median Ki-67 index		27.97	21.06
(Min-Max)		(12.10-39.40)	(5.39-56.36)
D		0.017	

^a Immunohistochemical evaluation of ER and P-cadherin was available in 96 cases; SMA was available in 100 cases and Ki-67 was available in 95 cases; ^b Score for CK5, CK14, SMA and p63: 0: <10% positive cells; 1: 10-50% positive cells; and 2: >50% positive cells; score for P-cadherin: 0: <10% positive cells; 1: 10-25% positive cells; 2: 26-50% positive cells and 3: >50% positive cells.

		Overall survival			Disease-	free survival
		Mean survival	Average 1 year		Mean survival	Average 1 year
CK19 status	n^a	(months)	survival rate n (%)	n	(months)	survival rate n (%)
Negative	14	5	1 (9.5)	14	4	0 (0.0)
Positive	55	43	30 (62.9)	54	13	19 (41.6)
P		0.0001			0.0004	

Table 5. Survival rates in dogs with available follow up

^a Follow up data was available in 69 cases for OS and 68 cases for DFS.

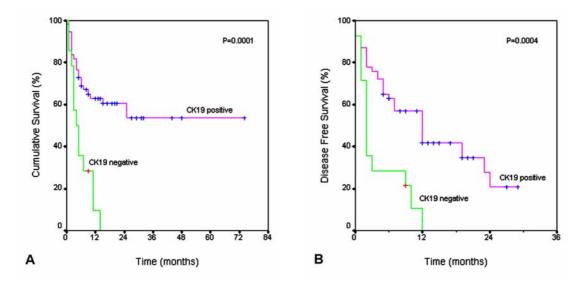


Fig. 2. Kaplan-Meier overall survival (A) and disease-free survival (B) curves of groups with positive and negative CK19 expression (P=0.0001 and P=0.0004, respectively).

Discussion

About half of canine mammary tumours are considered to be malignant and it is of great importance to recognise and identify reliable prognostic factors to estimate the individual risk of an unfavourable clinical outcome (Misdorp, 2002; Zaidan Dagli, 2008). Down-regulation of CK19 has been identified as an independent prognostic factor in human breast cancer (Parikh *et al.*, 2008) and the present study demonstrates that CK19 might have a putative role in canine mammary tumour progression although it is not an independent prognostic parameter.

CKs have been recognised as epithelial cell markers in diagnostic histopathology for over 20 years (Moll *et al.*, 1982; Coulombe and Omary, 2002). In the human mammary gland, immunohistochemistry has demonstrated that luminal cells usually express CK8,

18, and 19, while basal/myoepithelial cells express CK5, CK14, CK15 and CK17 (Moll *et al.*, 1982; Bocker *et al.*, 2002). CK19 expression also has been described in canine mammary tissues as a luminal cell marker (Destexhe *et al.*, 1993; Vos *et al.*, 1993c; Sarli *et al.*, 2007). To the best of our knowledge, this is the first study correlating its expression pattern with clinicopathological parameters and clinical outcome for canine mammary carcinomas.

Immunohistochemical studies have shown that human breast carcinomas usually retain the CK pattern of normal luminal epithelial cells (Moll *et al.*, 1982). Monoclonal antibodies directed against luminal CK18 and 19 have been used to identify primary and metastatic human breast cancer cells (Malzahn *et al.*, 1998). A subset of tumours has been identified with down-regulation of luminal CKs associated with aggressive biological behaviour and poor outcome (Woelfle *et al.*, 2004; Parikh *et al.*, 2008), challenging the view that CKs are purely marker proteins.

Moreover, a number of regulatory changes in CK expression at the transcriptional and post-transcriptional level have been described in experimental cell studies (Blouin *et al.*, 1992; Choi *et al.*, 2000). Regulatory mechanisms include interaction with keratin associated proteins (KAPs), which results in CK phosphorylation, glycosylation, transglycosylation, caspase cleavage and ubiquitination (Coulombe and Omary, 2002). CKs may also associate with other cytoskeletal elements (Coulombe and Omary, 2002). Additional evidence for a more widespread role of CKs has come from mouse gene knockout studies, in which the double deletion of CK18 and 19 results in the lack of a functional CK skeleton and causes embryonic lethality (Hesse *et al.*, 2000).

Our immunohistochemical results demonstrated a variable CK19 expression among canine mammary malignant tumours, with down-regulation of CK19 in 23.5% of tumours. Similar observations have already been described in the literature on canine mammary tumours (Destexhe *et al.*, 1993; Vos at al., 1993b). With regard to clinicopathological parameters, most CK19 negative tumours were associated with aggressive phenotypical features, such as high histological grade and stromal/vascular invasion and high Ki-67 (proliferation) index. These results are in accordance with several reports in the human literature for luminal CKs markers, which found that loss of luminal CKs was significantly associated with a higher tumour grade and a higher mitotic index (Schaller *et al.*, 1996; Abd El-Rehim *et al.*, 2004; Woelfle *et al.*, 2004; Willipinski-Stapelfeldt *et al.*, 2005; Parikh *et al.*, 2008).

Recent studies using gene array technology on human breast tumours have identified distinct subtypes of breast carcinomas (luminal A, luminal B, Her2 over-expressing and basal-like) that are associated with different clinical outcomes (Perou *et al.*, 2000; Sorlie *et al.*, 2001; Sorlie *et al.*, 2003). Luminal A and B subtypes are based on the expression of ER, usually with CK19 expression (Birnbaum *et al.*, 2004) whereas the basal-like subtype is characterised by the absence of hormonal receptors and expression of basal cell markers, such as CK5, p63 or P-cadherin. In the present study, we have described a subset of carcinomas with down-regulation of CK19 expression, which lack ER, and associated with basal and/or myoepithelial cell differentiation. In fact, tumours negative for both CK19 and ER were always associated with the expression of at least one basal or myoepithelial cell marker. So, this subset might correspond to the so-called basal-like subtype, which has been found to be associated with a particularly poor clinical outcome in human patients (Sorlie *et al.*, 2001; Abd El-Rehim *et al.*, 2004; Nielsen *et al.*, 2004; Rakha *et al.*, 2006); however, additional studies are needed in order to confirm this hypothesis.

The loss of expression of luminal cytokeratins in conjunction with lack of ER expression, similar to that found in this study, has been described previously in human breast carcinomas (Abd El-Rehim *et al.*, 2004; Willipinski-Stapelfeldt *et al.*, 2005; Parikh *et al.*, 2008). Choi *et al.* (2000) demonstrated that CK19 is under rapid and direct regulation by oestrogen in the MCF-7 breast cancer cell line, which suggest that cytoskeletal organisation might also be driven by hormonally-mediated stimuli in human breast cancer (Santini *et al.*, 1996; Choi *et al.*, 2000).

In the present study, we also performed a clinical follow-up analysis to assess CK19 prognostic significance and our results suggest that CK19 is associated with a more aggressive phenotype, since its down-regulation was significantly related with shorter OS and DFS. Previous studies on human breast cancer have made similar observations for luminal CK18 and 19 (Schaller *et al.*, 1996; Woelfle *et al.*, 2004; Parikh *et al.*, 2008). However, a multivariate analysis including well-known prognostic variables showed that CK19 expression was not an independent prognostic factor in our series of canine mammary tumours, in contrast to human breast cancer (Parikh *et al.*, 2008).

Considering that changes in the composition of the cytoskeleton of tumour cells may result in increased plasticity, which is required for epithelial tumour cells to become mobile and invasive (Thiery, 2002), luminal CK expression down-regulation might

among other factors lead to less differentiated tumour cells. Gene expression profile studies on breast cancer cell lines have found that expression of CK19 is consistently elevated in the less aggressive cell lines, whereas the highly aggressive cell lines expressed vimentin, a mesenchymal cell marker (Zajchowski *et al.*, 2001). Similarly, Willipinski-Stapelfeldt *et al.* (2005) found that micrometastatic breast cancer cell lines displayed loss of luminal cytokeratins (CK8, CK18, and CK19) and showed an ectopic expression of vimentin, which is indicative of epithelial-mesenchymal transition.

Conclusions

Our findings demonstrate that CK19 down-regulation is associated with an aggressive tumour phenotype, identifying a group of dogs with higher risk of tumour progression. However, additional studies are warranted to confirm these findings and to investigate whether CK19 plays an active role, or whether the observed changes at the expression level merely reflect more upstream processes.

Acknowledgements

The authors thank Prof. Fátima Gärtner (Institute of Biomedical Science at the University of Porto, Portugal) for the contribution of some cases included in this study. We also thank Mrs Lígia Bento for expert technical assistance. This work was supported by the Centro de Ciência Animal e Veterinária (CECAV), University of Trás os Montes e Alto Douro (UTAD), Vila Real, Portugal, and by Portuguese Science and Technology Foundation, project POCTI/CVT/57795/2004.

References

- Abd El-Rehim DM, Pinder SE, Paish CE, Bell J, Blamey RW, Robertson JF, Nicholson RI, Ellis IO. (2004). Expression of luminal and basal cytokeratins in human breast carcinoma. J Pathol 203:661-671.
- Birnbaum D, Bertucci F, Ginestier C, Tagett R, Jacquemier J, Charafe-Jauffret E. (2004). Basal and luminal breast cancers: basic or luminous? Int J Oncol 25:249-258.
- Blouin R, Swierenga SH, Marceau N. (1992). Evidence for post-transcriptional regulation of cytokeratin gene expression in a rat liver epithelial cell line. Biochem Cell Biol 70:1-9.
- Bocker W, Moll R, Poremba C, Holland R, Van Diest PJ, Dervan P, Burger H, Wai D, Ina Diallo R, Brandt B, Herbst H, Schmidt A, Lerch MM, Buchwallow IB. (2002). Common adult stem cells in the human breast give rise to glandular and myoepithelial cell lineages: a new cell biological concept. Lab Invest 82:737-746.
- Choi I, Gudas LJ, Katzenellenbogen BS. (2000). Regulation of keratin 19 gene expression by estrogen in human breast cancer cells and identification of the estrogen responsive gene region Mol Cell Endocrinol 164:225–237.
- Chu PG, Weiss LM. (2002). Keratin expression in human tissues and neoplasms. Histopathology 40:403-439.
- Coulombe PA, Omary MB. (2002). "Hard" and "soft" principles defining the structure, function and regulation of keratin intermediate filaments. Curr Opin Cell Biol 14:110-122.
- Destexhe E, Lespagnard L, Degeyter M, Heymann R, Coignoul F. (1993). Immunohistochemical identification of myoepithelial, epithelial, and connective tissue cells in canine mammary tumors. Vet Pathol 30:146-154.
- Elston CW, Ellis IO. (1998). Assessment of histological grade. In: Systemic Pathology. The Breast. CW Elston and IO Ellis (Eds), 3rd Ed., Churchill Livingstone, London, UK, pp. 365-384.
- Espinosa de Los Monteros A, Millán MY, Ordás J, Carrasco L, Reymundo C, Martín de Las Mulas J. (2002). Immunolocalization of the smooth muscle-specific protein calponin in complex and mixed tumors of the mammary gland of the dog: assessment of the morphogenetic role of the myoepithelium. Vet Pathol 39:247-256.
- Ferguson HR. (1985). Canine mammary gland tumors. Vet Clin North Am Small Anim Pract 15:501-511.
- Gama A, Alves A, Gartner F, Schmitt FC. (2003). P63: a novel myoepithelial cell marker in canine mammary tissues. Vet Pathol 40:412-420.

- Gama A, Paredes J, Albergaria A, Gartner F, Schmitt FC. (2004). P-cadherin expression in canine mammary tissues. J Comp Pathol 130:13-20.
- Gama A, Paredes J, Gärtner F, Alves A, Schmitt FC. (2008). Expression of E-cadherin, P-cadherin and β-catenin in canine malignant mammary tumours in relation to clinicopathological parameters, proliferation and survival. Vet J 177:45-53.
- Griffey SM, Madewell BR, Dairkee SH, Hunt JE, Naydan DK, Higgins RJ. (1993). Immunohistochemical reactivity of basal and luminal epithelium-specific cytokeratin antibodies within normal and neoplastic canine mammary glands. Vet Pathol 30:155-161.
- Hellmén E, Lindgren A. (1989). The expression of intermediate filaments in canine mammary glands and their tumors. Vet Pathol 26:420-428.
- Hesse M, Franz T, Tamai Y, Taketo MM, Magin TM. (2000). Targeted deletion of keratins 18 and 19 leads to trophoblast fragility and early embryonic lethality. EMBO J 19:5060-5070.
- Malzahn K, Mitze M, Thoenes M, Moll R. (1998). Biological and prognostic significance of stratified epithelial cytokeratins in infiltrating ductal breast carcinomas. Virchows Arch 433:119-129.
- Misdorp W, Else RW, Hellmén E, Lipscomb TP. (1999). Histological Classification of Mammary Tumors of the Dog and the Cat, 2nd series, Vol VII, Armed Forces Institute of Pathology, American Registry of Pathology, Washington D.C., and the World Health Organization Collaborating Center for Worldwide Reference on Comparative Oncology, pp. 1-59.
- Misdorp W. (2002). Tumours of the mammary gland. In: Tumors in Domestic Animals, DJ Meuten (Ed.), 4th Edition, Iowa State Press, Blackwell Publishing Company, pp. 575-606.
- Moll R, Franke WW, Schiller DL, Geiger B, Krepler R. (1982). The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. Cell 31:11-24.
- Moulton JE (1990). Tumors of the mammary gland. In: Tumors in domestic animals, JE Moulton (Ed.), 3rd Edition, University of California Press, Berkeley, pp. 518-553.
- Nerurkar VR, Chitale AR, Jalnakurpar BV, Naik SN, Lalitha VS. (1989). Comparative pathology of canine mammary tumours. J Comp Pathol 101:389-397.
- Nielsen TO, Hsu FD, Jensen K, Cheang M, Karaca G, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslen LA, Ragaz J, Gown AM, Gilks CB, van de Rijn M, Perou CM. (2004). Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. Clin Cancer Res 10:5367-5374.

- Parikh RR, Yang Q, Higgins SA, Haffty BG. (2008). Outcomes in young women with breast cancer of triple-negative phenotype: the prognostic significance of CK19 expression. Int J Radiat Oncol Biol Phys 70:35-42.
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lonning PE, Borresen-Dale AL, Brown PO, Botstein D. (2000). Molecular portraits of human breast tumours. Nature 406:747-752.
- Rabanal RM, Else RW. (1994). Immunohistochemical localisation of cytokeratin and vimentin intermediate filament proteins in canine mammary tumours. Res Vet Sci 56:225-233.
- Rakha EA, Putti TC, Abd El-Rehim DM, Paish C, Green AR, Powe DG, Lee AH, Robertson JF, Ellis IO. (2006). Morphological and immunophenotypic analysis of breast carcinomas with basal and myoepithelial differentiation. J Pathol 208:495-506.
- Ramalho LNZ, Ribeiro-Silva A, Cassali GD, Zucoloto S. (2006). The expression of p63 and cytokeratin 5 in mixed tumors of the canine mammary gland provides new insights into the histogenesis of these neoplasms. Vet Pathol 43:424-429.
- Romano V, Bosco P, Rocchi M, Costa G, Leube RE, Franke WW, Romeo G. (1988). Chromosomal assignments of human type I and type II cytokeratin genes to different chromosomes. Cytogenet Cell Genet 48:148-151.
- Santini D, Ceccarelli C, Taffurelli M, Pileri S, Marrano D. (1996). Differentiation pathways in primary invasive breast carcinoma as suggested by intermediate filament and biopathological marker expression. J Pathol 179:386-391.
- Sarli G, Sassi F, Brunetti B, Benazzi C. (2007). Luminal-like A and B types in canine mammary carcinomas. 25th Annual Meeting European Society for Veterinary Pathology, Munich, Germany, pp. 187.
- Schaller G, Fuchs I, Pritze W, Ebert A, Herbst H, Pantel K, Weitzel H, Lengyel E. (1996). Elevated keratin 18 protein expression indicates a favorable prognosis in patients with breast cancer. Clin Cancer Res 2:1879-1885.
- Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, Rijn MV, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Lonning PE, Borresen-Dale AL. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98:10869-10874.
- Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lonning PE, Brown PO, Borresen-Dale AL, Botstein D. (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 100:8418-8423.

- Thiery JP. (2002). Epithelial-mesenchymal transitions in tumour progression. Nat Rev Cancer 2:442-454.
- van de Rijn M, Perou CM, Tibshirani R, Haas P, Kallioniemi O, Kononen J, Torhorst J, Sauter G, Zuber M, Kochli OR, Mross F, Dieterich H, Seitz R, Ross D, Botstein D, Brown P. (2002). Expression of cytokeratins 17 and 5 identifies a group of breast carcinomas with poor clinical outcome. Am J Pathol 161:1991-1996.
- Vos JH, Van den Ingh TS, Misdorp W, Molenbeek RF, Van Mil FN, Rutterman GR, Ivanyi D, Ramaekers FCS. (1993a). Immunohistochemistry with keratin, vimentin, desmin, and α-smooth muscle actin monoclonal antibodies in canine mammary gland: benign mammary tumors and duct ectasias. Vet Q 14:89-95.
- Vos JH, Van den Ingh TS, Misdorp W, Molenbeek RF, Van Mil FN, Rutterman GR, Ivanyi D, Ramaekers FCS. (1993b). Immunohistochemistry with keratin, vimentin, desmin, and α-smooth muscle actin monoclonal antibodies in canine mammary gland: malignant mammary tumors. Vet Q 14:96-102.
- Vos JH, Van den Ingh TS, Misdorp W, Molenbeek RF, Van Mil FN, Rutterman GR, Ivanyi D, Ramaekers FCS. (1993c). Immunohistochemistry with keratin, vimentin, desmin, and α-smooth muscle actin monoclonal antibodies in canine mammary gland: normal mammary tissue. Vet Q 14:102-107.
- Willipinski-Stapelfeldt B, Riethdorf S, Assmann V, Woelfle U, Rau T, Sauter G, Heukeshoven J, Pantel K. (2005). Changes in cytoskeletal protein composition indicative of an epithelial-mesenchymal transition in human micrometastatic and primary breast carcinoma cells. Clin Cancer Res 11:8006-8014.
- Woelfle U, Sauter G, Santjer S, Brakenhoff R, Pantel K. (2004). Downregulated expression of cytokeratin 18 promotes progression of human breast cancer. Clin Cancer Res 10:2670-2674.
- Zaidan Dagli ML. (2008). The search for suitable prognostic markers for canine mammary tumors: A promising outlook. Vet J 177:3-5.
- Zajchowski DA, Bartholdi MF, Gong Y, Webster L, Liu HL, Munishkin A, Beauheim C, Harvey S, Ethier SP, Johnson PH. (2001). Identification of gene expression profiles that predict the aggressive behavior of breast cancer cells. Cancer Res 61:5168-5178.

Chapter VI

Identification of molecular phenotypes in canine mammary carcinomas with clinical implications: application of the human classification

Gama A, Alves A, Schmitt FC

Virchows Archiv (in press)

Identification of molecular phenotypes in canine mammary carcinomas with clinical implications: application of the human classification

Gama A^a, Alves A^a, Schmitt F^{b,c*}

^a Department of Veterinary Sciences, CECAV, University of Trás-os-Montes and Alto Douro

(UTAD), 5001-811 Vila Real, Portugal

^b Institute of Molecular Pathology and Immunology of the University of Porto (IPATIMUP),

4200-465 Porto, Portugal

^c Medical Faculty, University of Porto, Porto, Portugal

* Corresponding author:

Tel.: 351-22-557 07 00; fax: 351-22-557 07 99.

E-mail address: fernando.schmitt@ipatimup.pt (F. Schmitt).

Abstract

Similarly to humans, canine mammary cancer represents a heterogeneous group in

terms of morphology and biological behaviour. In the present study we evaluated a

series of canine mammary carcinomas based on a new human classification, initially

based on gene expression profiling analysis. Similarly to human breast cancer, by using

an immunohistochemistry surrogate panel based on five molecular markers (estrogen

receptor, HER2, cytokeratin 5, p63 and P-cadherin), we were able to classify canine

mammary carcinomas into four different subtypes: luminal A (ER+/HER2-; 44.8%),

luminal B (ER+/HER2+; 13.5%), basal (ER-/HER2- and a basal marker positive;

29.2%) and HER2 overexpressing tumours (ER-/HER2+; 8.3%). Luminal A-type

tumours were characterized by lower grade and proliferation rate, whereas basal-type

tumours were mostly high grade, high proliferative and positive for CK5, p63 and P-

cadherin. In addition, as in humans, basal subtype was significantly associated with

shorter disease-free and overall survival rates and we propose canine mammary

carcinomas as a suitable natural model for the study of this particular subset of human

carcinomas.

Keywords: canine, mammary carcinoma, immunohistochemistry, classification

149

Introduction

Mammary gland tumours are the most commonly occurring neoplasm in the female dog and represent a remarkably heterogeneous group in terms of morphology and biological behaviour [32, 43]. About half of canine mammary tumours are considered malignant and the identification of reliable prognostic factors is essential in order to estimate the individual risk of unfavourable clinical outcome [8, 29].

Several studies have recognized some reliable prognostic factors such as tumour size, histologic type, histologic grade and lymph node status [19, 30, 31]. Moreover, in recent literature we found an increasing number of investigations searching for suitable prognostic markers for canine mammary cancer [54], including proliferation markers [25], hormone receptors [23], p53 and Human Epidermal Growth Factor Receptor 2 (HER2) [21, 24] and adhesion molecules [14, 26], among others. The clinical experience is still limited, however, and reliable results of prospective studies are not always available.

Human and canine mammary cancer studies based on single molecular markers probably cannot accurately account for the heterogeneity of this disease [39]. Given the large number of cellular events involved in cell growth, differentiation, proliferation, invasion and metastases [4], the investigation of multiple molecular alterations in concert has been assuming great importance, due to the introduction of high-throughput technologies [39]. In fact, recent gene expression profiling studies on human breast tumours have identified distinct molecular subtypes of breast carcinomas which differ in their pathobiology and clinical outcomes [36, 47, 48]. Sorlie *et al.* [48] analyzed the expression profiles of 115 sporadic breast tumour samples and categorized them into five main groups: luminal A, luminal B, HER2-overexpressing, basal-like and normal breast tissue-like. Luminal A and B subtypes are based on the expression of estrogen receptor (ER), usually with luminal cytokeratin (CK) expression whereas the basal-like subtype is characterized by the absence of hormonal receptors and expression of basal cell markers [5, 33].

Given that gene expression profiling is impractical as a routine diagnostic tool, there are immunohistochemistry surrogate panels proposed that can potentially distinguish breast cancer subtypes [27, 33]. In the present study, we sought to identify phenotypical subtypes in canine mammary cancer with possible clinical implications. To accomplish this goal, we have characterized by immunohistochemical analysis a hundred and two

canine mammary carcinomas based on the immunohistochemical panel proposed by Matos *et al.* [27], which involved the evaluation of five molecular markers (ER, HER2, CK5, p63 and P-cadherin).

Material and methods

Tumour specimens

The present study is based on a series of a hundred and two cases of canine malignant mammary tumours (n=102) selected from the histopathological files of the University of Trás-os-Montes and Alto Douro, Vila Real and from the Institute of Biomedical Science at the University of Porto, Portugal. The material was fixed in 10% neutral formalin and embedded in paraffin wax. Sections (3 μ m) were cut and stained with haematoxylin and eosin (HE) for histological examination, or used to perform immunohistochemistry.

Follow up data

Sixty nine cases (n=69) had available follow up data, with a median overall survival time of 15 months (range 5-74 months). Overall survival (OS) was defined as the period between surgery and animal natural death or euthanasia due to cancer. Disease-free survival (DFS) was defined as the period of time between surgery and recurrent or metastatic disease. During the follow up period, according to the referring surgeons, 35 animals died or euthanized due to metastatic disease and/or local recurrence.

Histological Examination

Tumours were diagnosed according to the WHO criteria for canine mammary neoplasms [31]. Clinicopathological variables included in the present study were: age, ovariohysterectomy status, contraceptive administration, tumour size, tumour location, tumour histological type and grade, presence of intra-tumoral necrosis, presence of vascular invasion and presence of lymph node metastasis.

Tumours were evaluated for grade in accordance with the Nottingham method for human breast tumours [11], based on the assessment of three morphological features: tubule formation, nuclear pleomorphism and mitotic counts. Each of these features was scored on a scale of 1 to 3 to indicate whether it was present in slight, moderate or

marked degree, giving a putative total of 3-9 points. Grade was allocated by an arbitrary division of the total points as follows: grade I (well differentiated), 3, 4 or 5 points; grade II (moderately differentiated), 6 or 7 points; and grade III (poorly differentiated), 8 or 9 points.

Immunohistochemistry

Tissue sections were incubated with primary monoclonal antibodies against ER, HER2, CK5, p63, P-cadherin and Ki-67. Table 1 summarizes the antibodies used and the staining procedures adopted for each antibody. Antigen retrieval was carried out by microwave treatment in a 10mM citrate buffer, pH 6.0, with the exception of Pcadherin, which was performed with an EDTA buffer, pH 8.0 (Lab Vision) in a boiling bath, during 20 minutes. For Ki-67, slides were previously incubated with 0.2 mg/mL trypsin (Merck) in phosphate buffered saline (PBS) for 10 min at 37°C. After cooling (20 minutes at room temperature), the sections were immersed in 3% hydrogen peroxide (H₂O₂) and distillated water during 30 minutes to block endogenous peroxidase activity. All slides were then incubated with a blocking serum (Lab Vision, USA) for 10 min and then incubated with the specific antibody. After incubation, slides sections were incubated with biotinylated secondary antibody, followed by streptavidin-conjugated peroxidase (Lab Vision, USA), except for ER and HER2. For these antibodies, a polymeric labelling methodology was used as a detection system (Novolink Polymer Detection System, Novocastra, Newcastle, United Kingdom), following the manufacturer's instructions. Subsequently, the colour was developed with 3,3diaminobenzidine tetrahydrochloride (DAB) and slides were counterstained with Gill's hematoxylin, dehydrated, and mounted for evaluation by light microscopy.

Adjacent normal mammary tissues were used as internal positive controls for CK5, p63, P-cadherin (basal and myoepithelial cells) and Ki-67. As positive controls, we also used canine uterus sections for ER and a human breast carcinoma with proved amplification (by FISH) and overexpression for HER2. Negative controls were carried out by replacing the primary antibody with PBS.

Table 1. Primary monoclonal antibodies and immunostaining protocols used.

Antibody	Origin	Clone	Dilution	Pretreatment	Incubation
ER	Novocastra, UK	NCL-LH2	1:40	Microwave	2h
HER2	Novocastra, UK	NCL-CB11	1:40	Microwave	Overnight
CK5	Neomarkers, USA	XM26	1:25	Microwave	Overnight
P63	Neomarkers, USA	4A4	1:150	Microwave	Overnight
PCAD	BD Transduction, USA	56	1:50	Water bath, 98°C	Overnight
Ki-67	Dako, Denmark	Mib1	1:50	Trypsin + Microwave	Overnight

Evaluation of the immunohistochemical data

Nuclear ER immunoreactivity was considered positive when more than 10% of the neoplastic cells expressed this marker. To evaluate HER2 expression, Herceptest scoring system was applied (0=no membrane staining or <10% of cells stained; 1+=incomplete membrane staining in >10% of cells; 2+=>10% of cells with weak to moderate complete membrane staining; and 3+=strong and complete membrane staining in >10% of cells), with 2+ and 3+ cases considered positive. As for CK5 and p63, a semi-quantitative analysis was performed as follows: 0, <10% positive cells; 1, 10-50% positive cells and 2, >50% positive cells, with a cytoplasmic (CK5) or nuclear (p63) pattern of cellular distribution. Ki-67 and P-cadherin immunostainings were evaluated as previously described in canine tissues [14, 15]. CK5, p63 and P-cadherin were considered positive when more than 50% of the neoplastic cells expressed each marker.

Statistical Analysis

For statistical analysis, association between subtype tumour groups and continuous variables (mitotic and Ki-67 indices) was assessed with non parametric Kruskal-Wallis test. Associations between groups and categorical variables such as tumour size, histological type, histological grade and invasion were performed using the chi-square test. Survival curves were generated by the Kaplan-Meier method and the survival rates were compared using the log-rank test. All statistical analysis was performed using SPSS 11.5 statistical software. A *P* value <0.05 was considered statistically significant.

Results

Patients and tumour characteristics

The mean age of dogs at the time of surgical removal of tumours was 9.7 ± 2.5 years (range 4–16 years of age). The mean maximum tumour diameter was 4.21 ± 3.4 cm (range 0.5-18cm), with tumours more frequently located in caudal mammary glands (n=36; 59%). In 10 (15.2%) out of the 66 female dogs with available clinical information, ovariohysterectomy (OHE) was performed prior to the removal of mammary tumours. Contraceptive administration was confirmed in 8 (13.8%) cases. Histological evaluation yielded 39 (42.4%) simple carcinoma, 41 (44.6%) complex carcinoma and 12 (13%) carcinosarcoma subtypes. According to the Nottingham method, tumours were classified as grade I (n=14, 15.2%), grade II (n=33, 35.9%) and grade III (n=45, 48.9%). Necrosis was present in 87 (94.6%) and vascular invasion in 51 (55.4%) cases. Lymph nodes were available in 49 cases, with confirmed metastasis in 26 cases (53.1%).

Immunohistochemistry profiles in canine tumours

The results of the immunohistochemical analysis performed for ER, HER2, CK5, p63 and P-cadherin are shown in Table 2 and Fig. 1. The immunohistochemical detection of ER was reliable in 96 cases: the remaining tumours have lost ER antigenicity (adjacent mammary gland was negative) and were excluded. Immunohistochemical evaluation of HER2 and P-cadherin was available in 100 and 96 cases, respectively.

ER and p63 positive cases showed the characteristic nuclear staining, whereas CK5 positive ones showed a cytoplasmic pattern of expression. HER2 positive tumours showed a membranous staining and P-cadherin positive tumours showed a cytoplasmic and/or membranous immunostaining. We observed that 58.3% of canine mammary carcinomas in our series were ER positive, whereas 21% were HER2 positive (2+ and 3+). A positive basal cell marker expression was present in 32.4% tumours for both CK5 and p63 and in 42.8% tumours for P-cadherin.

According to Nielsen *et al.* [33], we classified each tumour based on its ER and HER2 expression. A total of 96 cases were immunohistochemically interpretable to allow sample characterization into one of five categories (Table 3). If a tumour was ER

positive, it was classified as luminal; moreover, we distinguish luminal A and B on the basis of HER2 overexpression. If a tumour was ER positive and HER2 negative (0 or 1+), it would be classified as luminal A (ER+/HER2-); however, if it was ER and HER2 positive, it would be classified as luminal B (ER+/HER2+). If a tumour was ER negative and HER2 positive (ER-/HER2+), it would be classified as HER2-overexpressing, and if it was both ER- and HER2- negative but positive for at least one basal marker (CK5 and/or p63 and/or P-cadherin), it would be classified as basal (ER-/HER2-). If a tumour did not show expression for any of these markers, it would be classified as negative (null phenotype) and would not be considered in the remaining analyses.

Using this definition, we observed that luminal A and B subtypes comprised 44.8 and 13.5% of all tumours, respectively; basal subtype comprised 29.2%; HER2 overexpressing subtype represented 8.3% and negative/null phenotype accounted for 4.2% in this tumour series (Table 3).

Table 2. Imunohistochemical results in the present study.

Molecular marker	Positive staining n (%)	Negative staining n (%)
ER^a	56 (58.3)	40 (41.7)
HER2 ^a	21 (21)	79 (79)
CK5	33 (32.4)	69 (67.6)
P63	33 (32.4)	69 (67.6)
$PCAD^{a}$	42 (42.8)	54 (56.3)

^a Immunohistochemical evaluation of ER and P-cadherin was available in 96 cases and HER2 was available in 100 cases.

Table 3. Frequencies of immunohistochemically defined subtypes of canine mammary carcinomas (n=96).

Subtype	ER	HER2	P-CD and/or p63 and/or CK5	Frequency [n (%)]
Luminal A	Positive	Negative	Positive/negative	43 (44.8%)
Luminal B	Positive	Positive	Positive/negative	13 (13.5%)
Basal	Negative	Negative	Positive	28 (29.2%)
HER2-overexpressing	Negative	Positive	Positive/negative	8 (8.3%)
Negative/null phenotype	Negative	Negative	Negative	4 (4.2%)

Statistically strong significant differences between the four groups were observed in this study, when related with some relevant clinicopathological parameters (Table 4). Basal and HER2 overexpressing subtypes were associated with simple or carcinosarcoma histological types, whereas complex carcinomas were mostly of luminal A subtype (P<0.0001). In addition, basal subtype tumours presented higher histological grade, representing 55.6% of grade III tumours (P<0.0001) and were also significantly associated with the presence of vascular invasion (P<0.0001).

Basal marker expression clearly differed across distinct molecular subtypes (Table 5). Basal and HER2-overexpressing tumours demonstrated a higher frequency of the basal cell markers p63 and P-cadherin (P<0.0001 and P=0.001) and CK5 positive tumours were frequently basal subtype tumours (P=0.001). In contrast, luminal pattern was associated with a lower expression of basal markers. In fact, when analysing basal marker expression simultaneously, we found that the majority of luminal tumours were simultaneously negative to CK5, p63 and P-cadherin. All HER2-overexpressing tumours expressed at least one basal marker and the basal subtype tumours showed frequently the expression of two or all basal markers (P<0.0001).

With regard to proliferation indices, luminal A tumours showed lower median mitotic and Ki-67 labelling indices (P=0.001 and P<0.0001, respectively), whereas all other groups were characterized by higher proliferation rates, with basal subtype showing the highest Ki-67 index.

Follow up data revealed that basal subtype was significantly associated with lower overall (P=0.002, Fig. 2A) and disease-free (P=0.01, Fig.2B) survival rates, whereas the other groups showed higher survival rates, including the HER2-overexpressing group.

Discussion

Recently, gene expression profiling has redefined breast cancer taxonomy and identified five distinct subtypes of carcinomas: luminal A, luminal B, normal breast-like, HER2 overexpressing and basal-like [36, 47, 48, 53]. These molecular subtypes not only reflect the heterogeneity of breast carcinomas and the possible different cell lineage pathways in breast carcinogenesis, but also demonstrate the difference in clinical outcome, with basal-like subtype associated with a more aggressive behaviour [1, 47, 48, 52, 53].

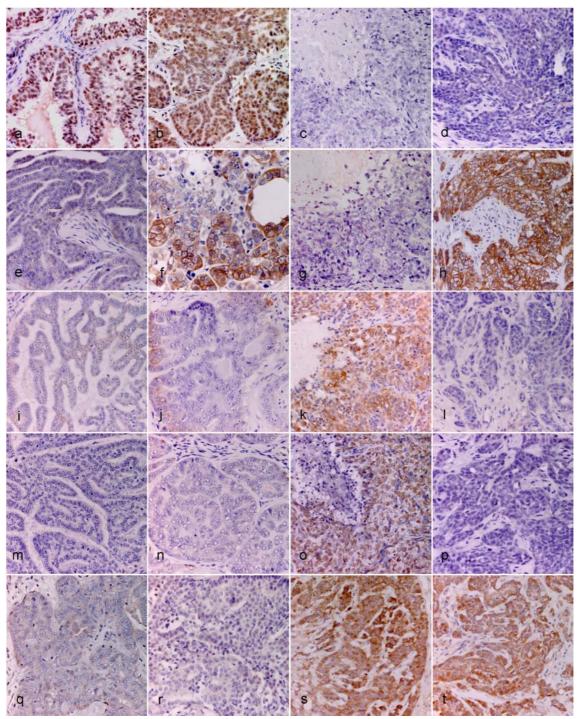


Fig. 1 Immunohistochemical expression of the different proteins studied by IHC in canine mammary carcinomas. **a-d** ER staining; **e-h** HER2 staining; **i-l** CK5 staining; **m-p** p63 staining; **q-t** P-cadherin staining. Each column represents a distinct molecular subtype. From left to right, each column represents luminal A, luminal B, basal and HER2 overexpressing subtypes. [original magnification x400]

Table 4. Association between tumour subtypes and clinicopathological characteristics.

	Luminal A [n (%)]	Luminal B [n (%)]	Basal [n (%)]	HER2 overexpressing [n (%)]	P
Age				<u> </u>	
≤ 9 years old	18 (43.9%)	6 (14.6%)	13 (31.7%)	4 (9.8%)	0.90
> 9 years old	24 (51.1%)	6 (12.8%)	14 (29.8%)	3 (6.4%)	
Tumour size					
<3 cm	17 (53.1%)	6 (18.8%)	8 (25%)	1 (3.1%)	0.37
3-5 cm	14 (46.7%)	4 (13.3%)	8 (26.7%)	4 (13.3%)	
>5 cm	9 (39.1%)	1 (4.3%)	10 (43.5%)	3 (13%)	
Tumour location					
Cranial glands	2 (50%)	0 (0%)	1 (25%)	1 (25%)	0.09
Medial gland	6 (60%)	2 (20%)	1 (10%)	1 (10%)	
Caudal glands	12 (33.3%)	4 (11.1%)	10 (50%)	2 (5.6%)	
Multiple	8 (72.7%)	2 (18.2%)	0 (0%)	1 (20%)	
Ovariohysterectomy					
No	18 (39.1%)	7 (15.2%)	17 (37%)	4 (8.7%)	0.057
Yes, prior to tumour development	9 (90%)	0 (0%)	0 (0%)	1 (10%)	
Yes, performed with mastectomy	6 (60%)	0 (0%)	4 (40%)	0 (0%)	
Contraception					
No	22 (44%)	6 (12%)	17 (34%)	5 (10%)	0.36
Yes	6 (75%)	0 (0%)	2 (25%)	0 (0%)	
Histological type					
Simple carcinoma	9 (23.1%)	8 (20.5%)	17 (43.6%)	5 (12.8%)	< 0.0001
Complex carcinoma	32 (78%)	5 (12.2%)	3 (7.3%)	1 (2.4%)	
Carcinosarcoma	2 (16.7%)	0 (0%)	8 (66.7%)	2 (16.7%)	
Histological grade					
Grade I	14 (100%)	0 (0%)	0 (0%)	0 (0%)	< 0.0001
Grade II	23 (69.7%)	5 (15.2%)	3 (9.1%)	2 (6.1%)	
Grade III	6 (13.3%)	8 (17.8%)	25 (55.6%)	6 (13.3%)	
Necrosis					
Absent	4 (80%)	1 (20%)	0 (0%)	0 (0%)	0.29
Present	39 (44.8%)	12 (13.8%)	28 (32.3%)	8 (9.2%)	
Lymphovascular					
Invasion					.0.000:
Absent	29 (70.7%)	6 (14.6%)	4 (9.8%)	2 (4.9%)	< 0.0001
Present	14 (27.5%)	7 (13.7%)	24 (47.1%)	6 (11.8%)	
Lymph node					
metastasis ^a	12 (56 50/)	6 (26 10/)	2 (120/)	1 (4 20/)	Λ 1
Absent	13 (56.5%)	6 (26.1%)	3 (13%)	1 (4.3%)	0.1
Present	8 (30.8%)	5 (19.2%)	11 (42.3%)	2 (7.7%)	

^aLymph nodes were available in 49 cases.

m 11 f A 1 .	1:00 11	1 1 1		1.0 (1.
Table 5. Association between	different subtypes <i>versu</i>	c hasal marker ex	enression and	nroliteration indices
1 dole 5. 1 issociation between	different subtypes versus	ousui illuikei er	Apression and	prometation maters.

	Luminal A [n (%)]	Luminal B [n (%)]	Basal [n (%)]	HER2 overexpressing [n (%)]	P	
CK5						
Negative	36 (60%)	8 (13.3%)	11 (18.3%)	5 (8.3%)	0.001	
Positive	7 (21.9%)	5 (15.6%)	17 (53.1%)	3 (9.4%)		
P63						
Negative	32 (53.3%)	13 (21.7%)	11 (18.3%)	4 (6.7%)	< 0.0001	
Positive	11 (34.4%)	0 (0%)	17 (53.1%)	4 (12.5%)		
P-cadherin						
Negative	26 (56.5%)	10 (21.7%)	9 (19.6%)	1 (2.2%)	0.001	
Positive	13 (31.7%)	3 (7.3%)	18 (43.9%)	7 (17.1%)		
Basal markers						
All negative	21 (77.8%)	6 (22.2%)	0 (0%)	0 (0%)	< 0.0001	
One positive	11(39.3%)	6 (21.4%)	8 (28.6%)	3 (10.7%)		
Two positive	5 (20%)	1 (4%)	15 (60%)	4 (16%)		
All positive	2 (25%)	0 (0%)	5 (62.5%)	1 (12.5%)		
Median Mitotic index ^a	0.44	1.0	0.94	0.7	0.001	
(Min-Max)	(0-1.59)	(0.1-2.99)	(0.1-2.09)	(0.3-1.9)	0.001	
Median Ki-67 index ^a (Min-Max)	17.89 (5.39-56.36)	26.7 (15-44.8)	28.14 (12.10-49.2)	26.4 (22.5-35.86)	<0.0001	

^a Proliferative indices were available in 86 cases.

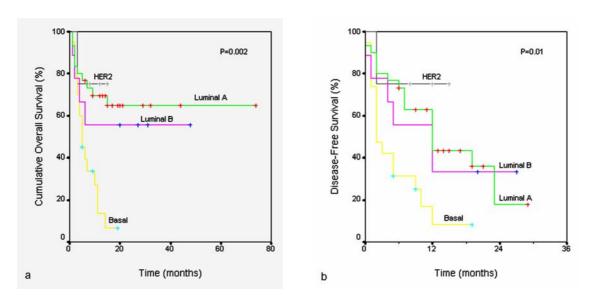


Fig. 2 Kaplan-Meier overall survival (A) and disease-free survival (B) curves of the different subtype groups.

Although gene expression profiling is still considered the "gold standard" for the identification of breast carcinoma subtypes, this technology requires highly sophisticated technical equipment and is not readily available for clinical application or for retrospective studies using formalin fixed, paraffin-embedded samples [39]. For this reason, immunohistochemistry has been used in several studies and the evaluation of a limited panel of immunohistochemical cell markers have shown that breast carcinomas can be subdivided into subgroups remarkably similar to the ones defined by gene expression profiling [1, 3, 22, 27, 33, 38, 52].

In the present study we found in a series of canine mammary tumours, similar findings observed in human breast cancer. We have also identified distinct phenotypical subtypes in a series of canine mammary carcinomas, by using an immunohistochemical panel which included five molecular markers (ER, HER2, CK5, p63 and P-cadherin). Based on ER/HER2 molecular classification, we defined four main subgroups: luminal A (ER+/HER2-, 44.8%), luminal B (ER+/HER2+, 13.5%), basal-like (ER-/HER2-, 29.2%) and HER2 overexpressing (ER-/HER2+, 8.3%). In contrast, Sarli *et al.* [44] have only identified luminal A and B subtypes when studying a series of 39 canine mammary carcinomas. Although using a similar terminology, they used a distinct panel of molecular markers and the adopted classification was not identical, with luminal subtype defined as CK19 positive tumours, regardless of hormonal status (luminal A, HER2- and luminal B, HER2+), and HER2 overexpressing and basal-like subtypes defined as CK19 negative tumours, HER2+ and HER2-, respectively.

In the current study, we found statistically strong significant differences between the four groups, with ER positive luminal A tumours more frequently associated with complex tumour type, low histological grade, less invasive and low proliferative tumours, whereas basal-like and HER2 overexpressing subtypes were associated with simple and carcinosarcoma tumour types, high histological grade, lymphovascular invasion and high proliferation, features that are in accordance to the ones described in recent human literature for basal-like cancers [20, 22, 27, 40].

CK5, p63 and P-cadherin are proteins that are expressed early in epithelial differentiation and may contribute to a committed stem cell and/or progenitor phenotype [6, 7, 9, 35]. In this study, we demonstrate that these markers are upregulated in the basal subtype, similarly to Matos *et al.* previous results [27]. In fact, the basal subtype rarely expressed just one basal marker but frequently expressed them simultaneously,

which suggests a more undifferentiated profile. HER2-overexpressing subtype was also characterized by an up-regulation of basal markers, confirming some human breast studies which suggested that HER2-overexpressing tumours should be included in a bona fide basal-like subclass [5, 27]. In contrast, the majority of luminal tumours in our series were simultaneously negative for basal cell markers, with some cases showing basal marker expression, which was also described by some authors, who reported tumours co-expressing basal CK and hormone receptors or HER2 [40, 50].

Similarly to human breast cancers, in this study we further demonstrate the molecular heterogeneity of canine mammary cancer. A "hierarchy or stem cell" model of breast cancer oncogenesis has been proposed to elucidate the observed functional heterogeneity of tumours. In this model, transformation occurs in a stem cell, or in a progenitor "highly proliferating" cell, and expansion proceeds until various maturation stages, depending on the genomic alterations. Specific genetic alterations would lead to distinct cellular transcriptomic programmes, including the change of hormonal receptors and CK expression pattern, characterising distinct subgroups of breast carcinomas (5, 8, 39].

Survival analysis revealed that distinct subtypes were associated with different clinical outcomes, with basal subtype associated with lower survival rates, similarly to human breast cancer studies [36, 47, 48]. These results also corroborate a previous study in canine mammary cancer performed by Griffey *et al.* [16] which firstly described basal carcinomas as having poor prognostic features. Despite many different studies associating basal-like tumours with a more aggressive clinical history and shorter survival [3, 33, 37, 47, 48, 49, 52], others did not find such a prognostic significance [12, 18]. These variations are probably related to differences between studies in patient cohorts, analytic methods and, most importantly, the immunohistochemical definitions of basal-like breast cancer [39]. Recently, Tang *et al.* [51] comparing several classifications with similar terminology but different definitions (such as ER/HER and triple negative classification) concluded that these classifications are related but not interchangeable.

In contrast to basal subgroup, luminal and HER2 overexpressing subtypes showed increased survival rates. The fact that luminal tumours were associated with a better prognosis is not surprising, since ER positive human breast carcinomas are usually associated with a more favourable clinical outcome. In veterinary pathology, however,

the prognostic value of ER in canine mammary cancer is still a matter of debate. Previous studies using biochemical [45] and immunohistochemical [34] methodologies have demonstrated the prognostic value of ER, but others have failed this confirmation [23, 28]. The observed discrepancies between different studies are probably related with sample selection, differences in antibodies, staining procedure and evaluation or sensitivity of the detection system. In our series, luminal tumours were mostly of complex type, which comes in accordance to previous canine studies reporting complex carcinomas as being more likely ER positive [15, 23, 28]. Given that this tumour type is generally associated with a better clinical outcome, its high proportion in luminal subtype groups is probably in part responsible for their favourable prognosis.

Despite HER2 recognition as a prognostic factor in human breast cancer [41, 46], the significance of HER2 overexpression in dogs with mammary carcinoma is still unclear. Some studies have shown that either HER2 amplification or protein overexpression are present in canine mammary carcinomas [2, 42], while others found no gene amplification [24]. Similarly to previous studies [10, 24], HER-2 overexpressing tumours were found usually associated with established indicators of poor prognosis such as large tumour size, high histologic grade, invasion, simple histologic type and high proliferative indices. However, Kaplan-Meier analysis revealed that this subtype was related with a more favourable clinical outcome, findings that are in contrast with human studies, which describe similar survival rates for HER2 overexpressing and basal-like subtypes [36, 47, 48], and are probably related to the small number of cases that comprise the HER2 overexpressing subtype. However, a recent study performed by Hsu *et al.* [17] revealed that HER2 overexpression in canine malignant mammary tumours is associated with higher survival rates. Additional large-scale studies are warranted to further explore the value of HER2 in canine mammary carcinomas.

In conclusion, as in humans, our study defined distinct molecular phenotypes in canine mammary carcinomas based on immunohistochemical analysis. Moreover, we have identified a basal-like subtype representing almost 30% of our series, which was associated with a more aggressive clinical behaviour. We believe that canine mammary carcinomas would be suitable natural models for the study of this particular subset of carcinomas. However, more studies are needed regarding the prognostic value of these immunohistochemically determined subtypes in canine mammary cancer.

Acknowledgements

The authors thank Prof. Fátima Gärtner (Institute of Biomedical Science at the University of Porto, Portugal) for the contribution of some cases included in this study. We also thank Mrs Lígia Bento for expert technical assistance. This work was supported by the Centro de Ciência Animal e Veterinária (CECAV) – University of Trás os Montes e Alto Douro (UTAD), Vila Real, Portugal and by Portuguese Science and Technology Foundation, project POCTI/CVT/57795/2004.

References

- Abd El-Rehim DM, Pinder SE, Paish CE, Bell J, Blamey RW, Robertson JF, Nicholson RI, Ellis IO (2004) Expression of luminal and basal cytokeratins in human breast carcinoma. J Pathol 203: 661–671
- 2. Ahern TE, Bird RC, Church Bird AE, Wolfe LG (1996) Expression of the oncogene c-*erb*B-2 in canine mammary cancers and tumor-derived cell lines. Am J Vet Res 57:693–696
- Banerjee S, Reis-Filho JS, Ashley S, Steele D, Ashworth A, Lakhani SR, Smith IE. (2006)
 Basal-like breast carcinomas: clinical outcome and response to chemotherapy. J Clin Pathol 59:729–735
- 4. Beckmann MW, Niederacher D, Schnürch HG, Gusterson BA, Bender HG (1997) Multistep carcinogenesis of breast cancer and tumour heterogeneity. J Mol Med 75:429–439
- 5. Birnbaum D, Bertucci F, Ginestier C, Tagett R, Jacquemier J, Charafe-Jauffret E (2004) Basal and luminal breast cancers; basic or luminous? Int J Oncol 25:249–258.
- 6. Boecker W, Buerger H (2003) Evidence of progenitor cells of glandular and myoepithelial cell lineages in the human adult female breast epithelium: a new progenitor (adult stem) cell concept. Cell Prolif 36(Suppl 1):73–84
- 7. Boecker W, Moll R, Poremba C, Holland R, Van Diest PJ, Dervan P, Burger H, Wai D, Ina Diallo R, Brandt B, Herbst H, Schmidt A, Lerch MM, Buchwallow IB (2002) Common adult stem cells in the human breast give rise to glandular and myoepithelial cell lineages: a new cell biological concept. Lab Invest 82:737–746
- 8. Dick JE (2003) Breast cancer stem cells revealed. Proc Natl Acad Sci U S A 100:3547–3549
- DiRenzo J, Signoretti S, Nakamura N, Rivera-Gonzalez R, Sellers W, Loda M, Brown M
 (2002) Growth factor requirements and basal phenotype of an immortalized mammary epithelial cell line. Cancer Res 62:89–98
- 10. Dutra AP, Granja NVM, Schmitt FC, Cassali GD (2004) c-erbB-2 expression and nuclear pleomorphism in canine mammary tumors. Braz J Med Biol Res 37:1673-1681
- Elston CW, Ellis IO (1998) Assessment of histological grade. In: Elston CW and Ellis IO (Eds). Systemic Pathology. The Breast, 3rd Edn. Churchill Livingstone, London, UK, pp 365-384
- 12. Fulford LG, Reis-Filho JS, Ryder K, Jones C, Gillett CE, Hanby A, Easton D, Lakhani SR (2007) Basal-like grade III invasive ductal carcinoma of the breast: patterns of metastasis and long-term survival. Breast Cancer Res 9:R4
- 13. Gama A, Paredes J, Albergaria A, Gartner F, Schmitt FC (2004) P-cadherin expression in canine mammary tissues. J Comp Pathol 130:13-20

- 14. Gama A, Paredes J, Gartner F, Alves A, Schmitt FC (2008) Expression of E-cadherin, P-cadherin and β-catenin in canine malignant mammary tumours in relation to clinicopathological parameters, proliferation and survival. Vet J 177:45-53.
- 15. Geraldes M, Gärtner F, Schmitt F (2000) Immunohistochemical study of hormonal receptors and cell proliferation in normal canine mammary glands and spontaneous mammary tumours. Vet Rec 146:403–406
- 16. Griffey SM, Madewell BR, Dairkee SH, Hunt JE, Naydan DK, Higgins RJ (1993) Immunohistochemical reactivity of basal and luminal epithelium-specific cytokeratin antibodies within normal and neoplastic canine mammary glands. Vet Pathol 30:155-161
- 17. Hsu W-L, Huang H-M, Liao J-W, Wong M-L, Chang S-C (2007) Increased survival in dogs with malignant mammary tumours overexpressing HER-2 protein and detection of a silent single nucleotide polymorphism in the canine HER-2 gene. Vet Journal. Doi:10.1016/j.tvjl.2007.10.013
- 18. Jumppanen M, Gruvberger-Saal S, Kauraniemi P, Tanner M, Bendahl PO, Lundin M, Krogh M, Kataja P, Borg A, Fernö M, Isola J (2007) Basal-like phenotype is not associated with patient survival in estrogen receptor negative breast cancers. Breast Cancer Res 9: R16.
- 19. Karayannopoulou M, Kaldrymidou E, Constantinidis TC, Dessiris A (2005) Histological grading and prognosis in dogs with mammary carcinomas: application of a human grading method. J Comp Pathol 133:246–252
- 20. Kim MJ, Ro JY, Ahn SH, Kim HH, Kim SB, Gong G. (2006) Clinicopathologic significance of the basal-like subtype of breast cancer: a comparison with hormone receptor and Her2/neu-overexpressing phenotypes. Hum Pathol 37:1217-1226
- 21. Lee CH, Kim WH, Lim JH, Kang MS, Kim DY, Kweon OK (2004) Mutation and overexpression of p53 as a prognostic factor in canine mammary tumors. J Vet Sci 5:63–69
- 22. Livasy CA, Karaca G, Nanda R, Tretiakova MS, Olopade OI, Moore DT, Perou CM. (2006) Phenotypic evaluation of the basal-like subtype of invasive breast carcinoma. Mod Pathol 19:264–271
- 23. Martin de las Mulas J, Millán Y, Dios R (2005) A prospective analysis of immunohistochemically determined Estrogen Receptor α and Progesterone Receptor Expression and host and tumor factors as predictors of disease-free period in mammary tumors of the dog. Vet Pathol 42:200–212
- 24. Martin de las Mulas J, Ordás J, Millán Y, Fernández-Soria V, Ramón y Cajal S (2003) Oncogene HER-2 in canine mammary gland carcinomas: an immunohistochemical and chromogenic in situ hybridization study. Breast Cancer Res Treat 80:363–367

- 25. Matos AJ, Lopes CC, Faustino AM, Carvalheira JG, dos Santos MS, Rutteman GR, Gartner F (2006) MIB-1 labelling índices according to clínico-pathological variables in canine mammary tumours: a multivariate study. Anticancer Res 26:1821-1826
- 26. Matos AJ, Lopes CC, Faustino AM, Carvalheira JG, Rutteman GR, Gärtner MF (2007) E-cadherin, beta-catenin, invasion and lymph node metastases in canine malignant mammary tumours. APMIS 115: 327-334
- 27. Matos I, Dufloth R, Alvarenga M, Zeferino LC, Schmitt F (2005) p63, cytokeratin 5, and P-cadherin: three molecular markers to distinguish basal phenotype in breast carcinomas. Virchows Arch 447:688-694
- 28. Millanta F, Calandrella M, Bari G, Niccolini M, Vannozzi I, Poli A (2005) Comparison of steroid receptor expression in normal, dysplastic, and neoplastic canine and feline mammary tissues. Res Vet Sci 79:225-232
- 29. Misdorp W (2002) Tumours of the mammary gland. In: Meuten DJ (Ed) Tumors in Domestic Animals, 4th Edn. Iowa State Press, Blackwell Publishing Company, pp 575-606
- 30. Misdorp W, Else RW, Hellmén E, Lipscomb TP (1999) Histological Classification of Mammary Tumors of the Dog and the Cat, Vol VII, 2nd series. Armed Forces Institute of Pathology, American Registry of Pathology, Washington D.C., and the World Health Organization Collaborating Center for Worldwide Reference on Comparative Oncology, pp 1-59
- 31. Misdorp W, Hart AA (1976) Prognostic factors in canine mammary cancer. J Natl Cancer Inst 56:779-786
- 32. Nerurkar VR, Chitale AR, Jalnakurpar BV, Naik SN, Lalitha VS (1989) Comparative pathology of canine mammary tumours. J Comp Pathol 101:389-397
- 33. Nielsen TO, Hsu FD, Jensen K, Cheang M, Karaca G, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslen LA, Ragaz J, Gown AM, Gilks CB, van de Rijn M, Perou CM (2004) Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. Clin Cancer Res 10:5367–5374.
- 34. Nieto A, Peña L, Perez-Alenza MD, Sanchez MA, Flores JM, Castaño M (2000) Immunohistologic detection of estrogen receptor alpha in canine mammary tumors: clinical and pathologic associations and prognostic significance. Vet Pathol 37:239–247.
- 35. Peralta Soler A, Knudsen KA, Salazar H, Han AC, Keshgegian AA (1999) P-cadherin expression in breast carcinoma indicates poor survival. Cancer 86:1263-1272.
- 36. Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lonning PE,

- Borresen-Dale AL, Brown PO, Botstein D (2000) Molecular portraits of human breast tumours. Nature 406:747–752
- 37. Rakha EA, El-Rehim DA, Paish C, Green AR, Lee AH, Robertson JF, Blamey RW, Macmillan D, Ellis IO. (2006a) Basal phenotype identifies a poor prognostic subgroup of breast cancer of clinical importance. Eur J Cancer 42:3149-3156
- 38. Rakha EA, El-Sayed ME, Green AR, Paish EC, Lee AH, Ellis IO (2007) Breast carcinoma with basal differentiation: a proposal for pathology definition based on basal cytokeratin expression. Histopathology 50: 434–438
- 39. Rakha EA, El-Sayed ME, Reis-Filho J, Ellis IO (2008) Patho-biological aspects of basal-like breast cancer. Breast Cancer Res Treat. Doi 10.1007/s10549-008-9952-1
- 40. Rakha EA, Putti TC, Abd El-Rehim DM, Paish C, Green AR, Powe DG, Lee AH, Robertson JF, Ellis IO (2006b) Morphological and immunophenotypic analysis of breast carcinomas with basal and myoepithelial differentiation. J Pathol 208:495–506.
- 41. Revillion F, Bonnetarre J, Peyrat JP (1998) ErbB-2 oncogene in human breast cancer and its clinical significance. Eur J Cancer 34:791–808
- 42. Rungsipipat A, Tateyama S, Yamaguchi R, Uchida K, Miyoshi N, Hayashi T (1999) Immunohistochemical analysis of c-yes and c-erbB-2 oncogene products and p53 tumor suppressor protein in canine mammary tumors. J Vet Med Sci 61:27–32
- 43. Rutteman GR, Withrow SJ, MacEwen EG (2001) Tumors of the mammary gland. In: Withrow SJ and MacEwen BR (ed) Small Animal Clinical Oncology, WB Saunders Company, Philadelphia, pp 455–477
- 44. Sarli G, Sassi F, Brunetti B, Benazzi C (2007) Luminal-like A and B types in canine mammary carcinomas. 25th Annual Meeting ESVP, Munich (Germany), pp. 187.
- 45. Sartin EA, Barnes S, Kwapien RP, Wolfe LG (1992) Estrogen and progesterone receptor status of mammary carcinomas and correlation with clinical outcome in dogs. Am J Vet Res 53:2196–2200
- 46. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL (1987) Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. Science 235:177–182
- 47. Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, Rijn MV, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Lonning PE, Borresen-Dale AL (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98:10869–10874
- 48. Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lonning PE, Brown PO, Borresen-Dale AL, Botstein

- D (2003) Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 100:8418–8423.
- 49. Sotiriou C, Neo SY, McShane LM, Korn EL, Long PM, Jazaeri A, Martiat P, Fox SB, Harris AL, Liu ET (2003) Breast cancer classification and prognosis based on gene expression profiles from a population-based study. Proc. Natl Acad. Sci. USA 100:10393–10398.
- 50. Tan DS, Marchió C, Jones RL, Savage K, Smith IE, Dowsett M, Reis-Filho JS (2008) Triple negative breast cancer: molecular profiling and prognostic impact in adjuvant anthracycline-treated patients. Breast Cancer Res Treat. 111:27-44
- 51. Tang P, Wang J, Bourne P (2008) Molecular classifications of breast carcinoma with similar terminology and different definitions: are they the same? Hum Pathol 39:506-513
- 52. van de Rijn M, Perou CM, Tibshirani R, Haas P, Kallioniemi O, Kononen J, Torhorst J, Sauter G, Zuber M, Kochli OR, Mross F, Dieterich H, Seitz R, Ross D, Botstein D, Brown P (2002) Expression of cytokeratins 17 and 5 identifies a group of breast carcinomas with poor clinical outcome. Am J Pathol 161:1991–1996.
- 53. van 't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M, Peterse HL, van der Kooy K, Marton MJ, Witteveen AT, Schreiber GJ, Kerkhoven RM, Roberts C, Linsley PS, Bernards R, Friend SH (2002) Gene expression profiling predicts clinical outcome of breast cancer. Nature 415:530–536
- 54. Zaidan Dagli M (2008) The search for suitable prognostic markers for canine mammary tumors: A promising outlook. Vet J 177:3-5

Chapter VII

General Discussion and Concluding Remarks

General Discussion and Concluding Remarks

Until recently, the main role of the veterinary pathologist laid on the establishment of a basic diagnosis. In the context of canine mammary tumours, apart from the examination of regional lymph nodes for the presence or absence of metastasis it was unusual for any other prognostic information to be supplied, or indeed, requested. Nowadays, veterinary oncologists want to know not only the standard histological features of mammary tumours, such its type and grade, but also relevant information concerning the prognosis of an individual animal.

Canine mammary cancer represents a very important disease and there has been a persistent drive in order to identify reliable prognostic factors. The prognostic value of clinicopathological parameters in canine mammary tumours is still a matter of debate and the controversial results obtained in a rather small number of prognostic studies also prevent the routine use of molecular markers. The conducted studies included in this thesis aimed to study the prognostic impact of several clinicopathological and molecular characteristics of canine mammary gland tumours.

With regard to clinicopathological parameters, several variables were found of prognostic value on univariate analysis, including tumour size, ulceration, histological type, tumour growth pattern, histological grade, stromal/vascular invasion and lymph node status. Thus, large sized tumours, skin ulceration, simple and carcinosarcoma histological type, infiltrative tumour growth and the presence of stromal/vascular invasion and node metastasis were significantly associated with reduced survival rates. In addition, high proliferation indices (both mitotic and Ki-67 indices) were also associated with poorer survival times.

The histological heterogeneity observed in canine mammary neoplasms presents considerable difficulties in the design of a classification system that will assure reproducibility of a prognostically meaningful categorization (Gilbertson *et al.*, 1983). In our study, the categorization of mammary malignant tumours in simple carcinoma, complex carcinoma and carcinosarcoma types has been associated with distinct biological behaviours. As previously observed by de las Mulas and Peña (2004), we have also found some difficulties in the application of the new WHO tumour classification, mostly in the definition of carcinoma in benign tumour. In the present study, these tumours were included in the complex carcinoma group, given that they

were characterized by an extensive complex phenotype, although some minor areas of metaplastic changes occurred in a few cases. In addition, these tumours showed very similar clinical behaviour with other complex carcinomas. As for simple carcinoma group, it harboured several distinct morphological entities and it remains our goal to gather a large number of tumour cases in order to perform a prospective analysis considering separate histological types. Even so, simple carcinomas and carcinosarcomas were strongly associated with shorter survival rates, when compared to complex carcinomas. Ongoing work includes the establishment and characterization of canine mammary cell lines of epithelial and myoepithelial differentiation, in order to unravel myoepithelial cell putative role as a natural invasion tumour suppressor in canine mammary tumours.

Post-surgical prognosis has been the subject of several prospective studies, but only a few multivariate analyses have been carried out to determine which clinicopathological parameters have independent prognostic value (Hellmén *et al.*, 1993; Peña *et al.*, 1998; Nieto *et al.*, 2000; Chang *et al.*, 2005; Itoh *et al.*, 2005; de Matos *et al.*, 2006). On multivariate Cox-regression analysis, only lymph node status represented an independent prognostic factor, which stresses the critical importance of the standard availability of regional lymph nodes. Based on our findings, we confirm that veterinary pathologists are in an ideal position to supply clinical colleagues with a substantial amount of useful prognostic information, just from the routine examination of canine mammary tumours.

When searching for suitable prognostic markers, it is important to focus on cell adhesion properties, which might be related to the cells' ability to detach from neighbouring cells leading to the first steps of invasion and metastasis (Zaidan Dagli, 2008). Inspired by this idea, we have investigated the immunohistochemical expression of the cell adhesion molecules E-cadherin, P-cadherin and β -catenin in a series of canine mammary malignant tumours, and their relationship with clinicopathological parameters, proliferation and survival. Our study revealed that a reduced E-cadherin and β -catenin expression was significantly associated with several aggressive clinicopathological features, such as high histological grade and invasion, as shown in previous canine mammary studies (Brunetti *et al.*, 2005; Matos *et al.*, 2006; de Matos *et al.*, 2007). In addition, abnormal E-cadherin and β -catenin expression was significantly associated with poorer survival times, in contrast to Brunetti *et al.* findings (Brunetti *et al*

al., 2005). From our results, we confirmed that an altered expression of the cadherincatenin complex is associated with cell invasion and might play a central role in canine mammary tumour progression. However, to further validate E-cadherin and β -catenin molecules as prognostic markers, additional studies are warranted, including a larger series of tumours and a longer follow-up period.

Similarly to canine mammary gland tumour findings, E-cadherin expression studies have revealed some contradictory results in human breast cancer, where some authors failed to reveal a prognostic value. In fact, some breast cancers with aggressive characteristics present high levels of E-cadherin and many metastases are E-cadherin-positive (Shiozaki *et al.*, 1996; Howard *et al.*, 2005). Thus, the expression of E-cadherin is likely to be dynamic; it is possible that temporary or localized downregulation of E-cadherin promotes detachment of cells from the primary tumour and invasion into the local environment and that posterior re-expression of E-cadherin in a new environment might foster their survival as they are carried to a distant site (Knudsen and Wheelock, 2005).

Another consideration is that E-cadherin, even if it is expressed in mammary cancers, is not fully functional unless it forms a complex with catenins and anchors to the cytoskeleton. In this study we have investigated both E-cadherin and β-catenin, considering a separate and a combined expression of these molecules, and we have confirmed that idea, given that the loss of at least one of these proteins was associated with an aggressive tumour phenotype. Additional studies are required in order to shed some light on E-cadherin regulation and signalling in canine mammary tumours. Recent investigations point out to regulatory signals between oestrogen and E-cadherin, with loss of oestrogen resulting in its subsequent repression (Fujita *et al.*, 2003; Park *et al.*, 2008). In fact, it was recently described an association between E-cadherin absence/reduction and a basal-like phenotype in human breast carcinomas (Mahler-Araujo *et al.*, 2008).

As for P-cadherin, to the best of our knowledge, no study had previously studied its prognostic value in canine mammary tissues. P-cadherin expression was only significantly associated with an invasive tumour phenotype, with no association with survival. Our results are discordant with the majority of available studies in human breast cancer, which found P-cadherin significantly associated with several aggressive characteristics, such as high histological grade and proliferation, as well as with a poor

prognosis (Peralta Soler et al., 1999; Gamallo et al., 2001; Arnes et al., 2005). Yet, in this first study we have analysed a relatively small number of cases. Later on, we have expanded our series and we have found P-cadherin expression significantly associated with high histological grade and with a poor prognosis, being strongly associated with a basal-like phenotype. However, although P-cadherin positive carcinomas indeed appear to have a myoepithelial/basal-like transcriptomic programme, this explanation is unlikely to account for the high percentage of P-cadherin expressing tumours, as suggested for human breast. It is easier to accept that some molecular mechanisms would lead to activation of P-cadherin expression (Paredes et al., 2007). In fact, a significant correlation was recently described between P-cadherin expression and hypomethylation of a specific region of the CDH3 promoter, suggesting an important regulatory role for cytosine methylation in the aberrant expression of P-cadherin in breast cancer (Paredes et al., 2005). On the other hand, the lack of ER signalling was found responsible for the increase in P-cadherin, categorizing CDH3 as an oestrogenrepressed gene and pointing to E2 as a key regulator of this cadherin (Paredes et al., 2004). As already discussed, the role of P-cadherin in breast cancer remains incompletely understood. Whether it represents a useful prognostic marker or plays a causal role is open to question, both in canine and in the human species.

The availability of molecular targeted therapies that interfere with specific targets having critical roles in tumour growth and progression is promising for cancer treatment, and the recent availability and US Food and Drug Administration approval of specific EGFR tyrosine kinase inhibitors has increased the interest on this growth factor receptor in human breast cancer studies (Baselga and Arteaga, 2005; Jorgensen *et al.*, 2007; Widakowich *et al.*, 2007). In the present thesis, we report for the first time an immunohistochemical study of EGFR expression in benign and malignant canine mammary gland tumours, confirming previous biochemical findings at the cellular level. Our results have found EGFR expression significantly associated with a malignant tumour phenotype. However, EGFR was not significantly associated with clinicopathological variables, other than animal age and tumour size. In addition, although dogs affected by EGFR-overexpressing malignant tumours showed poorer survival rates compared to dogs harbouring EGFR negative tumours, the differences observed failed to reach statistically significant levels. Given the tendency of positive EGFR cases towards poor prognosis, the possibility exists that a number of dogs might

benefit from EGFR-targeted therapy, as in human breast and lung cancer patients (Lambros *et al.*, 2007; Faratian and Bartlett, 2008). However, to test this hypothesis and to find out if this receptor has prognostic value in canine mammary cancer, additional studies are warranted with a larger series of tumours and follow-up analysis.

Previous studies concerning EGFR expression in canine mammary samples were performed by using biochemical assays (Nerurkar *et al.*, 1987; Donnay *et al.*, 1993; Rutteman *et al.*, 1990, 1994; Donnay *et al.*, 1996), with no differences observed between benign and malignant tumours or with clinicopathological parameters. The distinct approach used between our and previous canine studies makes difficult a direct comparison. Similarly, in human literature, conflicting results are also found, and EGFR is not a consensual prognostic marker. The lack of standardized assessment method and interpretation criteria for EGFR expression may contribute to these apparent contradictory findings (Klijn *et al.*, 1992; Bhargava *et al.*, 2005; Park *et al.*, 2007). At a practical level, there is no universal method for evaluating EGFR expression in human breast tumours and it is of particular interest to disclose if EGFR expression levels can really predict the response to therapy, keeping in mind that EGFR signalling network is comprised of a complex series of interconnecting pathways and each component is likely to affect the level of EGFR signalling output (Ciardiello and Tortora, 2003; Milanezi *et al.*, 2008).

In the near future, we intend to study the underlying mechanisms of EGFR protein overexpression, such as *EGFR* gene amplification, in canine mammary tumours. It is possible that, similarly to the findings in human breast carcinomas, gene amplification does not represent the main mechanism, but this remains unknown in the canine species. On the other hand, several authors have suggested other regulatory mechanisms, namely at transcriptional level (Berquin *et al.*, 2001, 2005; Kersting *et al.*, 2004; Milanezi *et al.*, 2008), which can be related with specific cellular transcriptomic programmes, such as myoepithelial/"basal-like" differentiation (Reis-Filho *et al.*, 2005). Given that EGFR is consistently expressed in normal canine myoepithelial cells, further studies are required to elucidate if EGFR is associated with a basal-like phenotype in canine tumours, as it has been shown in human breast cancer.

Additionally, we have studied the expression of several cell differentiation markers in a series of canine mammary malignant tumours and we have identified a subset of canine mammary carcinomas with lack or reduction of CK19 epithelial expression, a luminal

epithelial cell marker, strongly associated with the expression of basal and/or myoepithelial cell markers. A reduced or absent CK19 expression was also significantly associated with several clinicopathological variables, such as invasion and high histological grade, as well as with high proliferative index.

To our knowledge, this is the first study dealing with CK expression in canine mammary carcinomas in which survival analyses have been performed. These analyses revealed a less favourable disease course for tumours with a basal phenotype than for those with a luminal phenotype, identified by CK19 expression. However, Cox regression multivariate analysis has not revealed CK19 as an independent prognostic variable in canine mammary malignant tumours. It remains to be elucidated if CK19 is merely a reflection of cell differentiation or if it plays an active role during tumour progression. Future studies involving larger number of cases will be needed before such questions can be satisfactorily answered.

In human breast cancer studies, a high level of luminal CK (CK8, CK18 or CK19) immunostaining has been also correlated with a more favourable prognosis (Takei *et al.*, 1995; Schaller *et al.*, 1996; Woelfle *et al.*, 2004; Parikh *et al.*, 2008). However, contradictory findings were described in distinct tumours, since it was recently suggested that the expression of CK19 in pancreatic endocrine tumours may be correlated with a poor prognosis (La Rosa *et al.*, 2007).

The biological significance of the expression of basal CK in poorly differentiated canine mammary carcinomas remains an enigma. One hypothesis is that its expression might indicate derivation from, or toward, myoepithelial cells. In fact, the expression of the smooth muscle actin and calponin has been observed in several carcinoma cases, as previously described in human cancer studies (Santini *et al.*, 1996; Tsuda *et al.*, 2000). Both canine and human mammary cancer has been recognized as a heterogeneous disease in terms of morphology and biological behaviour. Recent studies based on gene expression profiling have reflected human breast cancer heterogeneity at the molecular level and lead the way into modern breast cancer taxonomy (Perou *et al.*, 2000; Sorlie *et al.*, 2001, 2003). The introduction of high-throughput microarray technology allowed the analysis of the expression levels of thousands of genes and the distinction of breast cancer subclasses with diverse biological behaviours (Perou *et al.*, 2000; Sorlie *et al.*, 2001, 2003; Sotiriou *et al.*, 2003). Several immunohistochemical studies have reinforced this novel breast cancer taxonomy at the protein level, by using a surrogate

panel of immunohistochemical markers (Nielsen *et al.*, 2004; Matos *et al.*, 2005; Rakha *et al.*, 2008). In this study, we have applied this novel classification on canine mammary carcinomas and we have identified similar molecular subtypes to the ones found in human breast cancer, by using a surrogate panel of immunohistochemical cell markers, which included ER, HER-2, CK5, P63 and P-cadherin. Four main subtypes were identified: luminal A, luminal B, HER-2 overexpressing and basal-like, with basal-like subtype significantly associated with poor prognosis.

Currently, in the human setting, hormonal and HER-2 status are routinely used to predict prognosis and to determine a patient's specific treatment (Payne *et al.*, 2008). Looking at the situation in dogs, we are far behind from humans; no consensus was reached on the prognostic value of ER or HER-2 status and surgery still remains the treatment of choice for most dogs with mammary gland tumours (Rutteman *et al.*, 2001).

Our study confirmed the reliability of monoclonal antibodies directed against ER and HER-2 proteins in routinely processed formalin-fixed canine mammary tissues, by using a highly sensitive polymeric detection system. We have demonstrated that ER expression (luminal subtypes) is significantly associated with better survival rates, confirming some previous results (Nieto *et al.*, 2000). Future prospective studies are required to validate ER as a prognostic and predictive marker in canine mammary tumours. In fact, the possibility exists that ER may still represent a rationale therapeutic target in canine mammary cancer (Soremno, 2003).

HER-2 status also needs further consideration in subsequent studies. Similarly to EGFR, HER-2 also represents an appealing target molecule for cancer therapy. As already described elsewhere in this thesis, the development of trastuzumab, a humanized monoclonal antibody against HER-2 extracellular domain, settled HER-2 importance as a therapeutic target in human breast cancer (Milanezi *et al.*, 2008), and this fact opens up the possibility of using HER-2 directed therapies in canine metastatic cancer. However, in the present investigation, HER-2 overexpressing tumours were associated with increased survival rates, which is in accordance to the recent results of Hsu and coworkers (2007) in canine tumours but largely contradicts the available human literature (Slamon *et al.*, 1987). Given that our series comprised a small number of HER-2 overexpressing tumours, additional prospective studies are in order to further explore HER-2 prognostic value.

The identification of a subset of carcinomas with a basal-like phenotype confirms, at some point, a previous study performed by Griffey and co-workers (1993), which used this terminology for the first time in canine mammary carcinomas. In the present series, basal-like phenotype was significantly associated with a high histological grade and proliferation, as well as with shorter disease-free and overall survival rates. It is not known to what extent basal-like phenotype represents a signature derived from the cell of origin of these cancers, or more is the result of differentiation from a precursor that is common to all breast cancers and therefore does not reflect histogenesis. In fact, two hypothetical models of mammary oncogenesis showing the potential origin of breast cancer cell have been proposed. The "stochastic" model suggests that clonal tumour expansion originates from any cell, whatever its stage of differentiation, after it has been randomly hit by enough genomic alterations to trigger transformation; the tumor cell acquires a self-renewing capacity but preserves characteristics of its origin. The "hierarchy" or "stem cell" model suggests that transformation occurs in a stem cell, or in a progenitor cell, and expansion proceeds concomitantly to usual maturation until various stages, depending on the identity of genomic alterations (Birnbaum et al., 2004; Rakha et al., 2008).

In human studies, it has been shown that the basal-like subtype is a rather heterogeneous tumour group. Although basal-like tumours are mainly high grade invasive ductal carcinomas, showing morphological characteristic features, such as large central necrotic areas, pushing margins of invasion, high-grade nuclear features and high mitotic index (Livasy *et al.*, 2006; Rakha *et al.*, 2006), other histological types display a basal-like phenotype, namely medullary carcinomas, metaplastic carcinomas and myoepithelial type carcinomas (Rakha *et al.*, 2008). Our next step will be a thorough characterization of this basal-like subtype in canine mammary cancer, performing a large scale study of canine mammary tumours, which will also verify the robustness and independent significance of the present findings.

To further substantiate this classification in canine mammary cancer, the analysis of gene expression levels by high-throughput microarray technology will certainly be the ultimate proof. However, this methodology is expensive, requires access to large numbers of fresh frozen tumour samples and is impractical as a routine diagnostic tool (Rakha *et al.*, 2008). Very recently, a cDNA microarray study was conducted on three canine mammary tumour cell lines, revealing distinct gene expression profiles

pertaining to their phenotype. However, these authors were not able to directly compare canine cell data with human gene sets because of a lack of representation of these genes on the canine microarray. Even so, pathway analysis identified a striking similarity in the pathway profiles of canine mammary tumour, human breast cancer cell line and breast carcinoma intrinsic gene sets (Rao *et al.*, 2008).

A major challenge nowadays is to identify therapeutic targets for the basal-like subtype of human breast cancer, which is not responsive to endocrine therapy or HER-2-directed therapy. Despite the extensive research on this tumour phenotype, the specific genes that drive its aggressive behaviour are poorly understood. A number of attractive gene products have been identified by gene expression profiling in the basal-like cluster, some of them implicated in cellular proliferation, suppression of apoptosis or cell invasion (Perou et al., 2000; Sorlie et al., 2001). These gene products include EGFR, αB-crystallin, TGFβ2, MMP14, cyclin E1 and c-KIT, and although quite diverse, several activate similar signalling pathways such as MAPK-ERK and PI3-kinase-AKT pathways, which may play a central role in the pathogenesis of basal-like carcinomas (Vogelstein and Kinzler, 2004; Yehiely et al., 2006). This specific basal gene profile provides several potential targets for therapy, namely EGFR and its downstream signalling pathways (Siziopikou and Cobleigh, 2007). It is our objective to study EGFR expression and amplification in the basal-like subset of canine carcinomas, in order to investigate this molecule as a potential therapeutic target. Although we consider the present findings as preliminary results, canine mammary carcinomas might represent a suitable natural model for the study of human breast carcinomas, in particular to the basal-like subset, given the putative high percentage of basal carcinomas identified in the canine species.

The main goal of prognostic studies is to identify reliable prognostic factors, which might be used in the routine setting, preferably with therapeutic potential. We hope that our present study on canine mammary tumours would contribute in some way in the understanding of this complex disease. From our results, we stress the following major conclusions:

1. With regard to clinicopathological parameters, univariate analysis revealed that tumour size, ulceration, histological type, tumour growth pattern, histological grade,

stromal/vascular invasion, proliferation indices and lymph node status were significantly associated with poor survival rates, whereas multivariate analysis disclosed lymph node status as the only independent prognostic factor in canine mammary tumours.

- 2. Alterations in the expression of the cadherin-catenin complex represent a common event in canine mammary malignant tumours, with reduced E-cadherin and β -catenin expression significantly associated with several aggressive clinicopathological features, such as high histological grade and invasion, as well as with poorer survival times. P-cadherin was only associated with invasion. Cadherin-mediated cell adhesion molecules might play a central role in canine mammary tumour progression and they may be of prognostic value in canine malignant mammary tumours.
- 3. EGFR is consistently expressed by canine mammary myoepithelial cells. Yet, it is also expressed by epithelial cells, being significantly associated with malignancy. EGFR expression was only associated with animal age and tumour size variables. Female dogs affected by EGFR-overexpressing malignant tumours showed poorer survival rates compared to dogs harbouring EGFR negative tumours, but the differences observed failed to reach statistically significant levels.
- 4. The reduction or absence of luminal CK19 expression was found significantly associated with several clinicopathological variables, such as invasion, high histological grade, high proliferative index, as well as with the expression of basal and/or myoepithelial cell markers. Univariate analysis revealed that the reduction of CK19 was significantly associated with poor survival rates, but multivariate analysis failed to confirm CK19 as an independent prognostic variable in canine mammary tumours.
- 5. By using a surrogate panel of immunohistochemical cell markers (which included ER, HER-2, CK5, P63 and P-cadherin), a novel human classification was applied on canine mammary carcinomas, revealing similar molecular subtypes to the ones found in human breast cancer (luminal A, luminal B, basal-like and HER-2 overexpressing), associated with distinct clinical behaviours. Basal-like phenotype was significantly associated with poor prognostic features, such as high histological grade and proliferation, as well as with shorter disease-free and overall survival rates.

References

- Arnes JB, Brunet JS, Stefansson I, Bégin RL, Wong N, Chappuis PO, Akslen LA, Foulkes WD. (2005). Placental cadherin and the basal epithelial phenotype of BRCA1-related breast cancer. Clin Cancer Res 11:4003-4011.
- Baselga J, Arteaga CL. (2005). Critical update and emerging trends in epidermal growth factor receptor targeting in cancer. J Clin Oncol 23:2445-2459.
- Berquin IM, Dziubinski ML, Nolan GP, Ethier SP. (2001). A functional screen for genes inducing epidermal growth factor autonomy of human mammary epithelial cells confirms the role of amphiregulin. Oncogene 20:4019-4028.
- Berquin IM, Pang B, Dziubinski ML, Scott LM, Chen YQ, Nolan GP, Ethier SP. (2005). Y-box binding protein 1 confers EGFR independence to human mammary epithelial cells. Oncogene 24:3177-3186.
- Bhargava R, Gerald WL, Li AR, Pan Q, Lal P, Ladanyi M, Chen B. (2005). EGFR gene amplification in breast cancer: correlation with epidermal growth factor receptor mRNA and protein expression and HER-2 status and absence of EGFR-activating mutations. Modern Pathol 18:1027-1033.
- Birnbaum D, Bertucci F, Ginestier C, Tagett R, Jacquemier J, Charafe-Jauffret E. (2004). Basal and luminal breast cancers: basic or luminous? Int J Oncol 25:249–258.
- Brunetti B, Sarli G, Preziosi R, Monari I, Benazzi C. (2005). E-cadherin and β-catenin reduction influence invasion but not proliferation in canine malignant mammary tumors. Vet Pathol 42:781-787.
- Chang SC, Chang CC, Chang TJ, Wong ML. (2005). Prognostic factors associated with survival two years after surgery in dogs with malignant mammary tumours: 79 cases (1998-2002). J Am Vet Med Ass 227:1625-1629.
- Ciardiello F, Tortora G. (2003). Epidermal growth factor receptor (EGFR) as a target in cancer therapy: understanding the role of receptor expression and other molecular determinants that could influence the response to anti-EGFR drugs. Eur J Cancer 39:1348-1354.
- De las Mulas JM, Peña L. (2004). Comparison of canine mammary tumours routine histopathological diagnoses given by different groups of pathologists and applicability of the 1999 WHO's classification. Proceedings of 12th Annual Meeting of the Portuguese Society of Animal Pathology jointly with 16th Annual Meeting of the Spanish Society of Veterinary Pathology, pp. 46-47.
- De Matos AJ, Lopes CC, Faustino AM, Carvalheira JG, Dos Santos MS, Rutteman GR, Gärtner MF. (2006). MIB-1 labelling indices according to clinico-pathological variables in canine mammary tumours: a multivariate study. Anticancer Res 26:1821–1826.

- De Matos AJF, Lopes C, Faustino AMR, Carvalheira J, Rutteman GR, Gartner F (2007). E-cadherin, β-catenin, invasion and lymph node metastases in canine malignant mammary tumours. APMIS 115:327-334.
- Donnay I, Rauis J, Wouters-Ballman P, Devleeschouwer N, Leclercq G, Verstegen JP. (1993). Receptors for oestrogen, progesterone and epidermal growth factor in normal and tumorous canine mammary tissues. J Repr Fertil Suppl 47:501-512.
- Donnay I, Devleeschouwer N, Wouters-Ballman P, Leclercq G, Verstegen J. (1996). Relationship between receptors for epidermal growth factor and steroid hormones in normal, dysplastic and neoplastic canine mammary tissues. Res Vet Sci 60:251-254.
- Faratian D, Bartlett J. (2008). Predictive markers in breast cancer-the future. Histopathology 52:91-98.
- Fujita N, Jaye DL, Kajita M, Geigerman C, Moreno CS, Wade PA. (2003). MTA3, a Mi-2/NuRD complex subunit, regulates an invasive growth pathway in breast cancer. Cell 113:207-219.
- Gamallo C, Moreno-Bueno G, Sarrio D, Calero F, Hardisson D, Palacios J. (2001). The prognostic significance of P-cadherin in infiltrating ductal breast carcinoma. Mod Pathol 14:650-654.
- Gilbertson SR, Kurzman ID, Zachrau RE, Hurvitz AI, Black MM. (1983). Canine mammary epithelial neoplasms: biologic implications of morphologic characteristics assessed in 232 dogs. Vet Pathol 20:127-142.
- Griffey SM, Madewell BR, Dairkee SH, Hunt JE, Naydan DK, Higgins RJ. (1993). Immunohistochemical reactivity of basal and luminal epithelium-specific cytokeratin antibodies within normal and neoplastic canine mammary glands. Vet Pathol 30:155-161.
- Hellmén E, Bergstrom R, Holmberg L, Spangberg I-B, Hansson K, Lindgren A. (1993). Prognostic factors in canine mammary tumors: a multivariate study of 202 consecutive cases. Vet Pathol 30:20-27.
- Howard EM, Lau SK, Lyles RH, Birdsong GG, Umbreit JN, Kochhar R. (2005). Expression of e-cadherin in high-risk breast cancer. J Cancer Res Clin Oncol 131:14-18.
- Hsu W-L, Huang H-M, Liao J-W, Wong M-L, Chang S-C. (2007). Increased survival in dogs with malignant mammary tumours overexpressing HER-2 protein and detection of a silent single nucleotide polymorphism in the canine HER-2 gene. Vet J Doi:10.1016/j.tvjl.2007.10.013
- Itoh T, Uchida K, Ishikawa K, Kushima K, Kushima E, Tama H, Moritake T, Nakao H, Shii H. (2005). Clinicopathological survey of 101 canine mammary gland tumors: differences between small-breed dogs and others. J Vet Med Sci 67:345-347.

- Jorgensen JT, Nielsen KV, Ejlertsen B. (2007). Pharmacodiagnostics and targeted therapies- a rational approach for individualizing medical anticancer therapy in breast cancer. The Oncologist 12:397-405.
- Kersting C, Tidow N, Schmidt H, Liedtke C, Neumann J, Boecker W, van Diest PJ, Brandt B, Buerger H. (2004). Gene dosage PCR and fluorescence in situ hybridization reveal low frequency of EGFR amplifications despite protein overexpression in invasive breast carcinoma. Lab Invest 84:582-587.
- Klijn JG, Berns PM, Schmitz PI, Foekens JA. (1992). The clinical significance of epidermal growth factor receptor (EGF-R) in human breast cancer: a review on 5232 patients. Endocr Rev 13:3–17.
- Knudsen KA, Wheelock MJ. (2005). Cadherins and the mammary gland. J Cell Biochem 95:488-496.
- La Rosa S, Rigoli E, Uccella S, Novario R, Capella C. (2007). Prognostic and biological significance of cytokeratin 19 in pancreatic endocrine tumours. Histopathology 50:597-606.
- Lambros MB, Natrajan R, Reis-Filho JS. (2007). Chromogenic and fluorescent in situ hybridization in breast cancer. Hum Pathol 38:1105–1122.
- Livasy CA, Karaca G, Nanda R, Tretiakova MS, Olopade OI, Moore DT, Perou CM. (2006). Phenotypic evaluation of the basal-like subtype of invasive breast carcinoma. Mod Pathol 19:264–271.
- Mahler-Araujo B, Savage K, Parry S, Reis-Filho JS. (2008). Reduction of E-cadherin expression is associated with non-lobular breast carcinomas of basal-like and triple negative phenotype. J Clin Pathol. 61:615-20.
- Matos AJF, Lopes C, Carvalheira J, Santos M, Rutteman GR, Gartner F. (2006). E-cadherin expression in canine malignant mammary tumours: relationship to other clinicopathological variables. J Comp Pathol 134:182-189.
- Matos I, Dufloth R, Alvarenga M, Zeferino LC, Schmitt F. (2005). p63, cytokeratin 5, and P-cadherin: three molecular markers to distinguish basal phenotype in breast carcinomas. Virchows Arch 447:688-694.
- Milanezi F, Carvalho S, Schmitt FC. (2008). EGFR/HER2 in breast cancer: a biological approach for molecular diagnosis and therapy. Expert Rev Mol Diagn 8:417-434.
- Nerurkar VR, Seshadri R, Mulherkar R, Ishwad CS, Lalitha VS, Naik SN. (1987). Receptors for epidermal growth factor and estradiol in canine mammary tumors. Int J Cancer 40:230-232.
- Nielsen TO, Hsu FD, Jensen K Cheang M, Karaca G, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslen LA, Ragaz J, Gown AM, Gilks CB, van de Rijn M, Perou

- CM. (2004). Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. Clin Cancer Res 10:5367–5374.
- Nieto A, Peña L, Perez-Alenza MD, Sanchez MA, Flores JM, Castaño M. (2000). Immunohistologic detection of estrogen receptor alpha in canine mammary tumors: clinical and pathologic associations and prognostic significance. Vet Pathol 37:239–247.
- Paredes J, Albergaria A, Oliveira J, Jerónimo C, Milanezi F, Schmitt F. (2005). P-cadherin overexpression is an indicator of clinical outcome in invasive breast carcinomas and is associated with CDH3 promoter hypomethylation. Clin Cancer Res 11:5869-5877.
- Paredes J, Correia AL; Ribeiro AS, Albergaria A, Milanezi F, Schmitt FC. (2007). P-cadherin expression in breast cancer: a review. Breast Cancer Res 9:214.
- Paredes J, Stove C, Stove V, Milanezi F, van Marck V, Derycke L, Mareel M, Bracke M, Schmitt F. (2004). P-cadherin is up-regulated by the antiestrogen ICI 182,780 and promotes invasion of human breast cancer cells. Cancer Res 64:8309-8317.
- Parikh RR, Yang Q, Higgins SA, Haffty BG. (2008). Outcomes in young women with breast cancer of triple-negative phenotype: the prognostic significance of CK19 expression. Int J Radiat Oncol Biol Phys 70:35-42.
- Park K, Han S, Shin E, Kim HJ, Kim JY. (2007). EGFR gene and protein expression in breast cancers. Eur J Surg Oncol 33:956-960.
- Park SH, Cheung LW, Wong AS, Leung PC. (2008). Estrogen regulates Snail and Slug in the downregulation of E-cadherin and induces metastatic potential of ovarian cancer cells through estrogen receptor alpha. Mol Endocrinol doi:10.1210/me.2007-0512.
- Payne SJL, Bowen RL, Jones JL, Wells CA. (2008). Predictive markers in breast cancer the present. Histopathology 52:82–90.
- Peña L, Nieto A, Pérez-Alenza D, Cuesta P, Castano M. (1998). Immunohistochemical detection of Ki-67 and PCNA in canine mammary tumors: relationship to clinical and pathologic variables. J Vet Diagn Invest 10:237-246.
- Peralta Soler A, Knudsen KA, Salazar H, Han AC, Keshgegian AA. (1999). P-cadherin expression in breast carcinoma indicates poor survival. Cancer 86:1263-1272.
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lonning PE, Borresen-Dale AL, Brown PO, Botstein D. (2000). Molecular portraits of human breast tumours. Nature 406:747-752.
- Rakha EA, Putti TC, Abd El-Rehim DM, Paish C, Green AR, Powe DG, Lee AH, Robertson JF, Ellis IO. (2006). Morphological and immunophenotypic analysis of breast carcinomas with basal and myoepithelial differentiation. J Pathol 208:495–506.

- Rakha EA, El-Sayed ME, Reis-Filho J, Ellis IO. (2008). Expression profiling technology: its contribution to our understanding of breast cancer. Histopathology 52:67-81.
- Rakha EA, El-Sayed ME, Reis-Filho J, Ellis IO. (2008). Patho-biological aspects of basal-like breast cancer. Breast Cancer Res Treat Doi 10.1007/s10549-008-9952-1.
- Rao NAS, van Wolferen ME, van den Ham R, van Leenen D, Groot Koerkamp MJA, Holstege FCP, Mol JA. (2008). cDNA microarray profiles of canine mammary tumour cell lines reveal deregulated pathways pertaining to their phenotype. Animal Genetics Doi:10.1111/j.1365-2052.2008.01733.x.
- Reis Filho JS, Milanezi F, Carvalho S, Simpson PT, Steele D, Savage K, Lambros MB, Pereira EM, Nesland JM, Lakhani SR, Schmitt FC. (2005). Metaplastic breast carcinomas exhibit EGFR, but not HER2, gene amplification and overexpression: immunohistochemical and chromogenic in situ hybridization analysis. Breast Cancer Res 7:RA1028-1035.
- Rutteman GR, Foekens JA, Blankenstein MA, Vos JH, Misdorp W. (1990). EGF-receptors in non-affected and tumorous dog mammary tissues. Eur J Cancer 26:182-186.
- Rutteman GR, Foekens JA, Portengen H, Vos JH, Blankenstein MA, Teske E, Cornelisse CJ, Misdorp W. (1994). Expression of epidermal growth factor receptor (EGFR) in non-affected and tumorous mammary tissue of female dogs. Breast Cancer Res Treat 30:139-146.
- Rutteman GR, Withrow SJ, MacEwen EG. (2001). Tumors of the mammary gland. In: SJ Withrow and BR MacEwen (Ed.) Small Animal Clinical Oncology, WB Saunders Company, Philadelphia, pp. 455–477.
- Santini D, Ceccarelli C, Taffurelli M, Pileri S, Marrano D. (1996). Differentiation pathways in primary invasive breast carcinoma as suggested by intermediate filament and biopathological marker expression. J Pathol 179:386-391.
- Schaller G, Fuchs I, Pritze W, Ebert A, Herbst H, Pantel K, Weitzel H, Lengyel E. (1996). Elevated keratin 18 protein expression indicates a favorable prognosis in patients with breast cancer. Clin Cancer Res 2:1879–1885.
- Shiozaki H, Oka H, Inoue M, Tamura S, Monden M. (1996). E-cadherin mediated adhesion system in cancer cells. Cancer 77:1605-1613.
- Siziopikou KP, Cobleigh M. (2007). The basal subtype of breast carcinomas may represent the group of breast tumors that could benefit from EGFR targeted therapies. Breast 16:104–107.
- Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. (1987). Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. Science 235:177–182.
- Sorenmo K. (2003). Canine mammary gland tumors. Vet Clin North Am 33: 573-596.

- Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, Rijn MV, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Lonning PE, Borresen-Dale AL. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98:10869–10874.
- Sorlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lonning PE, Brown PO, Borresen-Dale AL, Botstein D. (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 100:8418–8423.
- Sotiriou C, Neo SY, McShane LM, Korn EL, Long PM, Jazaeri A, Martiat P, Fox SB, Harris AL, Liu ET. (2003). Breast cancer classification and prognosis based on gene expression profiles from a population-based study. Proc Natl Acad Sci. USA 100:10393–10398.
- Takei H, Iino Y, Horiguchi J, Kanoh T, Takao Y, Oyama T, Morishita Y. (1995). Immunohistochemical analysis of cytokeratin #8 as a prognostic factor in invasive breast carcinoma. Anticancer Res 15:1101-1105.
- Tsuda H, Takarabe T, Hasegawa F, Fukutomi T, Hirohashi S. (2000). Large, central acellular zones indicating myoepithelial tumor differentiation in high-grade invasive ductal carcinomas as markers of predisposition to lung and brain metastases. Am J Surg Pathol 24:197–202.
- Vogelstein B, Kinzler KW. (2004). Cancer genes and the pathways they control. Nat Med 10:789–799.
- Widakowich C, Azambuja E, Gil T, Cardoso F, Dihn P, Awada A, Piccart-Gebhart M. (2007). Molecular targeted therapies in breast cancer: where are we now? Int J Biochem Cell Biol 39:1375-1387.
- Woelfle U, Sauter G, Santjer S, Brakenhoff R, Pantel K. (2004). Downregulated expression of cytokeratin 18 promotes progression of human breast cancer. Clin Cancer Res 10:2670–2674.
- Yehiely F, Moyano JV, Evans JR, Nielsen TO, Cryns VL. (2006). Deconstructing the molecular portrait of basal-like breast cancer. Trends Mol Med 12:537-544.
- Zaidan Dagli ML. (2008). The search for suitable prognostic markers for canine mammary tumors: A promising outlook. Vet J 177:3-5.